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## PRIORITY DOCUMENT

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### Novel cell cycle genes and uses thereof

The present invention relates to DNA sequences encoding cell cycle interacting proteins as well as to methods for obtaining the same. The present invention also provides vectors comprising said DNA sequences, wherein the DNA sequences are operatively linked to regulatory elements allowing expression in prokaryotic and/or eukaryotic host cells. In addition, the present invention relates to the proteins encoded by said DNA sequences, antibodies to said proteins and methods for their production. Furthermore, the present invention relates to regulatory sequences which naturally regulate the expression of the above described DNA sequences. The present invention also relates to a method for controlling or altering growth characteristics of a plant and/or a plant cell comprising introduction and/or expression of one or more cell cycle regulatory proteins functional in a plant or parts thereof and/or one or more DNA sequences encoding such proteins. Also provided by the present invention is a process for disruption plant cell division by interfering in the expression of a substrate for cyclin-dependent protein kinase using a DNA sequence according to the invention wherein said plant cell is part of a transgenic plant. The present invention further relates to diagnostic compositions comprising the aforementioned DNA sequences, proteins and antibodies. The present invention also relates to methods for the identification of compounds being capable of activating or inhibiting the cell cycle. Furthermore, the present invention relates to transgenic plant cells, plant tissue and plants containing the above-described DNA sequences, regulatory sequences and vectors as well as to the use of the aforementioned DNA sequences, regulatory sequences, vectors, proteins, antibodies and/or compounds identified by the method of the invention in plant cell and tissue culture, plant breeding and/or agriculture.

Several documents are cited throughout the text of this specification. Each of the documents cited herein (including any manufacturer's specifications, instructions, etc.) are hereby incorporated herein by reference; however, there is no admission that any document cited is indeed prior art as to the present invention.

Cell division is fundamental for growth in humans, animals and plants. Prior to dividing in two daughter cells, the mother cell needs to replicate its DNA. The cell cycle is traditionally divided into 4 distinct phases:

G1: the gap between mitosis and the onset of DNA synthesis;

S : the phase of DNA synthesis;

G2: the gap between S and mitosis;

M : mitosis, the process of nuclear division leading up to the actual cell division.

The distinction of these 4 phases provides a convenient way of dividing the interval between successive divisions. Although they have served a useful purpose, a recent flurry of experimental results, much of it as a consequence of cancer research, has resulted in a more intricate picture of the cell cycle's "four seasons" (Nasmyth, Science 274, 1643-1645, 1996; Nurse, Nature, 344, 503-508, 1990). The underlying mechanism controlling the cell cycle control system has only recently been studied in greater detail. In all eukaryotic systems, including plants, this control mechanism is based on two key families of proteins which regulate the essential process of cell division, namely protein kinases (cyclin dependent kinases or CDKs) and their activating associated subunits, called cyclins. The activity of these protein complexes is switched on and off at specific points of the cell cycle. Particular CDK-cyclin complexes activated at the G1/S transition trigger the start of DNA replication. Different CDK-cyclin complexes are activated at the G2/M transition and induce mitosis leading to cell division. Each of the CDK-cyclin complexes execute their regulatory role via modulating different sets of multiple target proteins. Furthermore, the large variety of developmental and environmental signals affecting cell division all converge on the regulation of CDK activity. CDKs can therefore be seen as the central engine driving cell division.

In animal systems and in yeast, knowledge about cell cycle regulations is now quite advanced. The activity of CDK-cyclin complexes is regulated at five levels: (i)

transcription of the CDK and cyclin genes; (ii) association of specific CDK's with their specific cyclin partner; (iii) phosphorylation/dephosphorylation of the CDK and cyclins; (iv) interaction with other regulatory proteins such as SUC1/CKS1 homologues and cell cycle kinase inhibitors (CKI); and (v) cell cycle phase-dependent destruction of the cyclins and CKIs.

The study of cell cycle regulation in plants has lagged behind that in animals and yeast. Some basic mechanisms of cell cycle control appear to be conserved among eukaryotes, including plants. Plants were shown to also possess CDK's, cyclins and CKI's. However plants have unique developmental features which are reflected in specific characteristics of the cell cycle control. These include for instance the absence of cell migration, the formation of organs throughout the entire lifespan from specialized regions called *meristems*, the formation of a cell wall and the capacity of non-dividing cells to re-enter the cell cycle. Another specific feature is that many plant cells, in particular those involved in storage (e.g. endosperm), are polyploid due to rounds of DNA synthesis without mitosis. This so-called endoreduplication is intimately related with cell cycle control.

Due to these fundamental differences, multiple components of the cell cycle of plants are unique compared to their yeast and animal counterparts. For example, plants contain a unique class of CDKs, such as CDC2b in *Arabidopsis*, which are both structurally and functionally different from animal and yeast CDKs.

The further elucidation of cell cycle regulation in plants and its differences and similarities with other eukaryotic systems is a major research challenge. Strictly for the case of comparison, some key elements about yeast and animal systems are described below in more detail.

As already mentioned above, the control of cell cycle progression in eukaryotes is mainly exerted at two transition points: one in late G<sub>1</sub>, before DNA synthesis, and one at the G<sub>2</sub>/M boundary. Progression through these control points is mediated by cyclin-dependent protein kinase (CDK) complexes, which contain, in more detail, a catalytic subunit of approximately 34-kDa encoded by the *CDK* genes. Both *Saccharomyces cerevisiae* and *Schizosaccharomyces pombe* only utilize one *CDK* gene for the regulation of their cell cycle. The kinase activity of their gene products p34<sup>CDC2</sup> and

p34<sup>CDC28</sup> in *Sch. pombe* and in *S. cerevisiae*, respectively, is dependent on regulatory proteins, called cyclins. Progression through the different cell cycle phases is achieved by the sequential association of p34<sup>CDC2/CDC28</sup> with different cyclins. Although in high eukaryotes this regulation mechanism is conserved, the situation is more complex since they have evolved to use multiple CDKs to regulate the different stages of the cell cycle. In mammals, seven CDKs have been described, defined as CDK1 to CDK7, each binding a specific subset of cyclins.

In animal systems, CDK activity is not only regulated by its association with cyclins but also involves both stimulatory and inhibitory phosphorylations. Kinase activity is positively regulated by phosphorylation of a Thr residue located between amino acids 160-170 (depending on the CDK protein). This phosphorylation is mediated by the CDK-activating kinase (CAK) which interestingly is a CDK/cyclin complex itself. Inhibitory phosphorylations occur at the ATP-binding site (the Tyr15 residue together with Thr14 in higher eukaryotes) and are carried out by at least two protein kinases. A specific phosphatase, CDC25, dephosphorylates these residues at the G<sub>2</sub>/M checkpoint, thus activating CDK activity and resulting in the onset of mitosis. CDK activity is furthermore negatively regulated by a family of mainly low-molecular weight proteins, called cyclin-dependent kinase inhibitors (CKIs). Kinase activity is inhibited by the tight association of these CKIs with the CDK/cyclin complexes.

With respect to cell cycle regulation in plants a summary of the state of the art is given below. In *Arabidopsis*, thusfar only two CDK genes have been isolated, *CDC2aAt* and *CDC2bAt*, of which the gene products share 56% amino acid identity. Both CDKs are distinguished by several features. First, only *CDC2aAt* is able to complement yeast p34<sup>CDC2/CDC28</sup> mutants. Second, *CDC2aAt* and *CDC2bAt* bear different cyclin-binding motifs (PSTAIRE and PPTALRE, respectively), suggesting they may bind distinct types of cyclins. Third, although both *CDC2aAt* and *CDC2bAt* show the same spatial expression pattern, they exhibit a different cell cycle phase-specific regulation. The *CDC2aAt* gene is expressed constitutively throughout the whole cell cycle. In contrast, *CDC2bAt* mRNA levels oscillate, being most abundant during the S and G<sub>2</sub> phases. In addition, multiple cyclins have been isolated from *Arabidopsis*. The majority displays the strongest sequence similarity with the animal A- or B-type class of cyclins, but also D-



type cyclins have been identified. Although the classification of *Arabidopsis* cyclins is mainly based upon sequence similarity, limited data suggests that this organization corresponds with differential functions of each cyclin class. Direct binding of any cyclin with an *Arabidopsis* CDK subunit has, however, not yet been demonstrated.

In order to manage problems related to plant growth, plant architecture and/or plant diseases, it is believed to be of utmost importance to identify and isolate plant genes and gene products involved in the regulation of the plant cell division, and more particularly coding for and interacting with CDK's and/or their interacting proteins, responsible for the control of the cell cycle and the completion of the S and M phase of the cell cycle. If such novel genes and/or proteins have been isolated and analyzed, the growth of the plant as a whole can be influenced. Also, the growth of specific tissues or organs and thus the architecture of the plant can be modified.

Thus, the technical problem underlying the present invention is to provide means and methods for modulating cell cycle proteins that are particular useful in agriculture and plant cell and tissue culture.

The solution to the technical problem is achieved by providing the embodiments characterized in the claims.

Accordingly, the invention relates to a DNA sequence encoding a cell cycle interacting protein or encoding an immunologically active and/or functional fragment of such a protein, selected from the group consisting of:

(a) DNA sequences

- (aa) comprising a nucleotide sequence encoding at least the mature form of a protein (LDV115) comprising the amino acid sequence as given in SEQ ID NO: 2;
- (ab) comprising the nucleotide sequence as given in SEQ ID NO: 1;
- (ac) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (aa) or (ab) under stringent hybridization conditions;

- (ad) comprising a nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (aa) or (ab);
- (ae) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (aa) to (ad);
- (b) DNA sequences
  - (ba) comprising a nucleotide sequence encoding at least the mature form of a protein (LDV24) comprising the amino acid sequence as given in SEQ ID NO: 4;
  - (bb) comprising the nucleotide sequence as given in SEQ ID NO: 3;
  - (bc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (ba) or (bb) under stringent hybridization conditions;
  - (bd) comprising a nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (ba) or (bb);
  - (be) comprising a nucleotide sequence encoding at least the cyclin-like interacting domain of the protein encoded by the nucleotide sequence of any one of (ba) to (bd);
- (c) DNA sequences
  - (ca) comprising a nucleotide sequence encoding at least the mature form of a protein (VB33) comprising the amino acid sequence as given in SEQ ID NO: 6;
  - (cb) comprising the nucleotide sequence as given in SEQ ID NO: 5;
  - (cc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (ca) or (cb) under stringent hybridization conditions;
  - (cd) comprising a nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (ca) or (cb);

- (ce) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (ca) to (cd);
- (d) DNA sequences
  - (da) comprising a nucleotide sequence encoding at least the mature form of a protein (VB89) comprising the amino acid sequence as given in SEQ ID NO: 8;
  - (db) comprising the nucleotide sequence as given in SEQ ID NO: 7;
  - (dc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (da) or (db) under stringent hybridization conditions;
  - (dd) comprising a nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (da) or (db);
  - (de) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (da) to (dd);
- (e) DNA sequences
  - (ea) comprising a nucleotide sequence encoding at least the mature form of a protein (VBSHF) comprising the amino acid sequence as given in SEQ ID NO: 10;
  - (eb) comprising the nucleotide sequence as given in SEQ ID NO: 9;
  - (ec) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (ea) or (eb) under stringent hybridization conditions;
  - (ed) comprising a nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (ea) or (eb);
  - (ee) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (ea) to (ed);

(f) DNA sequences

- (fa) comprising a nucleotide sequence encoding at least the mature form of a protein (VBDPBP) comprising the amino acid sequence as given in SEQ ID NO: 12;
- (fb) comprising the nucleotide sequence as given in SEQ ID NO: 11;
- (fc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (fa) or (fb) under stringent hybridization conditions;
- (fd) comprising an nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (fa) or (fb);
- (fe) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (fa) to (fd);

(g) DNA sequences

- (ga) comprising a nucleotide sequence encoding at least the mature form of a protein (VBDAHP) comprising the amino acid sequence as given in SEQ ID NO: 14;
- (gb) comprising the nucleotide sequence as given in SEQ ID NO: 13;
- (gc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (ga) or (gb) under stringent hybridization conditions;
- (gd) comprising an nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (ga) or (gb);
- (ge) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (ga) to (gd);

- (h) DNA sequences, the nucleotide sequence of which is degenerate as a result of the genetic code to a nucleotide sequence of a DNA sequence as defined in any one of (a) to (g); and

- (i) DNA sequences comprising a nucleotide sequence encoding a fragment of a protein encoded by a DNA sequence of any one of (a) to (h), wherein said fragment is capable of interacting with a cell cycle protein.

The term "cell cycle interacting protein" as denoted herein means a protein capable of binding to cyclin dependent kinases, in particular to CDC2a and/or CDC2b and preferably to plant cyclin dependent kinases or their subunits.

The term "cell cycle" means the cyclic biochemical and structural events associated with growth and proliferation of cells, and in particular with the regulation of the replication of DNA and mitosis. The cycle is divided into periods called: G<sub>0</sub>, Gap<sub>1</sub> (G<sub>1</sub>), DNA synthesis (S), Gap<sub>2</sub> (G<sub>2</sub>), and mitosis (M).

The term "proliferation" means growth and reproduction, i.e. division of cells.

The term "cell division" means mitosis, i.e. the usual process of cell reproduction.

The terms "gene", "polynucleotide", "nucleic acid sequence", "nucleotide sequence", "DNA sequence" or "nucleic acid molecule" as used herein refers to a polymeric form of nucleotides of any length, either ribonucleotides or deoxyribonucleotides. This term refers only to the primary structure of the molecule. Thus, this term includes double- and single-stranded DNA, and RNA. It also includes known types of modifications, for example, methylation, "caps" substitution of one or more of the naturally occurring nucleotides with an analog. Preferably, the DNA sequence of the invention comprises a coding sequence encoding at least the mature form of the above defined cell cycle interacting protein, i.e. the protein which is posttranslationally processed in its biologically active form, for example due to cleavage of leader or secretory sequences or a proprotein sequence or other natural proteolytic cleavage points.

By "functional fragment" and "biologically active form" polypeptides are meant that exhibit activity similar, but not necessarily identical, to an activity of the wild-type cell cycle interacting proteins of the invention or an activity that is enhanced over that of the wild-type proteins (either the full-length protein or, preferably, the mature protein), as measured in a particular biological assay. Assays of cell cycle interacting activity are disclosed, for example, in Example 1, below. These assays can be used to measure cell cycle interacting activity of partially purified or purified native or recombinant protein.

The cell cycle interacting protein of the invention binds to CDC2, i.e. CDC2a and/or CDC2b, e.g., from Arabidopsis. Thus, a polypeptide having a functional fragment or the "biological activity" of the cell cycle interacting protein of the invention will bind to CDCs as set forth in Example 1.

The term "immunologically active fragment" of a cell cycle interacting protein of the invention denotes proteins or peptides which have at least a part of the primary structural conformation for one or more epitopes capable of reacting specifically with antibodies to a protein which is encodable by a nucleic acid molecule as set forth above. Preferably, the peptides and proteins encoded by a nucleic acid molecule of the invention are recognized by an antibody that specifically recognizes an epitope of the cell cycle interacting protein comprising the amino acid residues that are unique for the protein encoded by any one of the aforementioned DNA sequences. Preferably, said peptides and proteins are capable of eliciting an effective immune response in a mammal, for example mouse or rabbit.

The DNA sequence which encodes for the predicted mature polypeptides of the proteins comprising SEQ ID NOS: 2, 4, 6, 8, 10, 12 or 14 or for the biologically active fragment thereof may include: only the coding sequence for the mature polypeptide or for a biologically active fragment thereof; the coding sequence for the mature polypeptide or for a biologically active fragment thereof and additional coding sequence such as a leader or secretory sequence or a proprotein sequence; the coding sequence for the mature polypeptide (and optionally additional coding sequence) and non-coding sequence, such as intron or non-coding sequence 5' and/or 3' of the coding sequence for the predicted mature polypeptide.

A "coding sequence" is a nucleotide sequence which is transcribed into mRNA and/or translated into a polypeptide when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a translation start codon at the 5'-terminus and a translation stop codon at the 3'-terminus. A coding sequence can include, but is not limited to mRNA, cDNA, recombinant nucleotide sequences or genomic DNA, while introns may be present as well under certain circumstances. Thus, the nucleotide sequences of the present invention can be engineered in order to alter a cell cycle interacting protein coding sequence for a variety

of reasons, including but not limited to, alterations which modify the cloning, processing and/or expression of the gene product. For example, mutations may be introduced using techniques which are well known in the art, e.g., site-directed mutagenesis to insert new restriction sites, to alter glycosylation patterns, to change codon preference, to produce splice variants, etc.

In accordance with the present invention a two-hybrid system (Fields et al., Nature 340 (1989), 245-246,) was exploited whereby CDC2aAt or CDC2bAt as bait and a cDNA library of a cell suspension as prey are used. Novel gene products interacting with CDC2aAt or CDC2bAt indicative of hitherto unknown plant cell cycle regulatory nucleotide sequences were identified. The library was made from a mixture mRNA from *Arabidopsis thaliana* cell suspensions harvested at various growing stages: early exponential, exponential, early stationary and stationary phase.

Nine cDNA clones have been identified in accordance with the invention comprising the nucleotide sequences as depicted in SEQ ID NOS: 1, 3, 5, 7, 9, 11 and 13, which encode proteins that are capable of specifically interacting with cdc2aAt or cdc2bAt; see Examples 1 and 2, below. The proteins encoded by the cDNA clones comprised the amino acid sequences depicted in SEQ ID NOS: 2, 4, 6, 8, 10, 12 and 14. Computer assisted homology search in genome data bases revealed that novel genes have been identified and/or genes where the (partial) cDNA was described but the particular function of the gene remained unknown. In particular, the examples of the present invention demonstrate that novel cell cycle interacting proteins and their encoding genes have been identified. The possible applications of the these cell cycle interacting proteins and their encoding nucleic acid molecules will be discussed further below and are evident from the description provided in Example 3.

The homology search was performed with the program BLASTX and BLASTN (version 2.0a19MP-WashU [build decunix3.2 01:53:29 05-feb-1998] (see Altschul, Nucleic Acids Res. 25 (1997), 3389-3402) on the *Arabidopsis thaliana* nucleic acids database at ATDB at Stanford (<http://genome-www2.stanford.edu/cgi-bin/AtDB/nph-blast2atdb>). The

function GAP (general alignment) (from the GCG 9.1 package, Genetics Computer Group Inc., Madison, USA) has been used with the parameters Gap weight = 12 and Length weight = 4 to quantify the percentage of homology and similarity. The protein sequences were then used to perform a BLASTP (version 2.0.4 [feb-24-1998]) with BEAUTY post-processing provided by the Human Genome Center, Baylor College of Medicine against the National Center for Biotechnology Information's non-redundant protein database (<http://dot.imgen.bcm.tmc.edu:9331/seq-search/protein-search.html>). The results of the homology search are described in the appended examples.

Thus it is evident that the genes comprising the nucleotide sequences of each SEQ ID NOS. 1, 3, 5, 7, 9, 11 and 13 each encode a member of a novel class of cell cycle interacting proteins.

The present invention also relates to DNA sequences hybridizing with the above-described DNA sequences and differ in one or more positions in comparison with these as long as they encode a cell cycle interacting protein. By "hybridizing" it is meant that such nucleic acid molecules hybridize under conventional hybridization conditions, preferably under stringent conditions such as described by, e.g., Sambrook (Molecular Cloning; A Laboratory Manual, 2nd Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1989)). An example of one such stringent hybridization condition is hybridization at 4XSSC at 65 °C, followed by a washing in 0.1XSSC at 65 °C for one hour. Alternatively, an exemplary stringent hybridization condition is in 50 % formamide, 4XSSC at 42 °C. Cell cycle interacting proteins derived from other organisms such as mammals, in particular humans, may be encoded by other DNA sequences which hybridize to the sequences for plant cell cycle interacting proteins under relaxed hybridization conditions and which code on expression for peptides having the ability to interact with cell cycle proteins. Examples of such non-stringent hybridization conditions are 4XSSC at 50 °C or hybridization with 30-40 % formamide at 42 °C. Such molecules comprise those which are fragments, analogues or derivatives of the cell cycle interacting protein of the invention and differ, for example, by way of amino acid and/or nucleotide deletion(s), insertion(s), substitution(s), addition(s) and/or recombination(s) or



any other modification(s) known in the art either alone or in combination from the above-described amino acid sequences or their underlying nucleotide sequence(s). Using the PESTFIND program (Rogers, Science 234 (1986), 364-368), PEST sequences (rich in proline, glutamic acid, serine, and threonine) can be identified, which are characteristically present in unstable proteins. Such sequences may be removed from the cell cycle interacting proteins in order to increase the stability and optionally the activity of the proteins. Methods for introducing such modifications in the nucleic acid molecules according to the invention are well-known to the person skilled in the art. The invention also relates to nucleic acid molecules the sequence of which differs from the nucleotide sequence of any of the above-described nucleic acid molecules due to the degeneracy of the genetic code. All such fragments, analogues and derivatives of the protein of the invention are included within the scope of the present invention, as long as the essential characteristic immunological and/or biological properties as defined above remain unaffected in kind, that is the novel nucleic acid molecules of the invention include all nucleotide sequences encoding proteins or peptides which have at least a part of the primary structural conformation for one or more epitopes capable of reacting with antibodies to cell cycle interacting proteins which are encodable by a nucleic acid molecule as set forth above and which have comparable or identical characteristics in terms of biological activity and/or the capability to interact with other proteins. It is preferred that proteins encoded by a nucleic acid molecule of the invention are at least capable of interacting with CDC2, particularly CDC2a and/or CDC2b, preferably from a plant such as *Arabidopsis thaliana*. Whilst the above described proteins may interact with a CDC2 with CDC2 from *Arabidopsis thaliana*, the most likely interaction is with a CDC2 from the same species from which the gene was isolated (homologous interaction). This capability allows advantageous uses of the proteins of the invention and their encoding nucleic acid molecules as will be described in more detail below. Part of the invention is therefore also nucleic acid molecules encoding a polypeptide comprising at least a functional part of a cell cycle interacting protein encoded by a nucleic acid sequence comprised in a nucleic acid molecule according to the invention. An example for this is that the polypeptide or a fragment thereof according to the invention is embedded in another amino acid sequence. Preferably, the DNA sequence

of the invention encodes a protein having substantially the same amino acid sequence as the proteins defined in SEQ ID NOS 2, 4, 6, 8, 10, 12 and 14.

### **Extending the polynucleotide sequence of the invention**

The polynucleotide sequences encoding the cell cycle interacting proteins may be extended utilizing partial nucleotide sequence and various methods known in the art to detect upstream sequences such as promoters and regulatory elements. Gobinda, (PCR Methods Applic. 2 (1993), 318-322) discloses "restriction-site" polymerase chain reaction (PCR) as a direct method which uses universal primers to retrieve unknown sequence adjacent to a known locus. First, genomic DNA is amplified in the presence of primer to a linker sequence and a primer specific to the known region. The amplified sequences are subjected to a second round of PCR with the same linker primer and another specific primer internal to the first one. Products of each round of PCR are transcribed with an appropriate RNA polymerase and sequenced using reverse transcriptase.

Inverse PCR can be used to amplify or extend sequences using divergent primers based on a known region (Triglia, Nucleic Acids Res. 16 (1988), 8186). The primers may be designed using OLIGO® 4.06 Primer Analysis Software (1992; National Biosciences Inc, Plymouth MN), or another appropriate program to be preferably 22-30 nucleotides in length, to have a GC content of preferably 50% or more, and to anneal to the target sequence at temperatures preferably about 68°-72°C. The method uses several restriction enzymes to generate a suitable fragment in the known region of a gene. The fragment is then circularized by intramolecular ligation and used as a PCR template.

Capture PCR (Lagerstrom, PCR Methods Applic. 1 (1991), 111-119) is a method for PCR amplification of DNA fragments adjacent to a known sequence in, e.g., human or plant yeast artificial chromosome DNA. Capture PCR also requires multiple restriction enzyme digestions and ligations to place an engineered double-stranded sequence into an unknown portion of the DNA molecule before PCR.

Another method which may be used to retrieve unknown sequences is that of Parker, (Nucleic Acids Res. 19 (1991), 3055-3060). Additionally, one can use PCR, nested primers and PromoterFinder libraries to walk in genomic DNA (PromoterFinder™ Clontech (Palo Alto CA). This process avoids the need to screen libraries and is useful in finding intron/exon junctions. Preferred libraries for screening for full length cDNAs are ones that have been size-selected to include larger cDNAs. Also, random primed libraries are preferred in that they will contain more sequences which contain the 5' and upstream regions of genes. A randomly primed library may be particularly useful if an oligo d(T) library does not yield a full-length cDNA. Genomic libraries are useful for extension into the 5' nontranslated regulatory region. Capillary electrophoresis may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products; see, e.g., Sambrook, supra. Systems for rapid sequencing are available from Perkin Elmer, Beckmann Instruments (Fullerton CA), and other companies.

#### **Computer-assisted identification of cell cycle interacting proteins and their encoding genes**

As is further described in the appended examples BLAST2, which stands for Basic Local Alignment Search Tool (Altschul, 1997; Altschul, J. Mol. Evol. 36 (1993), 290-300; Altschul, J. Mol. Biol. 215 (1990), 403-410), can be used to search for local sequence alignments. BLAST produces alignments of both nucleotide and amino acid sequences to determine sequence similarity. Because of the local nature of the alignments, BLAST is especially useful in determining exact matches or in identifying homologs. The fundamental unit of BLAST algorithm output is the High-scoring Segment Pair (HSP). An HSP consists of two sequence fragments of arbitrary but equal lengths whose alignment is locally maximal and for which the alignment score meets or exceeds a threshold or cutoff score set by the user. The BLAST approach is to look for HSPs between a query sequence and a database sequence, to evaluate the statistical significance of any matches found, and to report only those matches which satisfy the user-selected threshold of significance. The parameter E establishes the statistically significant threshold for reporting database sequence matches. E is interpreted as the

upper bound of the expected frequency of chance occurrence of an HSP (or set of HSPs) within the context of the entire database search. Any database sequence whose match satisfies E is reported in the program output.

Analogous computer techniques using BLAST (Altschul, 1997, 1993 and 1990, *supra*) are used to search for identical or related molecules in nucleotide databases such as GenBank or EMBL. This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or homologous. The basis of the search is the product score which is defined as:

$$\frac{\% \text{sequence identity} \times \% \text{maximum BLAST score}}{100}$$

and it takes into account both the degree of similarity between two sequences and the length of the sequence match. For example, with a product score of 40, the match will be exact within a 1-2% error; and at 70, the match will be exact. Homologous molecules are usually identified by selecting those which show product scores between 15 and 40, although lower scores may identify related molecules.

#### **Identifying derivatives, variants and homologs of the cell cycle interacting proteins of the invention**

As is demonstrated in the appended examples a two-hybrid screening assay has been developed in accordance with the present invention suitable for identifying cell cycle interacting proteins. Thus, in another aspect the present invention relates to a method for identifying and obtaining cell cycle interacting proteins comprising a two-hybrid screening assay wherein CDC2a or CDC2b as a bait and a cDNA library of cell suspension culture as prey are used. Preferably, said CDC2a and CDC2b is CDC2aAt and CDC2bAt, respectively. However, CDKs or their corresponding subunits from other plants or other organisms such as mammals may be employed as well. The cell culture may be from any organism possessing cell cycle interacting proteins such as animals, preferably mammals. Particularly preferred are plant cell suspension cultures such as from Arabidopsis. The nucleic acid molecules encoding proteins or peptides identified to

interact with CDC2a or CDC2b in the above mentioned assay can be easily obtained and sequenced by methods known in the art; see also the appended examples. Therefore, the present invention also relates to a DNA sequence encoding a cell cycle interacting protein obtainable by the method of the invention.

In a preferred embodiment the nucleic acid molecules according to the invention are RNA or DNA molecules, preferably cDNA, genomic DNA or synthetically synthesized DNA or RNA molecules. Since cell cycle interacting proteins are supposed to play a key role in plant cell division, corresponding proteins displaying similar properties should be present in other organisms including mammals as well. Nucleic acid molecules of the invention can be obtained, e.g., by hybridization of the above-described nucleic acid molecules with a (sample of) nucleic acid molecule(s) of any source. Nucleic acid molecules hybridizing with the above-described nucleic acid molecules can in general be derived from any organism, preferably plants possessing such molecules, preferably from monocotyledonous or dicotyledonous plants, in particular from plants of interest in agriculture, horticulture or wood culture, such as crop plants, namely those of the family Poaceae, any starch producing plants, such as potato, maniok, leguminous plants, oil producing plants, such as oilseed rape, linseed, etc., plants using polypeptide as storage substances, such as soybean, plants using sucrose as storage substance, such as sugar beet or sugar cane, trees, ornamental plants etc. Preferably, the nucleic acid molecules according to the invention are derived from crop plants (e.g. maize, rice, barley, wheat, rye, oats etc.), potatoes, oil producing plants (e.g. oilseed rape, sunflower, peanut, soybean, etc.), cotton, sugar beet, sugar cane, leguminous plants (e.g. beans, peas etc.), and, of course, from *Arabidopsis thaliana*. Nucleic acid molecules hybridizing to the above-described nucleic acid molecules can be isolated, e.g., from libraries, such as cDNA or genomic libraries by techniques well known in the art. For example, hybridizing nucleic acid molecules can be identified and isolated by using the above-described nucleic acid molecules or fragments thereof or complements thereof as probes to screen libraries by hybridizing with said molecules according to standard techniques. Possible is also the isolation of such nucleic acid molecules by applying a nucleic acid amplification technique such as the polymerase chain reaction

(PCR) using as primers oligonucleotides derived from the above-described nucleic acid molecules.

Nucleic acid molecules which hybridize with any of the aforementioned nucleic acid molecules also include fragments, derivatives and allelic variants of the above-described nucleic acid molecules that encode a cell cycle interacting protein or an immunologically active or functional fragment thereof. Fragments are understood to be parts of nucleic acid molecules long enough to encode the described protein or a functional or immunologically active fragment thereof as defined above.

The term "derivative" means in this context that the nucleotide sequence of these nucleic acid molecules differs from the sequences of the above-described nucleic acid molecules in one or more nucleotide positions and are highly homologous to said nucleic acid molecules. Homology is understood to refer to a sequence identity of at least 40 %, particularly an identity of at least 60 %, preferably more than 80 % and still more preferably more than 90 %. The term "substantially homologous" refers to a subject, for instance a nucleic acid, which is at least 50% identical in sequence to the reference when the entire ORF (open-reading frame) is compared, where the sequence identity is preferably at least 70%, more preferably at least 80%, still more preferably at least 85%, especially more than about 90%, most preferably 95% or greater, particularly 98% or greater. The deviations from the sequences of the nucleic acid molecules described above can, for example, be the result of nucleotide substitution(s), deletion(s), addition(s), insertion(s) and/or recombination(s); see supra.

Homology further means that the respective nucleic acid molecules or encoded proteins are functionally and/or structurally equivalent. The nucleic acid molecules that are homologous to the nucleic acid molecules described above and that are derivatives of said nucleic acid molecules are, for example, variations of said nucleic acid molecules which represent modifications having the same biological function, in particular encoding proteins with the same or substantially the same biological function. They may be naturally occurring variations, such as sequences from other plant varieties or species, or mutations. These mutations may occur naturally or may be obtained by

mutagenesis techniques. The allelic variations may be naturally occurring allelic variants as well as synthetically produced or genetically engineered variants; see supra.

The proteins encoded by the various derivatives and variants of the above-described nucleic acid molecules may share specific common characteristics, such as biological activity, molecular weight, immunological reactivity, conformation, etc., as well as physical properties, such as electrophoretic mobility, chromatographic behavior, sedimentation coefficients, pH optimum, temperature optimum, stability, solubility, spectroscopic properties, etc.

Examples of the different possible applications of the nucleic acid molecules according to the invention as well as molecules derived from them will be described in detail in the following.

#### **Uses of the nucleic acid molecules of the present invention**

In one embodiment, the present invention relates to a nucleic acid molecule which hybridizes with the complementary strand of the nucleic acid molecule of the invention and which encodes a mutated version of the protein as defined above which has lost its immunological and/or biological activity. This embodiment may prove useful for, e.g., generating dominant mutant alleles of the above-described cell cycle interacting proteins. Said mutated version is preferably generated by substitution, deletion and/or addition of 1 to 5 or 5 to 10 amino acid residues in the amino acid sequence of the above-described wild type proteins.

In a further embodiment, the invention relates to nucleic acid molecules of at least 15 nucleotides in length hybridizing specifically with a nucleic acid molecule as described above or with a complementary strand thereof. Specific hybridization occurs preferably under stringent conditions and implies no or very little cross-hybridization with nucleotide sequences encoding no or substantially different proteins. Such nucleic acid molecules may be used as probes and/or for the control of gene expression. Nucleic acid probe technology is well known to those skilled in the art who will readily appreciate

that such probes may vary in length. Preferred are nucleic acid probes of 16 to 35 nucleotides in length. Of course, it may also be appropriate to use nucleic acids of up to 100 and more nucleotides in length. The nucleic acid probes of the invention are useful for various applications. On the one hand, they may be used as primers for amplification of nucleic acid sequences according to the invention. The design and use of said primers is known by the person skilled in the art. Preferably such amplification primers comprise a contiguous sequence of at least 6 nucleotides, in particular 13 nucleotides, preferably 15 to 25 nucleotides or more, identical or complementary to the nucleotide sequence depicted in SEQ ID NOS: 1, 3, 5, 7, 9, 11 or 13 or to a nucleotide sequence encoding the amino acid sequence of SEQ ID NOS: 2, 4, 6, 8, 10, 12 or 14. Another application is the use as a hybridization probe to identify nucleic acid molecules hybridizing with a nucleic acid molecule of the invention by homology screening of genomic DNA or cDNA libraries. Nucleic acid molecules according to this preferred embodiment of the invention which are complementary to a nucleic acid molecule as described above may also be used for repression of expression of a cell cycle gene, for example due to an antisense or triple helix effect or for the construction of appropriate ribozymes (see, e.g., EP-A1 0 291 533, EP-A1 0 321 201, EP-A2 0 360 257) which specifically cleave the (pre)-mRNA of a gene comprising a nucleic acid molecule of the invention or part thereof. Selection of appropriate target sites and corresponding ribozymes can be done as described, for example, in Steinecke, Ribozymes, Methods in Cell Biology 50, Galbraith et al. eds Academic Press, Inc. (1995), 449-460. The above described nucleic acid molecules may either be DNA or RNA or a hybrid thereof. Furthermore, said nucleic acid molecule may contain, for example, thioester bonds and/or nucleotide analogues, commonly used in oligonucleotide anti-sense approaches. Said modifications may be useful for the stabilization of the nucleic acid molecule against endo- and/or exonucleases in the cell. Said nucleic acid molecules may be transcribed by an appropriate vector containing a chimeric gene which allows for the transcription of said nucleic acid molecule in the cell.

Furthermore, the person skilled in the art is well aware that it is also possible to label such a nucleic acid probe with an appropriate marker for specific applications, such as for the detection of the presence of a nucleic acid molecule of the invention in a sample



derived from an organism, in particular plants. A number of companies such as Pharmacia Biotech (Piscataway NJ), Promega (Madison WI), and US Biochemical Corp (Cleveland OH) supply commercial kits and protocols for these procedures. Suitable reporter molecules or labels include those radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents as well as substrates, cofactors, inhibitors, magnetic particles and the like. Patents teaching the use of such labels include US Patents US-A-3,817,837; US-A-3,850,752; US-A-3,939,350; US-A-3,996,345; US-A-4,227,437; US-A-4,275,149 and 4,366,241. Also, recombinant immunoglobulins may be produced as shown in US-A-4,816,567 incorporated herein by reference.

Furthermore, the so-called "peptide nucleic acid" (PNA) technique can be used for the detection or inhibition of the expression of a nucleic acid molecule of the invention. For example, the binding of PNAs to complementary as well as various single stranded RNA and DNA nucleic acid molecules can be systematically investigated using thermal denaturation and BIAcore surface-interaction techniques (Jensen, Biochemistry 36 (1997), 5072-5077). Furthermore, the nucleic acid molecules described above as well as PNAs derived therefrom can be used for detecting point mutations by hybridization with nucleic acids obtained from a sample with an affinity sensor, such as BIAcore; see Gotoh, Rinsho Byori 45 (1997), 224-228. Hybridization based DNA screening on peptide nucleic acids (PNA) oligomer arrays are described in the prior art, for example in Weiler, Nucleic Acids Research 25 (1997), 2792-2799. The synthesis of PNAs can be performed according to methods known in the art, for example, as described in Koch, J. Pept. Res. 49 (1997), 80-88; Finn, Nucleic Acids Research 24 (1996), 3357-3363. Further possible applications of such PNAs, for example as restriction enzymes or as templates for the synthesis of nucleic acid oligonucleotides are known to the person skilled in the art and are, for example, described in Veselkov, Nature 379 (1996), 214 and Bohler, Nature 376 (1995), 578-581.

#### **Detection and mapping of related polynucleotide sequences**

The nucleic acid sequence for a cell cycle interacting protein of the invention can also be used to generate hybridization probes for mapping the naturally occurring genomic

sequence. The sequence may be mapped to a particular chromosome or to a specific region of the chromosome using well known techniques. These include *in situ* hybridization to chromosomal spreads, flow-sorted chromosomal preparations, or artificial chromosome constructions such as yeast artificial chromosomes, bacterial artificial chromosomes, bacterial P1 constructions or single chromosome cDNA libraries as reviewed in Price (Blood Rev. 7 (1993), 127-134) and Trask (Trends Genet. 7 (1991), 149-154). The technique of fluorescent *in situ* hybridization of chromosome spreads has been described, among other places, in Verma, (1988) Human Chromosomes: A Manual of Basic Techniques, Pergamon Press, New York NY. Fluorescent *in situ* hybridization of chromosomal preparations and other physical chromosome mapping techniques may be correlated with additional genetic map data. Examples of genetic map data can be found in the 1994 Genome Issue of Science (265:1981f) and Meinke, Science 282 (1998), 662-682. Correlation between the location of the gene encoding a cell cycle interacting protein of the invention on a physical chromosomal map and a specific feature, e.g., plant growth, architecture, yield, disease etc. may help delimit the region of DNA associated with this feature. The nucleotide sequences of the subject invention may be used to detect differences in gene sequences between normal, carrier or affected individuals. Furthermore, the means and methods described herein can be used for marker-assisted breeding.

*In situ* hybridization of chromosomal preparations and physical mapping techniques such as linkage analysis using established chromosomal markers may be used for extending genetic maps. For example an sequence tagged site based map of the human genome was recently published by the Whitehead-MIT Center for Genomic Research (Hudson, Science 270 (1995), 1945-1954) on a map of the plant genome by way of the Arabidopsis genome is available from <http://genome.wwz.Stanford.edu/cgi-bin/AtDB/nph-blast2atdb>. Often the placement of a gene on the chromosome of another species may reveal associated marker even if the number or arm of a particular chromosome is not known. New sequences can be assigned to chromosomal arms, or parts thereof, by physical mapping. This provides valuable information to investigators searching for interacting genes using positional cloning or other gene discovery

techniques. Once such gene has been crudely localized by genetic linkage to a particular genomic region, any sequences mapping to that area may represent associated or regulatory genes for further investigation. The nucleotide sequence of the subject invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc. among normal, carrier or affected individuals.

### **Vectors and expression systems**

The present invention also relates to vectors, particularly plasmids, cosmids, viruses, bacteriophages and other vectors used conventionally in genetic engineering that contain a nucleic acid molecule according to the invention. Methods which are well known to those skilled in the art can be used to construct various plasmids and vectors; see, for example, the techniques described in Sambrook, *Molecular Cloning A Laboratory Manual*, Cold Spring Harbor Laboratory (1989) N.Y. and Ausubel, *Current Protocols in Molecular Biology*, Green Publishing Associates and Wiley Interscience, N.Y. (1989), (1994). Plasmids and vectors to be preferably employed in accordance with the present invention include those well known in the art. Alternatively, the nucleic acid molecules and vectors of the invention can be reconstituted into liposomes for delivery to target cells.

In a preferred embodiment the nucleic acid molecule present in the vector is linked to (a) control sequence(s) which allow the expression of the nucleic acid molecule in prokaryotic and/or eukaryotic cells.

The term "control sequence" refers to regulatory DNA sequences which are necessary to effect the expression of coding sequences to which they are ligated. The nature of such control sequences differs depending upon the host organism. In prokaryotes, control sequences generally include promoter, ribosomal binding site, and terminators. In eukaryotes generally control sequences include promoters, terminators and, in some instances, enhancers, transactivators or transcription factors. The term "control sequence" is intended to include, at a minimum, all components the presence of which

are necessary for expression, and may also include additional advantageous components.

The term "operably linked" refers to a juxtaposition wherein the components so described are in a relationship permitting them to function in their intended manner. A control sequence "operably linked" to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under conditions compatible with the control sequences. In case the control sequence is a promoter, it is obvious for a skilled person that double-stranded nucleic acid is preferably used.

Thus, the vector of the invention is preferably an expression vector. An "expression vector" is a construct that can be used to transform a selected host cell and provides for expression of a coding sequence in the selected host. Expression vectors can for instance be cloning vectors, binary vectors or integrating vectors. Expression comprises transcription of the nucleic acid molecule preferably into a translatable mRNA. Regulatory elements ensuring expression in prokaryotic and/or eukaryotic cells are well known to those skilled in the art. In the case of eukaryotic cells they comprise normally promoters ensuring initiation of transcription and optionally poly-A signals ensuring termination of transcription and stabilization of the transcript, for example in plants, those of the 35S RNA from Cauliflower Mosaic Virus (CaMV). Other promoters commonly used are the polyubiquitin promoter, and the actin promoter for ubiquitous expression. The termination signals usually employed are from the Nopaline Synthase promoter or from the CaMV 35S promoter. A plant translational enhancer often used is the TMV omega sequences, the inclusion of an intron (Intron-1 from the Shrunken gene of maize, for example) has been shown to increase expression levels by up to 100-fold. (Mait, Transgenic Research 6 (1997), 143-156; Ni, Plant Journal 7 (1995), 661-676). Additional regulatory elements may include transcriptional as well as translational enhancers. Possible regulatory elements permitting expression in prokaryotic host cells comprise, e.g., the  $P_L$ , *lac*, *trp* or *tac* promoter in *E. coli*, and examples of regulatory elements permitting expression in eukaryotic host cells are the AOX1 or GAL1 promoter in yeast or the CMV-, SV40-, RSV-promoter (Rous sarcoma virus), CMV-enhancer, SV40-enhancer or a globin intron in mammalian and other animal cells. In this context, suitable expression vectors are known in the art such as Okayama-Berg cDNA

expression vector pcDV1 (Pharmacia), pCDM8, pRc/CMV, pcDNA1, pcDNA3 (Invitrogen), pSPORT1 (GIBCO BRL). An alternative expression system which could be used to express a cell cycle interacting protein is an insect system. In one such system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in *Trichoplusia* larvae. The coding sequence of a nucleic acid molecule of the invention may be cloned into a nonessential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of said coding sequence will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein coat. The recombinant viruses are then used to infect *S. frugiperda* cells or *Trichoplusia* larvae in which the protein of the invention is expressed (Smith, J. Virol. 46 (1983), 584; Engelhard, Proc. Nat. Acad. Sci. USA 91 (1994), 3224-3227).

Advantageously, the above-described vectors of the invention comprises a selectable and/or scorable marker. Selectable marker genes useful for the selection of transformed plant cells, callus, plant tissue and plants are well known to those skilled in the art and comprise, for example, antimetabolite resistance as the basis of selection for dhfr, which confers resistance to methotrexate (Reiss, Plant Physiol. (Life Sci. Adv.) 13 (1994), 143-149); npt, which confers resistance to the aminoglycosides neomycin, kanamycin and paromycin (Herrera-Estrella, EMBO J. 2 (1983), 987-995) and hygromycin (Marsh, Gene 32 (1984), 481-485). Additional selectable genes have been described, namely trpB, which allows cells to utilize indole in place of tryptophan; hisD, which allows cells to utilize histinol in place of histidine (Hartman, Proc. Natl. Acad. Sci. USA 85 (1988), 8047); mannose-6-phosphate isomerase which allows cells to utilize mannose (WO 94/20627) and ODC (ornithine decarboxylase) which confers resistance to the ornithine decarboxylase inhibitor, 2-(difluoromethyl)-DL-ornithine, DFMO (McConlogue, 1987, In: Current Communications in Molecular Biology, Cold Spring Harbor Laboratory ed.) or deaminase from *Aspergillus terreus* which confers resistance to Blasticidin S (Tamura, Biosci. Biotechnol. Biochem. 59 (1995), 2336-2338).

Useful scorable marker are also known to those skilled in the art and are commercially available. Advantageously, said marker is a gene encoding luciferase (Giacomin, Pl.

Sci. 116 (1996), 59-72; Scikantha, J. Bact. 178 (1996), 121), green fluorescent protein (Gerdes, FEBS Lett. 389 (1996), 44-47) or  $\beta$ -glucuronidase (Jefferson, EMBO J. 6 (1987), 3901-3907). This embodiment is particularly useful for simple and rapid screening of cells, tissues and organisms containing a vector of the invention.

The present invention furthermore relates to host cells comprising a vector as described above or a nucleic acid molecule according to the invention wherein the nucleic acid molecule is foreign to the host cell.

By "foreign" it is meant that the nucleic acid molecule is either heterologous with respect to the host cell, this means derived from a cell or organism with a different genomic background, or is homologous with respect to the host cell but located in a different genomic environment than the naturally occurring counterpart of said nucleic acid molecule. This means that, if the nucleic acid molecule is homologous with respect to the host cell, it is not located in its natural location in the genome of said host cell, in particular it is surrounded by different genes. In this case the nucleic acid molecule may be either under the control of its own promoter or under the control of a heterologous promoter. The vector or nucleic acid molecule according to the invention which is present in the host cell may either be integrated into the genome of the host cell or it may be maintained in some form extrachromosomally. In this respect, it is also to be understood that the nucleic acid molecule of the invention can be used to restore or create a mutant gene via homologous recombination (Paszkowski (ed.), Homologous Recombination and Gene Silencing in Plants. Kluwer Academic Publishers (1994)).

The host cell can be any prokaryotic or eukaryotic cell, such as bacterial, insect, fungal, plant or animal cells. Preferred fungal cells are, for example, those of the genus *Saccharomyces*, in particular those of the species *S. cerevisiae*.

The term "prokaryotic" is meant to include all bacteria which can be transformed or transfected with a DNA or RNA molecules for the expression of a protein of the invention. Prokaryotic hosts may include gram negative as well as gram positive bacteria such as, for example, *E. coli*, *S. typhimurium*, *Serratia marcescens* and *Bacillus subtilis*. The term "eukaryotic" is meant to include yeast, higher plant, insect and preferably mammalian cells. Depending upon the host employed in a recombinant

production procedure, the protein encoded by the polynucleotide of the present invention may be glycosylated or may be non-glycosylated. The cell cycle interacting proteins of the invention may or may not also include an initial methionine amino acid residue. A polynucleotide of the invention can be used to transform or transfect the host using any of the techniques commonly known to those of ordinary skill in the art. Furthermore, methods for preparing fused, operably linked genes and expressing them in, e.g., mammalian cells and bacteria are well-known in the art (Sambrook, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989).

Another subject of the invention is a method for the preparation of cell cycle interacting proteins which comprises the cultivation of host cells according to the invention which, due to the presence of a vector or a nucleic acid molecule according to the invention, are able to express such a protein, under conditions which allow expression of the protein and recovering of the so-produced protein from the culture. It is also to be understood that the proteins can be expressed in a cell free system using for example in vitro translation assays known in the art.

The term "expression" means the production of a protein or nucleotide sequence in the cell. However, said term also includes expression of the protein in a cell-free system. It includes transcription into an RNA product, post-transcriptional modification and/or translation to a protein product or polypeptide from a DNA encoding that product, as well as possible post-translational modifications. Depending on the specific constructs and conditions used, the protein may be recovered from the cells, from the culture medium or from both. The terms "protein" and "polypeptide" used in this application are interchangeable. "Polypeptide" refers to a polymer of amino acids (amino acid sequence) and does not refer to a specific length of the molecule. Thus peptides and oligopeptides are included within the definition of polypeptide. This term does also refer to or include post-translational modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations and the like. Included within the definition are, for example, polypeptides containing one or more analogs of an amino acid (including, for example, unnatural amino acids, etc.), polypeptides with substituted linkages, as well as other modifications known in the art, both naturally occurring and

non-naturally occurring. For example, it is well known by the person skilled in the art that it is not only possible to express a native protein but also to express the protein as fusion polypeptides or to add signal sequences directing the protein to specific compartments of the host cell, e.g., ensuring secretion of the protein into the culture medium, etc. The protein of the invention may also be expressed as a recombinant protein with one or more additional polypeptide domains added to facilitate protein purification. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp, Seattle WA). The inclusion of a cleavable linker sequences such as Factor XA or enterokinase (Invitrogen, San Diego CA) between the purification domain and the protein of interest is useful to facilitate purification. One such expression vector provides for expression of a fusion protein comprising a cell cycle interacting protein and contains nucleic acid encoding 6 histidine residues followed by thioredoxin and an enterokinase cleavage site. The histidine residues facilitate purification on IMIAC (immobilized metal ion affinity chromatography as described in Porath, Protein Expression and Purification 3 (1992), 263-281) while the enterokinase cleavage site provides a means for purifying the cell cycle interacting protein from the fusion protein. In addition to recombinant production, fragments of the protein of the invention may be produced by direct peptide synthesis using solid-phase techniques (cf Stewart et al (1969) Solid Phase Peptide Synthesis, WH Freeman Co, San Francisco; Merrifield, J. Am. Chem. Soc. 85 (1963), 2149-2154). *In vitro* protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer, Foster City CA) in accordance with the instructions provided by the manufacturer. Various fragments of the cell cycle interacting protein of the invention may be chemically synthesized and/or modified separately and combined using chemical methods to produce the full length molecule. Once expressed or synthesized, the protein of the present invention can be purified according to standard procedures of the art, including ammonium sulfate precipitation, affinity columns, column chromatography, gel



electrophoresis and the like; see, Scopes, "Protein Purification", Springer-Verlag, N.Y. (1982). Substantially pure proteins of at least about 90 to 95% homogeneity are preferred, and 98 to 99% or more homogeneity are most preferred, for pharmaceutical uses. Once purified, partially or to homogeneity as desired, the proteins may then be used therapeutically (including extracorporeally) or in developing and performing assay procedures.

### **Cell cycle interacting proteins of the invention**

The present invention furthermore relates to cell cycle interacting proteins encoded by the nucleic acid molecules according to the invention or produced or obtained by the above-described methods, and to functional and/or immunologically active fragments of such cell cycle interacting proteins. The proteins and polypeptides of the present invention are not necessarily translated from a designated nucleic acid sequence; the polypeptides may be generated in any manner, including for example, chemical synthesis, or expression of a recombinant expression system, or isolation from a suitable viral system. The polypeptides may include one or more analogs of amino acids, phosphorylated amino acids or unnatural amino acids. Methods of inserting analogs of amino acids into a sequence are known in the art. The polypeptides may also include one or more labels, which are known to those skilled in the art. In this context, it is also understood that the proteins according to the invention may be further modified by conventional methods known in the art. By providing the proteins according to the present invention it is also possible to determine fragments which retain biological activity. This allows the construction of chimeric proteins and peptides comprising an amino sequence derived from the protein of the invention, which is crucial for its, e.g., binding activity and other functional amino acid sequences, e.g. GUS marker gene (Jefferson, EMBO J. 6 (1987), 3901-3907). The other functional amino acid sequences may be either physically linked by, e.g., chemical means to the proteins of the invention or may be fused by recombinant DNA techniques well known in the art.

The term "fragment of a sequence" or "part of a sequence" means a truncated sequence of the original sequence referred to. The truncated sequence (nucleic acid or

protein sequence) can vary widely in length; the minimum size being a sequence of sufficient size to provide a sequence with at least a comparable function and/or activity of the original sequence referred to, while the maximum size is not critical. In some applications, the maximum size usually is not substantially greater than that required to provide the desired activity and/or function(s) of the original sequence. Typically, the truncated amino acid sequence will range from about 5 to about 60 amino acids in length. More typically, however, the sequence will be a maximum of about 50 amino acids in length, preferably a maximum of about 30 amino acids. It is usually desirable to select sequences of at least about 10, 12 or 15 amino acids, up to a maximum of about 20 or 25 amino acids.

Furthermore, folding simulations and computer redesign of structural motifs of the protein of the invention can be performed using appropriate computer programs (Olszewski, *Proteins* 25 (1996), 286-299; Hoffman, *Comput. Appl. Biosci.* 11 (1995), 675-679). Computer modeling of protein folding can be used for the conformational and energetic analysis of detailed peptide and protein models (Monge, *J. Mol. Biol.* 247 (1995), 995-1012; Renouf, *Adv. Exp. Med. Biol.* 376 (1995), 37-45). In particular, the appropriate programs can be used for the identification of interactive sites of the cell cycle interacting protein and its receptor, its ligand or other interacting proteins by computer assisted searches for complementary peptide sequences (Fassina, *Immunomethods* 5 (1994), 114-120. Further appropriate computer systems for the design of protein and peptides are described in the prior art, for example in Berry, *Biochem. Soc. Trans.* 22 (1994), 1033-1036; Wodak, *Ann. N. Y. Acad. Sci.* 501 (1987), 1-13; Pabo, *Biochemistry* 25 (1986), 5987-5991. The results obtained from the above-described computer analysis can be used for, e.g., the preparation of peptide mimetics of the protein of the invention or fragments thereof. Such pseudopeptide analogues of the natural amino acid sequence of the protein may very efficiently mimic the parent protein (Benkirane, *J. Biol. Chem.* 271 (1996), 33218-33224). For example, incorporation of easily available achiral (-amino acid residues into a protein of the invention or a fragment thereof results in the substitution of amide bonds by polymethylene units of an aliphatic chain, thereby providing a convenient strategy for constructing a peptid mimetic (Banerjee, *Biopolymers* 39 (1996), 769-777). Superactive peptidomimetic analogues of small peptide hormones in other systems are

described in the prior art (Zhang, Biochem. Biophys. Res. Commun. 224 (1996), 327-331). Appropriate peptide mimetics of the protein of the present invention can also be identified by the synthesis of peptide mimetic combinatorial libraries through successive amide alkylation and testing the resulting compounds, e.g., for their binding and immunological properties. Methods for the generation and use of peptidomimetic combinatorial libraries are described in the prior art, for example in Ostresh, Methods in Enzymology 267 (1996), 220-234 and Dorner, Bioorg. Med. Chem. 4 (1996), 709-715.

Furthermore, a three-dimensional and/or crystallographic structure of the protein of the invention can be used for the design of peptide mimetic inhibitors of the biological activity of the protein of the invention (Rose, Biochemistry 35 (1996), 12933-12944; Rutenber, Bioorg. Med. Chem. 4 (1996), 1545-1558).

### **Antibodies**

Furthermore, the present invention relates to antibodies specifically recognizing a cell cycle interacting protein according to the invention or parts, i.e. specific fragments or epitopes, of such a protein. The antibodies of the invention can be used to identify and isolate other cell cycle interacting proteins and genes in any organism, preferably plants. These antibodies can be monoclonal antibodies, polyclonal antibodies or synthetic antibodies as well as fragments of antibodies, such as Fab, Fv or scFv fragments etc. Monoclonal antibodies can be prepared, for example, by the techniques as originally described in Köhler and Milstein, Nature 256 (1975), 495, and Galfré, Meth. Enzymol. 73 (1981), 3, which comprise the fusion of mouse myeloma cells to spleen cells derived from immunized mammals. Furthermore, antibodies or fragments thereof to the aforementioned peptides can be obtained by using methods which are described, e.g., in Harlow and Lane "Antibodies, A Laboratory Manual", CSH Press, Cold Spring Harbor, 1988. These antibodies can be used, for example, for the immunoprecipitation and immunolocalization of proteins according to the invention as well as for the monitoring of the synthesis of such proteins, for example, in recombinant organisms, and for the identification of compounds interacting with the protein according to the invention. For example, surface plasmon resonance as employed in the BIAcore system can be used to

increase the efficiency of phage antibodies selections, yielding a high increment of affinity from a single library of phage antibodies which bind to an epitope of the protein of the invention (Schier, Human Antibodies Hybridomas 7 (1996), 97-105; Malmberg, J. Immunol. Methods 183 (1995), 7-13). In many cases, the binding phenomena of antibodies to antigens is equivalent to other ligand/anti-ligand binding.

### **Transgenic plants**

Plant cell division can conceptually be influenced in three ways : (i) inhibiting or arresting cell division, (ii) maintaining, facilitating or stimulating cell division, (iii) uncoupling DNA synthesis from mitosis and cytokinesis or (iv) uncoupling cell division from intrinsic developmental or external environmental conditions. Modulation of the expression of a cell cycle interacting protein encoded by a nucleotide sequence according to the invention has surprisingly an advantageous influence on plant cell division characteristics, in particular on the disruption of the expression levels of genes involved in G1/S and/or G2/M transition and as a result thereof on the total make-up of the plant concerned or parts thereof. An example is that DNA synthesis, progression of DNA replication or mitosis will be negatively influenced by interfering with the formation of a cyclin-dependent protein kinase complex. Alternatively, overexpression of the cell cycle interacting protein accelerates reentry into the cell cycle.

The term "cyclin-dependent protein kinase complex" means the complex formed when a, preferably functional, cyclin associates with a, preferably, functional cyclin dependent kinase. Such complexes may be active in phosphorylating proteins and may or may not contain additional protein species.

The term "protein kinase" means an enzyme catalyzing the phosphorylation of proteins.

To analyse the industrial applicabilities of the invention, transformed plants can be made overproducing the nucleotide sequence according to the invention. Such an overexpression of the new gene(s), proteins or inactivated variants thereof will either positively or negatively have an effect on cell division. Methods to modify the expression levels and/or the activity are known to persons skilled in the art and include for instance

overexpression, co-suppression, the use of ribozymes, sense and anti-sense strategies, gene silencing approaches. "Sense strand" refers to the strand of a double-stranded DNA molecule that is homologous to a mRNA transcript thereof. The "anti-sense strand" contains an inverted sequence which is complementary to that of the "sense strand".

Hence, the nucleic acid molecules according to the invention are in particular useful for the genetic manipulation of plant cells in order to modify the characteristics of plants and to obtain plants with modified, preferably with improved or useful phenotypes. Similarly, the invention can also be used to modulate the cell division and the growth of cells, preferentially plant cells, in *in vitro* cultures. Specifically the plant cell division rate and/or the inhibition of a plant cell division can be influenced by overexpression or reducing the expression of a gene encoding a protein according to the invention. Overexpression of a cell cycle interacting protein encoding gene according to the invention promotes cell proliferation, while reducing gene expression arrests cell division or prevents reentry into the cell cycle. Part of the invention is thus the usage of the nucleic acid molecules as mentioned hereinbefore as a negative or positive regulator of cell proliferation. As a result of overproduction the G1/S generation time is shortened whereas the proliferation is less dependent on growth factors. A transformed plant can thus be obtained by transforming a plant cell with a gene encoding a polypeptide concerned or fragment thereof alone or in combination. For this purpose tissue specific promoters, in one construct or being present as a separate construct in addition to the sequence concerned, can be used. Alternatively the expression of the cell cycle interacting protein is inducible by cytokinines or sucrose. Surprisingly using a polypeptide or fragment thereof according to the invention or using antisense RNA for the gene according to the invention cell division of the meristems of the plant can be manipulated, positively and/or negatively respectively. Furthermore, overproduction of the cell cycle interacting protein of the invention enhances growth and results in cell division to be less sensitive to an arrest caused by environmental stress such as salt, drought, chilling and the like.

Thus, the present invention relates to a method for the production of transgenic plants, plant cells or plant tissue comprising the introduction of a nucleic acid molecule or vector of the invention into the genome of said plant, plant cell or plant tissue.

For the expression of the nucleic acid molecules according to the invention in sense or antisense orientation in plant cells, the molecules are placed under the control of regulatory elements which ensure the expression in plant cells. These regulatory elements may be heterologous or homologous with respect to the nucleic acid molecule to be expressed as well with respect to the plant species to be transformed. In general, such regulatory elements comprise a promoter active in plant cells. To obtain expression in all tissues of a transgenic plant, preferably constitutive promoters are used, such as the 35 S promoter of CaMV (Odell, Nature 313 (1985), 810-812) or promoters of the polyubiquitin genes of maize (Christensen, Plant Mol. Biol. 18 (1982), 675-689). In order to achieve expression in specific tissues of a transgenic plant it is possible to use tissue specific promoters (see, e.g., Stockhaus, EMBO J. 8 (1989), 2245-2251). Known are also promoters which are specifically active in tubers of potatoes or in seeds of different plants species, such as maize, Vicia, wheat, barley etc. Inducible promoters may be used in order to be able to exactly control expression. An example for inducible promoters are the promoters of genes encoding heat shock proteins. Also microspore-specific regulatory elements and their uses have been described (WO96/16182). Furthermore, the chemically inducible Tet-system may be employed (Gatz, Mol. Gen. Genet. 227 (1991); 229-237). Further suitable promoters are known to the person skilled in the art and are described, e.g., in Ward (Plant Mol. Biol. 22 (1993), 361-366). The regulatory elements may further comprise transcriptional and/or translational enhancers functional in plants cells. Furthermore, the regulatory elements may include transcription termination signals, such as a poly-A signal, which lead to the addition of a poly A tail to the transcript which may improve its stability.

In the case that a nucleic acid molecule according to the invention is expressed in sense orientation it is in principle possible to modify the coding sequence in such a way that the protein is located in any desired compartment of the plant cell. These include the nucleus,

endoplasmatic reticulum, the vacuole, the mitochondria, the plastids, the apoplast, the cytoplasm etc. Since the interacting component of the protein of the invention exerts its effects in the cytoplasm and/or nucleus, corresponding signal sequences are preferred to direct the protein of the invention in the same compartment. Methods how to carry out this modifications and signal sequences ensuring localization in a desired compartment are well known to the person skilled in the art.

Methods for the introduction of foreign DNA into plants are also well known in the art. These include, for example, the transformation of plant cells or tissues with T-DNA using *Agrobacterium tumefaciens* or *Agrobacterium rhizogenes*, the fusion of protoplasts, direct gene transfer (see, e.g., EP-A 164 575), injection, electroporation, biolistic methods like particle bombardment, pollen-mediated transformation, plant RNA virus-mediated transformation, liposome-mediated transformation, transformation using wounded or enzyme-degraded immature embryos, or wounded or enzyme-degraded embryogenic callus and other methods known in the art. The vectors used in the method of the invention may contain further functional elements, for example "left border"- and "right border"-sequences of the T-DNA of *Agrobacterium* which allow for stably integration into the plant genome. Furthermore, methods and vectors are known to the person skilled in the art which permit the generation of marker free transgenic plants, i.e. the selectable or scorable marker gene is lost at a certain stage of plant development or plant breeding. This can be achieved by, for example cotransformation (Lyznik, Plant Mol. Biol. 13 (1989), 151-161; Peng, Plant Mol. Biol. 27 (1995), 91-104) and/or by using systems which utilize enzymes capable of promoting homologous recombination in plants (see, e.g., WO97/08331; Bayley, Plant Mol. Biol. 18 (1992), 353-361); Lloyd, Mol. Gen. Genet. 242 (1994), 653-657; Maeser, Mol. Gen. Genet. 230 (1991), 170-176; Onouchi, Nucl. Acids Res. 19 (1991), 6373-6378). Methods for the preparation of appropriate vectors are described by, e.g., Sambrook (Molecular Cloning; A Laboratory Manual, 2nd Edition (1989), Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY).

Suitable strains of *Agrobacterium tumefaciens* and vectors as well as transformation of *Agrobacteria* and appropriate growth and selection media are well known to those

skilled in the art and are described in the prior art (GV3101 (pMK90RK), Koncz, Mol. Gen. Genet. 204 (1986), 383-396; C58C1 (pGV 3850kan), Deblaere, Nucl. Acid Res. 13 (1985), 4777; Bevan, Nucleic. Acid Res. 12(1984), 8711; Koncz, Proc. Natl. Acad. Sci. USA 86 (1989), 8467-8471; Koncz, Plant Mol. Biol. 20 (1992), 963-976; Koncz, Specialized vectors for gene tagging and expression studies. In: Plant Molecular Biology Manual Vol 2, Gelvin and Schilperoort (Eds.), Dordrecht, The Netherlands: Kluwer Academic Publ. (1994), 1-22; EP-A-120 516; Hoekema: The Binary Plant Vector System, Offsetdrukkerij Kanters B.V., Alblasserdam (1985), Chapter V, Fraley, Crit. Rev. Plant. Sci., 4, 1-46; An, EMBO J. 4 (1985), 277-287). Although the use of *Agrobacterium tumefaciens* is preferred in the method of the invention, other *Agrobacterium* strains, such as *Agrobacterium rhizogenes*, may be used, for example if a phenotype conferred by said strain is desired.

Methods for the transformation using biolistic methods are well known to the person skilled in the art; see, e.g., Wan, Plant Physiol. 104 (1994), 37-48; Vasil, Bio/Technology 11 (1993), 1553-1558 and Christou (1996) Trends in Plant Science 1, 423-431. Microinjection can be performed as described in Potrykus and Spangenberg (eds.), Gene Transfer To Plants. Springer Verlag, Berlin, NY (1995).

The transformation of most dicotyledonous plants is possible with the methods described above. But also for the transformation of monocotyledonous plants several successful transformation techniques have been developed. These include the transformation using biolistic methods as, e.g., described above as well as protoplast transformation, electroporation of partially permeabilized cells, introduction of DNA using glass fibers, etc.

The term "transformation" as used herein, refers to the transfer of an exogenous polynucleotide into a host cell, irrespective of the method used for the transfer. The polynucleotide may be transiently or stably introduced into the host cell and may be maintained non-integrated, for example, as a plasmid or as chimeric links, or alternatively, may be integrated into the host genome. The resulting transformed plant cell can then be used to regenerate a transformed plant in a manner known by a skilled person.



In general, the plants which can be modified according to the invention and which either show overexpression of a protein according to the invention or a reduction of the synthesis of such a protein can be derived from any desired plant species. They can be monocotyledonous plants or dicotyledonous plants, preferably they belong to plant species of interest in agriculture, wood culture or horticulture interest, such as crop plants (e.g. maize, rice, barley, wheat, rye, oats etc.), potatoes, oil producing plants (e.g. oilseed rape, sunflower, pea nut, soy bean, etc.), cotton, sugar beet, sugar cane, leguminous plants (e.g. beans, peas etc.), wood producing plants, preferably trees, etc.

Thus, the present invention relates also to a transgenic plant cell which contains (preferably stably integrated into its genome) a nucleic acid molecule according to the invention linked to regulatory elements which allow expression of the nucleic acid molecule in plant cells and wherein the nucleic acid molecule is foreign to the transgenic plant cell. For the meaning of foreign; see supra. The presence and expression of the nucleic acid molecule in the transgenic plant cells leads to the synthesis of a cell cycle interacting protein and leads to physiological and phenotypic changes in plants containing such cells.

Thus, the present invention also relates to transgenic plants and plant tissue comprising transgenic plant cells according to the invention. Due to the (over)expression of a cell cycle interacting protein of the invention, e.g., at developmental stages and/or in plant tissue in which they do not naturally occur these transgenic plants may show various physiological, developmental and/or morphological modifications in comparison to wild-type plants.

Therefore, part of this invention is the use of cell cycle genes and/or cell cycle interacting proteins to modulate plant cell division and/or growth in plant cells, plant tissues, plant organs and/or whole plants. To the scope of the invention also belongs a method to influence the activity of cyclin-dependent protein kinase in a plant cell by transforming the plant cell with a nucleic acid molecule according to the invention and/or manipulation of the expression of said molecule. More in particular using a nucleic acid molecule according to the invention, the disruption of plant cell division can be

accomplished by interfering in the expression of a substrate for cyclin-dependent protein kinase. The latter goal may be achieved, for example, with methods for reducing the amount of active cell cycle interacting proteins.

For example, to obtain transgenic plants overexpressing the *A. thaliana* LDV115 gene (see Example 3), its coding region can be cloned, e.g., into the pAT7002 vector (Aoyama and Chua, Plant J. 11 (1997), 605-612). This vector allows inducible expression of the cloned inserts by the addition of the glucocorticoid dexamethasone. For example, following a polymerase chain reaction (PCR) technology the coding region of LDV115 can be amplified using appropriate primers, whereby a first primer contains an XhoI and a second primer contains an SmaI restriction site. The obtained PCR fragment can be purified and cut with XhoI and SmaI. Subsequently the fragment can be cloned into the XhoI and SmaI sites of pTA7002. The resulted binary vector can be transferred into *Agrobacterium tumefaciens*. This strain can be used to transform *Nicotiana tabacum* cv. Petit havana using, e.g., the leaf disk protocol (Horsh, Science 227 (1985), 1229-1231) and *Arabidopsis thaliana* using, e.g., the root transformation protocol (Valvekens, PNAS 85 (1988), 5536-5540). Transgenic plants can then be selected on hygromycine 20 mg/l. Plants can be tested for LDV115 inducible expression as follows. 2 to 3 leaves of each transformant can be cut in two. Each half can be either submersed in 50 mM Na-citrate buffer (pH 5.8) with or without dexamethasone (0.03 mM concentration). After 24 hours of induction RNA can be extracted from these leaves using the Trizol reagents (Gibco-BRL) according to the manufactures and a northern gel can be run using, e.g., 5 µg of RNA. The gel can be blotted on a nitro-cellulose filter (HybondN+, Amersham) and hybridised with an LDV115 probe. Furthermore, seeds of transformants can be put on ½ MS medium with 1% sucrose, both with and without dexamethasone. As a control SR1 seeds should be included. In the presence of dexamethasone the growth behaviour of the transgenic plants as compared to the control plants is expected to be modified. For example, these transgenic plants may grow faster and/or have additional cells. Furthermore, said plant may be less sensitive to environmental stress compared to the corresponding wild type plant.

Furthermore, the invention also relates to a transgenic plant cell which contains (preferably stably integrated into its genome) a nucleic acid molecule according to the invention or part thereof, wherein the transcription and/or expression of the nucleic acid molecule or part thereof leads to reduction of the synthesis of a cell cycle interacting protein.

In a preferred embodiment, the reduction is achieved by an anti-sense, sense, ribozyme, co-suppression and/or dominant mutant effect.

"Antisense" and "antisense nucleotides" means DNA or RNA constructs which block the expression of the naturally occurring gene product.

The provision of the nucleic acid molecules according to the invention opens up the possibility to produce transgenic plant cells with a reduced level of the protein as described above and, thus, with a defect in cell division. Techniques how to achieve this are well known to the person skilled in the art. These include, for example, the expression of antisense-RNA, ribozymes, of molecules which combine antisense and ribozyme functions and/or of molecules which provide for a co-suppression effect; see also supra. When using the antisense approach for reduction of the amount of cell cycle interacting proteins in plant cells, the nucleic acid molecule encoding the antisense-RNA is preferably of homologous origin with respect to the plant species used for transformation. However, it is also possible to use nucleic acid molecules which display a high degree of homology to endogenously occurring nucleic acid molecules encoding a cell cycle interacting protein. In this case the homology is preferably higher than 80%, particularly higher than 90% and still more preferably higher than 95%. The reduction of the synthesis of a protein according to the invention in the transgenic plant cells can result in an alteration in, e.g., cell division. In transgenic plants comprising such cells this can lead to various physiological, developmental and/or morphological changes, preferably to improved regeneration and transformation capacity of, e.g., cultured cells or wounded tissue.

Thus, the present invention also relates to transgenic plants comprising the above-described transgenic plant cells. These may show, for example, a deficiency in cell

division and/or reduced growth characteristics compared to wild type plants due to the stable or transient presence of a foreign DNA resulting in at least one of the following features:

- (a) disruption of (an) endogenous gene(s) encoding a protein of the invention;
- (b) expression of at least one antisense RNA and/or ribozyme against a transcript comprising a nucleic acid molecule of the invention;
- (c) expression of a sense and/or non-translatable mRNA of the nucleic acid molecule of the invention;
- (d) expression of an antibody of the invention;
- (f) incorporation of a functional or non-functional copy of the regulatory sequence of the invention; or
- (g) incorporation of a recombinant DNA molecule or vector of the invention.

The present invention also relates to cultured plant tissues comprising transgenic plant cells as described above, which either show overexpression of a protein according to the invention or a reduction in synthesis of such a protein.

Any transformed plant obtained according to the invention can be used in a conventional breeding scheme or in *in vitro* plant propagation to produce more transformed plants with the same characteristics and/or can be used to introduce the same characteristic in other varieties of the same or related species. Such plants are also part of the invention. Seeds obtained from the transformed plants genetically also contain the same characteristic and are part of the invention. As mentioned before, the present invention is in principle applicable to any plant and crop that can be transformed with any of the transformation method known to those skilled in the art and includes for instance corn, wheat, barley, rice, oilseed crops, cotton, tree species, sugar beet, cassava, tomato, potato, numerous other vegetables, fruits.

In yet another aspect, the invention also relates to harvestable parts and to propagation material of the transgenic plants according to the invention which either contain transgenic plant cells expressing a nucleic acid molecule according to the invention or which contain

cells which show a reduced level of the described protein. Harvestable parts can be in principle any useful parts of a plant, for example, flowers, pollen, seedlings, tubers, leaves, stems, fruit, seeds, roots etc. Propagation material includes, for example, seeds, fruits, cuttings, seedlings, tubers, rootstocks etc.

### **Regulatory sequences of cell cycle interacting genes**

As mentioned above, the regulatory sequences that naturally drive the expression of the above described cell cycle interacting proteins may prove useful for the expression of heterologous DNA sequences in certain plant tissues and/or at different developmental stages in plant development.

Accordingly, in a further aspect the present invention relates to a regulatory sequence of a promoter naturally regulating the expression of a nucleic acid molecule of the invention described above or of a nucleic acid molecule homologous to a nucleic acid molecule of the invention. With methods well known in the art it is possible to isolate the regulatory sequences of the promoters that naturally regulate the expression of the above-described DNA sequences; see, e.g., Example 4. For example, using the above described nucleic acid molecules as probes a genomic library consisting of plant genomic DNA cloned into phage or bacterial vectors can be screened by a person skilled in the art. Such a library consists e.g. of genomic DNA prepared from seedlings, fractionized in fragments ranging from 5 kb to 50 kb, cloned into the lambda GEM11 (Promega) phages. Phages hybridizing with the probes can be purified. From the purified phages DNA can be extracted and sequenced. Having isolated the genomic sequences corresponding to the genes encoding the above-described cell cycle interacting proteins, it is possible to fuse heterologous DNA sequences to these promoters or their regulatory sequences via transcriptional or translational fusions well known to the person skilled in the art. In order to identify the regulatory sequences and specific elements of these cell cycle genes, 5'-upstream genomic fragments can be cloned in front of marker genes such as *luc*, *gfp* or the GUS coding region and the resulting chimeric genes can be introduced by means of *Agrobacterium tumefaciens*

mediated gene transfer into plants or transfected into plant cells or plant tissue for transient expression. The expression pattern observed in the transgenic plants or transfected plant cells containing the marker gene under the control of the regulatory sequences of the invention reveal the boundaries of the promoter and its regulatory sequences. Preferably, said regulatory sequence is capable of conferring expression of a heterologous DNA sequence in main and lateral root meristems, shoot apical meristems, embryos at the globular, heart and torpedo stages, floral meristems and/or cambial cells in the stem.

In context with the present invention, the term "regulatory sequence" refers to sequences which influence the specificity and/or level of expression, for example in the sense that they confer cell and/or tissue specificity; see supra. Such regions can be located upstream of the transcription initiation site, but can also be located downstream of it, e.g., in transcribed but not translated leader sequences.

The term "promoter", within the meaning of the present invention refers to nucleotide sequences necessary for transcription initiation, i.e. RNA polymerase binding, and may also include, for example, the TATA box.

The term "nucleic acid molecule homologous to a nucleic acid molecule of the invention", as used herein includes promoter regions and regulatory sequences of other cell cycle interacting protein encoding genes, such as genes from other species, for example, maize, alfalfa, potato, sorghum, millet, coix, barley, wheat and rice the coding region of which share substantial homology to the cell cycle interacting proteins of the invention and which display substantially the same expression pattern. Such promoters are characterized by their capability of conferring expression of a heterologous DNA sequence in meristematic tissue and cells and other tissues mentioned above.

Thus, according to the present invention, regulatory sequences from any species can be used that are functionally homologous to the regulatory sequences of the promoter of the above defined nucleic acid molecules, or promoters of genes that display an identical or similar pattern of expression, in the sense of being expressed in the above-mentioned tissues and cells. However, the expression conferred by the regulatory sequences of the invention may not be limited to, for example, root meristem cells but

can include or be restricted to, for example, subdomains of meristems. The particular expression pattern may also depend on the plant/vector system employed. However, expression of heterologous DNA sequences driven by the regulatory sequences of the invention predominantly occurs in the meristem unless certain elements of the regulatory sequences of the invention, were taken and designed by the person skilled in the art to control the expression of a heterologous DNA sequence in other cell types.

It is also immediately evident to the person skilled in the art that further regulatory elements may be added to the regulatory sequences of the invention. For example, transcriptional enhancers and/or sequences which allow for induced expression of the regulatory sequences of the invention may be employed. A suitable inducible system is for example tetracycline-regulated gene expression as described, e.g., by Gatz, *supra*.

The regulatory sequence of the invention may preferably be derived from the above described cell cycle interacting genes. Plants that may be suitable sources for such genes have been described above.

Usually, said regulatory sequence is part of a recombinant DNA molecule. In a preferred embodiment of the present invention, the regulatory sequence in the recombinant DNA molecule is operatively linked to a heterologous DNA sequence.

The term heterologous with respect to the DNA sequence being operatively linked to the regulatory sequence of the invention means that said DNA sequence is not naturally linked to the regulatory sequence of the invention. Expression of said heterologous DNA sequence comprises transcription of the DNA sequence, preferably into a translatable mRNA. Regulatory elements ensuring expression in eukaryotic cells, preferably plant cells, are well known to those skilled in the art. They usually comprise poly-A signals ensuring termination of transcription and stabilization of the transcript, see also *supra*. Additional regulatory elements may include transcriptional as well as translational enhancers; see *supra*.

In a preferred embodiment, the heterologous DNA sequence of the above-described recombinant DNA molecules encodes a peptide, protein, antisense RNA, sense RNA and/or ribozyme. The recombinant DNA molecule of the invention can be used alone or as part of a vector to express heterologous DNA sequences, which, e.g., encode proteins for, e.g., the control of disease resistance, modulation of nutrition value or diagnostics of cell cycle related gene expression. The recombinant DNA molecule or vector containing the DNA sequence encoding a protein of interest is introduced into the cells which in turn produce the RNA and optionally protein of interest. For example, the regulatory sequences of the invention can be operatively linked to a lethal gene for use in the production of male and female sterility in plants. Suitable lethal genes include the *Bacillus amyloliquefaciens* ribonuclease (Hartlet, J. Mol. Biol. 89 (1985)) and the *Bacillus amyloliquefaciens* ribonuclease expressed with or without its inhibitor, barstar. Another example for a lethal gene is the catalytic A fragment of diphtheria toxin (Tweeten, J. Bacteriol. 156 (1983), 680-685). Expression of diphtheria toxin within yeast cells causes ADP-ribosylation of elongation factor 2, which leads to inhibition of protein synthesis and eventual cell death (Mattheakis, Mol. Cell. Biol. 12 (1992), 4026-4037).

On the other hand, said protein can be a scorable marker, e.g., luciferase, green fluorescent protein or  $\beta$ -galactosidase. This embodiment is particularly useful for simple and rapid screening methods for compounds and substances described herein below capable of modulating cell cycle interacting protein gene expression. For example, a cell suspension can be cultured in the presence and absence of a candidate compound in order to determine whether the compound affects the expression of genes which are under the control of regulatory sequences of the invention, which can be measured, e.g., by monitoring the expression of the above-mentioned marker. It is also immediately evident to those skilled in the art that other marker genes may be employed as well, encoding, for example, a selectable marker which provides for the direct selection of compounds which induce or inhibit the expression of said marker.

The regulatory sequences of the invention may also be used in methods of antisense approaches. The antisense RNA may be a short (generally at least 10, preferably at



least 14 nucleotides, and optionally up to 100 or more nucleotides) nucleotide sequence formulated to be complementary to a portion of a specific mRNA sequence and/or DNA sequence of the gene of interest. Standard methods relating to antisense technology have been described; see, e.g., Klann, Plant Physiol. 112 (1996), 1321-1330. Following transcription of the DNA sequence into antisense RNA, the antisense RNA binds to its target sequence within a cell, thereby inhibiting translation of the mRNA and down-regulating expression of the protein encoded by the mRNA. Thus, in a further embodiment, the invention relates to nucleic acid molecules of at least 15 nucleotides in length hybridizing specifically with a regulatory sequence as described above or with a complementary strand thereof. For the possible applications of such nucleic acid molecules, see supra.

The present invention also relates to vectors, particularly plasmids, cosmids, viruses and bacteriophages used conventionally in genetic engineering that comprise a recombinant DNA molecule of the invention. Preferably, said vector is an expression vector and/or a vector further comprising a selection marker for plants. For example of suitable selector markers, see supra. Methods which are well known to those skilled in the art can be used to construct recombinant vectors; see, for example, the techniques described in Sambrook, Molecular Cloning A Laboratory Manual, Cold Spring Harbor Laboratory (1989) N.Y. and Ausubel, Current Protocols in Molecular Biology, Green Publishing Associates and Wiley Interscience, N.Y. (1989). Alternatively, the recombinant DNA molecules and vectors of the invention can be reconstituted into liposomes for delivery to target cells; see also supra.

The present invention furthermore relates to host cells transformed with a regulatory sequence, a DNA molecule or vector of the invention. Said host cell may be a prokaryotic or eukaryotic cell; see supra.

In a further preferred embodiment, the present invention provides for a method for the production of transgenic plants, plant cells or plant tissue comprising the introduction of a nucleic acid molecule, recombinant DNA molecule or vector of the invention into the

genome of said plant, plant cell or plant tissue. For the expression of the heterologous DNA sequence under the control of the regulatory sequence according to the invention in plant cells, further regulatory sequences such as poly A tail may be fused, preferably 3' to the heterologous DNA sequence, see also supra. Further possibilities might be to add Matrix Attachment Sites at the borders of the transgene to act as "delimiters" and insulate against methylation spread from nearby heterochromatic sequences. Methods for the introduction of foreign DNA into plants, plant cells and plant tissue are described above.

Thus, the present invention relates also to transgenic plant cells which contain stably integrated into the genome a recombinant DNA molecule or vector according to the invention.

Furthermore, the present invention also relates to transgenic plants and plant tissue comprising the above-described transgenic plant cells. These plants may show, for example, modified architecture, increased yield or an increased tolerance to diseases, e.g., nematodes, geminiviruses.

In yet another aspect the invention also relates to harvestable parts and to propagation material of the transgenic plants according to the invention which contain transgenic plant cells described above. Harvestable parts and propagation material can be in principle any useful part of a plant; see supra.

With the regulatory sequences of the invention, it will be possible to study *in vivo* gene expression related to cell cycle interacting proteins. Furthermore, since cell cycle interacting protein expression has different patterns in different stages of physiological and pathological conditions, it is now possible to determine further regulatory sequences which may be important for the up- or down-regulation of the expression or activity of cell cycle interacting proteins, for example in response to ions or elicitors. In addition, it is now possible to *in vivo* study mutations which affect different functional or regulatory aspects of specific gene expression in the cell cycle. Thus, the present invention also

relates to the use of the above described regulatory sequences and recombinant DNA molecules of the invention for the expression of heterologous DNA sequences.

The *in vivo* studies referred to above will be suitable to further broaden the knowledge on the mechanisms and genes involved in the control of the cell cycle. Expression of heterologous genes or antisense RNA under the control of the regulatory sequence of the present invention in plants and plant cells may allow the understanding of the function of each of these genes in the plant.

As mentioned hereinbefore, the nucleic acid molecules and proteins of the present invention provide a basis for the development of mimetic compounds that may be inhibitors or activators of cell cycle interacting proteins or their encoding genes. It will be appreciated that the present invention also provides cell based screening methods that allow a high-throughput-screening (HTS) of compounds that may be candidates for such inhibitors and activators.

Thus, the present invention further relates to a method for the identification of an activator or inhibitor of genes encoding cell cycle interacting proteins comprising the steps of:

- (a) culturing a plant cell or tissue or maintaining a plant comprising a recombinant DNA molecule comprising a readout system operatively linked to a regulatory sequence of the invention in the presence of a compound or a sample comprising a plurality of compounds under conditions which permit expression of said readout system;
- (b) identifying or verifying a sample and compound, respectively, which leads to suppression or activation and/or enhancement of expression of said readout system in said plant, plant cell, or plant tissue.

The present invention further relates to a method for identifying and obtaining an activator or inhibitor of cell cycle interacting proteins comprising the steps of:

- (a) combining a compound to be screened with a reaction mixture containing the cell cycle interacting protein of the invention and a readout system capable of interacting with the cell cycle interacting protein under suitable conditions which permit interaction of the cell cycle interacting protein with said readout system;
- (b) identifying or verifying a sample and compound, respectively, which leads to suppression or activation of the readout system.

The term "read out system" in context with the present invention means any substrate that can be monitored, for example due to enzymatically induced changes. It also includes DNA sequences which upon transcription and/or expression in a cell, tissue or organism provide for a scorable and/or selectable phenotype. Such read out systems are well known to those skilled in the art and comprise, for example, substrates for protein kinases, recombinant DNA molecules and marker genes as described above.

The term "plurality of compounds" in a method of the invention is to be understood as a plurality of substances which may or may not be identical.

Said compound or plurality of compounds may be chemically synthesized or microbiologically produced and/or comprised in, for example, samples, e.g., cell extracts from, e.g., plants, animals or microorganisms. Furthermore, said compound(s) may be known in the art but hitherto not known to be capable of suppressing or activating cell cycle interacting proteins. The reaction mixture may be a cell free extract or may comprise a cell or tissue culture. Suitable set ups for the method of the invention are known to the person skilled in the art and are, for example, generally described in Alberts et al., Molecular Biology of the Cell, third edition (1994), in particular Chapter 17. The plurality of compounds may be, e.g., added to the reaction mixture, culture medium, injected into the cell or sprayed onto the plant. The cell or tissue that may be employed in the method of the invention preferably is a host cell, plant cell or plant tissue of the invention described in the embodiments hereinbefore.

If a sample containing a compound or a plurality of compounds is identified in the method of the invention, then it is either possible to isolate the compound from the

original sample identified as containing the compound capable of suppressing or activating cell cycle interacting proteins, or one can further subdivide the original sample, for example, if it consists of a plurality of different compounds, so as to reduce the number of different substances per sample and repeat the method with the subdivisions of the original sample. Depending on the complexity of the samples, the steps described above can be performed several times, preferably until the sample identified according to the method of the invention only comprises a limited number of or only one substance(s). Preferably said sample comprises substances of similar chemical and/or physical properties, and most preferably said substances are identical. Several methods are known to the person skilled in the art for producing and screening large libraries to identify compounds having specific affinity for a target. These methods include the phage-display method in which randomized peptides are displayed from phage and screened by affinity chromatography to an immobilized receptor; see, e.g., WO 91/17271, WO 92/01047, US-A-5,223,409. In another approach, combinatorial libraries of polymers immobilized on a chip are synthesized using photolithography; see, e.g., US-A-5,143,854, WO 90/15070 and WO 92/10092. The immobilized polymers are contacted with a labeled receptor and scanned for label to identify polymers binding to the receptor. The synthesis and screening of peptide libraries on continuous cellulose membrane supports that can be used for identifying binding ligands of the polypeptide of the invention and thus possible inhibitors and activators is described, for example, in Kramer, *Methods Mol. Biol.* 87 (1998), 25-39. This method can also be used, for example, for determining the binding sites and the recognition motifs in the polypeptide of the invention. In like manner, the substrate specificity of the DnaK chaperon was determined and the contact sites between human interleukin-6 and its receptor; see Rudiger, *EMBO J.* 16 (1997), 1501-1507 and Weiergraber, *FEBS Lett.* 379 (1996), 122-126, respectively. Furthermore, the above-mentioned methods can be used for the construction of binding supertopes derived from the polypeptide of the invention. A similar approach was successfully described for peptide antigens of the anti-p24 (HIV-1) monoclonal antibody; see Kramer, *Cell* 91 (1997), 799-809. A general route to fingerprint analyses of peptide-antibody interactions using the clustered amino acid peptide library was described in Kramer, *Mol. Immunol.* 32 (1995), 459-465. In addition,

antagonists of the polypeptide of the invention can be derived and identified from monoclonal antibodies that specifically react with the polypeptide of the invention in accordance with the methods as described in Doring, Mol. Immunol. 31 (1994), 1059-1067.

More recently, WO 98/25146 described further methods for screening libraries of complexes for compounds having a desired property, especially, the capacity to agonize, bind to, or antagonize a polypeptide or its cellular receptor. The complexes in such libraries comprise a compound under test, a tag recording at least one step in synthesis of the compound, and a tether susceptible to modification by a reporter molecule. Modification of the tether is used to signify that a complex contains a compound having a desired property. The tag can be decoded to reveal at least one step in the synthesis of such a compound. Other methods for identifying compounds which interact with the proteins according to the invention or nucleic acid molecules encoding such molecules are, for example, the *in vitro* screening with the phage display system as well as filter binding assays or "real time" measuring of interaction using, for example, the BIAcore apparatus (Pharmacia).

All these methods can be used in accordance with the present invention to identify activators and antagonists of the polypeptide of the invention.

Various sources for the basic structure of such an activator or inhibitor can be employed and comprise, for example, mimetic analogs of the polypeptide of the invention. Mimetic analogs of the polypeptide of the invention or biologically active fragments thereof can be generated by, for example, substituting the amino acids that are expected to be essential for the biological activity with, e.g., stereoisomers, i.e. D-amino acids; see e.g., Tsukida, J. Med. Chem. 40 (1997), 3534-3541. Furthermore, in case fragments are used for the design of biologically active analogs Pro-mimetic components can be incorporated into a peptide to reestablish at least some of the conformational properties that may have been lost upon removal of part of the original polypeptide; see, e.g., Nachman, Regul. Pept. 57 (1995), 359-370. Furthermore, the polypeptide of the invention can be used to identify synthetic chemical peptide mimetics that bind to or can function as a ligand, substrate, binding partner or the receptor of the polypeptide of the

invention as effectively as does the natural polypeptide; see, e.g., Engleman, J. Clin. Invest. 99 (1997), 2284-2292.

The structure-based design and synthesis of low-molecular-weight synthetic molecules that mimic the activity of the native biological polypeptide is further described in, e.g., Dowd, Nature Biotechnol. 16 (1998), 190-195; Kieber-Emmons, Current Opinion Biotechnol. 8 (1997), 435-441; Moore, Proc. West Pharmacol. Soc. 40 (1997), 115-119; Mathews, Proc. West Pharmacol. Soc. 40 (1997), 121-125; Mukhija, European J. Biochem. 254 (1998), 433-438.

It is also well known to the person skilled in the art, that it is possible to design, synthesize and evaluate mimetics of small organic compounds that, for example, can act as a substrate or ligand to the polypeptide of the invention. For example, it has been described that D-glucose mimetics of hapalosin exhibited similar efficiency as hapalosin in antagonizing multidrug resistance assistance-associated protein in cytotoxicity; see Dinh, J. Med. Chem. 41 (1998), 981-987.

The nucleic acid molecule of the invention can also serve as a target for activators and inhibitors. Activators may comprise, for example, proteins that bind to the mRNA of a gene encoding a polypeptide of the invention, thereby stabilizing the native conformation of the mRNA and facilitating transcription and/or translation, e.g., in like manner as Tat protein acts on HIV-RNA. Furthermore, methods are described in the literature for identifying nucleic acid molecules such as an RNA fragment that mimics the structure of a defined or undefined target RNA molecule to which a compound binds inside of a cell resulting in retardation of cell growth or cell death; see, e.g., WO 98/18947 and references cited therein. These nucleic acid molecules can be used for identifying unknown compounds of pharmaceutical and/or agricultural interest, and for identifying unknown RNA targets for use in treating a disease. These methods and compositions can be used in screening for novel antibiotics, bacteriostatics, or modifications thereof or for identifying compounds useful to alter expression levels of proteins encoded by a nucleic acid molecule. Alternatively, for example, the

conformational structure of the RNA fragment which mimics the binding site can be employed in rational drug design to modify known antibiotics to make them bind more avidly to the target. One such methodology is nuclear magnetic resonance (NMR), which is useful to identify drug and RNA conformational structures. Still other methods are, for example, the drug design methods as described in WO 95/35367, US-A-5,322,933, where the crystal structure of the RNA fragment can be deduced and computer programs are utilized to design novel binding compounds which can act as antibiotics.

The compounds which can be tested and identified according to a method of the invention may be expression libraries, e.g., cDNA expression libraries, peptides, proteins, nucleic acids, antibodies, small organic compounds, hormones, peptidomimetics, PNAs or the like (Milner, *Nature Medicine* 1 (1995), 879-880; Hupp, *Cell* 83 (1995), 237-245; Gibbs, *Cell* 79 (1994), 193-198 and references cited supra). Furthermore, genes encoding a putative regulator of cell cycle interacting protein and/or which exert their effects up- or downstream the cell cycle interacting protein of the invention may be identified using, for example, insertion mutagenesis using, for example, gene targeting vectors known in the art (see, e.g., Hayashi, *Science* 258 (1992), 1350-1353; Fritze and Walden, *Gene activation by T-DNA tagging. In Methods in Molecular biology* 44 (Gartland, K.M.A. and Davey, M.R., eds). Totowa: Human Press (1995), 281-294) or transposon tagging (Chandlee, *Physiologia Plantarum* 78 (1990), 105-115). Said compounds can also be functional derivatives or analogues of known inhibitors or activators. Such useful compounds can be for example transacting factors which bind to the cell cycle interacting protein or regulatory sequences of the invention. Identification of transacting factors can be carried out using standard methods in the art (see, e.g., Sambrook, supra, and Ausubel, supra). To determine whether a protein binds to the protein or regulatory sequence of the invention, standard native gel-shift analyses can be carried out. In order to identify a transacting factor which binds to the protein or regulatory sequence of the invention, the protein or regulatory sequence of the invention can be used as an affinity reagent in standard protein purification methods, or as a probe for screening an expression library. The identification of nucleic acid molecules



which encode proteins which interact with the cell cycle interacting proteins described above can also be achieved, for example, as described in Scofield (Science 274 (1996), 2063-2065) by use of the so-called yeast "two-hybrid system"; see also the appended examples. In this system the protein encoded by the nucleic acid molecules according to the invention or a smaller part thereof is linked to the DNA-binding domain of the GAL4 transcription factor. A yeast strain expressing this fusion protein and comprising a lacZ reporter gene driven by an appropriate promoter, which is recognized by the GAL4 transcription factor, is transformed with a library of cDNAs which will express plant proteins or peptides thereof fused to an activation domain. Thus, if a peptide encoded by one of the cDNAs is able to interact with the fusion peptide comprising a peptide of a protein of the invention, the complex is able to direct expression of the reporter gene. In this way the nucleic acid molecules according to the invention and the encoded peptide can be used to identify peptides and proteins interacting with cell cycle interacting proteins. It is apparent to the person skilled in the art that this and similar systems may then further be exploited for the identification of inhibitors of the binding of the interacting proteins.

Once the transacting factor is identified, modulation of its binding to or regulation of expression of the cell cycle interacting protein of the invention can be pursued, beginning with, for example, screening for inhibitors against the binding of the transacting factor to the protein of the present invention. Activation or repression of cell cycle interacting proteins could then be achieved in plants by applying of the transacting factor (or its inhibitor) or the gene encoding it, e.g. in a vector for transgenic plants. In addition, if the active form of the transacting factor is a dimer, dominant-negative mutants of the transacting factor could be made in order to inhibit its activity. Furthermore, upon identification of the transacting factor, further components in the pathway leading to activation (e.g. signal transduction) or repression of a gene involved in the control of cell cycle then can be identified. Modulation of the activities of these components can then be pursued, in order to develop additional drugs and methods for modulating the cell cycle in animals and plants.

Thus, the present invention also relates to the use of the two-hybrid system as defined above for the identification of cell cycle interacting proteins or activators or inhibitors of such proteins

Determining whether a compound is capable of suppressing or activating cell cycle interacting proteins can be done, for example, by monitoring DNA duplication and cell division. It can further be done by monitoring the phenotypic characteristics of the cell of the invention contacted with the compounds and compare it to that of wild-type plants. In an additional embodiment, said characteristics may be compared to that of a cell contacted with a compound which is either known to be capable or incapable of suppressing or activating cell cycle interacting proteins.

The compounds isolated by the above methods also serve as lead compounds for the development of analog compounds. The analogs should have a stabilized electronic configuration and molecular conformation that allows key functional groups to be presented to the receptor in substantially the same way as the lead compound. In particular, the analog compounds have spatial electronic properties which are comparable to the binding region, but can be smaller molecules than the lead compound, frequently having a molecular weight below about 2 kD and preferably below about 1 kD. Identification of analog compounds can be performed through use of techniques such as self-consistent field (SCF) analysis, configuration interaction (CI) analysis, and normal mode dynamics analysis. Computer programs for implementing these techniques are available; e.g., Rein, Computer-Assisted Modeling of Receptor-Ligand Interactions (Alan Liss, New York, 1989). Methods for the preparation of chemical derivatives and analogues are well known to those skilled in the art and are described in, for example, Beilstein, Handbook of Organic Chemistry, Springer edition New York Inc., 175 Fifth Avenue, New York, N.Y. 10010 U.S.A. and Organic Synthesis, Wiley, New York, USA. Furthermore, said derivatives and analogues can be tested for their effects according to methods known in the art. Furthermore, peptidomimetics and/or computer aided design of appropriate derivatives and analogues can be used, for example, according to the methods described above.

The inhibitor or activator identified by the above-described method may prove useful as a herbicide, pesticide, insecticide, antibiotic, tumor suppressing agent and/or as a cell growth regulator. Thus, in a further embodiment the invention relates to a compound obtained or identified according to the method of the invention said compound being an activator of cell cycle interacting proteins or an inhibitor of cell cycle interacting proteins. The above-described compounds include, for example, cell cycle kinase inhibitors. "Cell-cycle kinase inhibitor" (CKI) is a protein which inhibit CDK/cyclin activity and is produced and/or activated when further cell division has to be temporarily or continuously prevented. The antibodies, nucleic acid molecules, inhibitors and activators of the present invention preferably have a specificity at least substantially identical to the binding specificity of the natural ligand or binding partner of the cell cycle protein of the invention, in particular if cell cycle stimulation is desired. An antibody or inhibitor can have a binding affinity to the cell cycle interacting protein of the invention of at least  $10^5\text{M}^{-1}$ , preferably higher than  $10^7\text{M}^{-1}$  and advantageously up to  $10^{10}\text{M}^{-1}$  in case cell cycle suppression should be mediated.

In a preferred embodiment, a suppressive antibody or inhibitor of the invention has an affinity of at least about  $10^{-7}$  M, preferably at least about  $10^{-9}$  M and most preferably at least about  $10^{-11}$  M; and cell cycle stimulating activator has an affinity of less than about  $10^{-7}$  M, preferably less than about  $10^{-6}$  M and most preferably in order of  $10^{-5}\text{M}$ .

In case of nucleic acid molecules it is preferred that they have a binding affinity to those encoding the amino acid sequences depicted in SEQ ID NO: 2, 4, 6, 8, 10, 12 or 14 of at most 2-, 5- or 10-fold less than an exact complement of 20 consecutive nucleotides of the above described nucleic acid molecules.

Preferably, the compound identified according to the above described method or its analog or derivative is further formulated in a therapeutically active form or in a form suitable for the application in plant breeding or plant cell and tissue culture. For example, it can be combined with a agriculturally acceptable carrier known in the art. Thus, the present invention also relates to a method of producing a therapeutic or plant effective composition comprising the steps of one of the above described methods of the invention and combining the compound obtained or identified in the method of the

invention or an analog or derivative thereof with a pharmaceutically acceptable carrier or with a plant cell and tissue culture acceptable carrier. As is evident from the above, the present invention generally relates to compositions comprising at least one of the aforementioned nucleic acid molecules, vectors, proteins, regulatory sequences, recombinant DNA molecules, antibodies or compounds. Advantageously, said composition is for use as a medicament, a diagnostic means, a kit or as a plant effective composition.

### **Compositions useful in agriculture and in plant cell and tissue culture**

Plant protection compositions can be prepared by employing the above-described methods of the invention and synthesizing the compound identified as inhibitor or activator in an amount sufficient for use in agriculture. Thus, the present invention also relates to a method for the preparation of an agricultural plant protection composition comprising the above-described steps of the method of the invention and synthesizing the compound so identified or an analog or derivative thereof.

In the plant protection composition of the invention, the compound identified by the above-described method may be preferentially formulated by conventional means commonly used for the application of, for example, herbicides and pesticides or agents capable of inducing systemic acquired resistance (SAR). For example, certain additives known to those skilled in the art stabilizers or substances which facilitate the uptake by the plant cell, plant tissue or plant may be used.

### **Pharmaceutical compositions**

The cell cycle interacting proteins of the invention appear to function in the cell division cycle which is similar in plants and animals. Accordingly, the nucleic acid molecules and proteins of the invention or derivatives thereof as well as the above described activators and inhibitors may be used to modulate the cell division cycle in animal, preferably mammalian cells which is integral to the development and spread of cancerous cells. A cell cycle interacting protein that acts as a basal transcription factor may promote

cancer cell growth. In conditions where cell cycle interacting protein activity is not desirable, cells could be transfected with antisense sequences to cell cycle interacting protein encoding polynucleotides or provided with antagonists to the protein or its encoding gene. Thus, the above described antagonists or antisense molecules may be used to slow, stop, or reverse cancer cell growth. Thus, the present invention also relates to a method of producing a therapeutic agent comprising the steps of the methods described hereinbefore and synthesizing the activator or inhibitor obtained or identified in step (c) or a. analog or derivative thereof in an amount sufficient to provide said agent in a therapeutically effective amount to a patient.

Compounds identified by the above methods or analogs are formulated for therapeutic use as pharmaceutical compositions. The compositions can also include, depending on the formulation desired, pharmaceutically acceptable, usually sterile, non-toxic carriers or diluents, which are defined as vehicles commonly used to formulate pharmaceutical compositions for animal or human administration. The diluent is selected so as not to affect the biological activity of the combination. Examples of such diluents are distilled water, physiological saline, Ringer's solutions, dextrose solution, and Hank's solution. In addition, the pharmaceutical composition or formulation may also include other carriers, adjuvants, or nontoxic, nontherapeutic, nonimmunogenic stabilizers and the like. A therapeutically effective dose refers to that amount of protein or its antibodies, antagonists, or inhibitors which ameliorate the symptoms or condition. Therapeutic efficacy and toxicity of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., ED50 (the dose therapeutically effective in 50% of the population) and LD50 (the dose lethal to 50% of the population). The dose ratio between therapeutic and toxic effects is the therapeutic index, and it can be expressed as the ratio, LD50/ED50.

### **Diagnostic means and kits**

The invention also relates to a diagnostic composition comprising at least one of the aforementioned nucleic acid molecules, vectors, proteins, antibodies or compounds and

optionally suitable means for detection. Said diagnostic compositions may be used for methods for determining expression of cell cycle interacting proteins by detecting the presence of the corresponding mRNA which comprises isolation of mRNA from a cell and contacting the mRNA so obtained with a probe comprising a nucleic acid probe as described above under hybridizing conditions, detecting the presence of mRNA hybridized to the probe, and thereby detecting the expression of the protein in the cell. Further methods of detecting the presence of a protein according to the present invention comprises immunotechniques well known in the art, for example enzyme linked immunosorbent assay. Furthermore, it is possible to use the nucleic acid molecules according to the invention as molecular markers in plant breeding. Moreover, the present invention relates to a kit comprising at least one of the aforementioned nucleic acid molecules, regulatory sequences, recombinant DNA molecules, vectors, proteins, compounds or antibodies of the invention. The kit of the invention may contain further ingredients such as selection markers and components for selective media suitable for the generation of transformed host cells and transgenic plant cells, plant tissue or plants. Furthermore, the kit may include buffers and substrates for reporter genes that may be present in the recombinant gene or vector of the invention. The kit of the invention may advantageously be used for carrying out the method of the invention and could be, inter alia, employed in a variety of applications referred to herein, e.g., in the diagnostic field or as research tool. The parts of the kit of the invention can be packaged individually in vials or in combination in containers or multicontainer units. Manufacture of the kit follows preferably standard procedures which are known to the person skilled in the art. The kit or its ingredients according to the invention can be used in plant cell and plant tissue cultures, for example, for any of the above described methods for detecting inhibitors and activators of cell cycle genes. The kit of the invention and its ingredients are expected to be very useful in breeding new varieties of, for example, plants which display improved properties such as nutritial value or disease resistance.

### Further applications of the invention

The person skilled in the art can use proteins according to the invention from other organisms such as yeast and animals to influence cell division progression in those other organisms such as mammals or insects. In a preferred embodiment one or more DNA sequences, vectors or proteins of the invention or the above-described antibody or compound are, for instance, used to specifically interfere in the disruption of the expression levels of genes involved in G1/S transition in the cell cycle process in transformed plants, particularly :

- in the complete plant
- in selected plant organs, tissues or cell types
- under specific environmental conditions, including abiotic stress such as cold, heat, drought or salt stress or biotic stress such as pathogen attack
- during specific developmental stages.

Specifically the plant cell division rate and/or the inhibition of a plant cell division can be influenced by (partial) elimination of a gene or reducing the expression of a gene encoding a protein according to the invention. Said plant cell division rate and/or the inhibition of a plant cell division can also be influenced by eliminating or inhibiting the activity of the protein according to the invention by using for instance antibodies directed against said protein. As a result of said elimination or reduction greater organisms or specific organs or tissues can be obtained; greater in volume and in mass too. Furthermore inhibition of cell division by various adverse environmental conditions such as drought, high salt content, chilling and the like can be delayed or prevented by reduction or enhancing (e.g. with a dominant negative version) of said expression of a gene according to the invention. The division rate of a plant cell can also be influenced in a transformed plant by overexpression of a nucleic acid molecule according to the invention. Therefore an important aspect of the current invention is a method to modify plant architecture by overproduction or reduction of expression of a sequence according to the invention under the control of a tissue, cell or organ specific promoter. Another aspect of the present invention is a method to modify the growth inhibition of plants

caused by environmental stress conditions above mentioned by appropriate use of sequences according to the invention. Surprisingly using a polypeptide or fragment thereof according to the invention or using antisense RNA or any method to reduce the expression of the gene according to the invention, cell division in the meristem of both main and lateral roots, shoot apical or the vascular tissue of a plant can be manipulated. Furthermore any of the DNA sequences of the invention can be used to manipulate (reduce or enhance) the level of endopolyploidy and thereby increasing the storage capacity of, for example, endosperm cells. Thus, another aspect of the current invention is that one or more DNA sequences, vectors or proteins, regulatory sequences or recombinant DNA molecules of the invention or the above-described antibody or compound can be used to modulate, for instance, endoreduplication in storage cells, storage tissues and/or storage organs of plants or parts thereof. The term "endoreduplication" means recurrent DNA replication without consequent mitosis and cytokinesis.

Preferred target storage organs and parts thereof for the modulation of endoreduplication are, for instance, seeds (such as from cereals, oilseed crops), roots (such as in sugar beet), tubers (such as in potato) and fruits (such as in vegetables and fruit species). Furthermore it is expected that increased endoreduplication in storage organs and parts thereof correlates with enhanced storage capacity and as such with improved yield. In yet another embodiment of the invention, a plant with modulated endoreduplication in the whole plant or parts thereof can be obtained from a single plant cell by transforming the cell, in a manner known to the skilled person, with the above-described means.

In view of the foregoing, the present invention also relates to the use of a DNA sequence, vector, protein, antibody, regulatory sequences, recombinant DNA molecule, nucleic acid molecules or compound of the invention for modulating plant cell cycle, plant cell division and/or growth, for influencing the activity of cell cycle interacting protein, for disrupting plant cell division by influencing the presence or absence or by interfering in the expression of a cyclin-dependent protein, for modifying growth inhibition of plants caused by environmental stress conditions, for inducing male or



female sterility, for influencing cell division progression in a host as defined above or for use in a screening method for the identification of inhibitors or activators of cell cycle proteins.

Furthermore, it is possible to use the nucleic acid molecules according to the invention as molecular markers in plant breeding. Thus, the present invention also relates to the use of a DNA sequence or regulatory sequence of the invention as a marker gene in plant or animal cell and tissue culture or as a marker in marker-assisted plant breeding. Moreover, the overexpression of nucleic acid molecules according to the invention may be useful for the alteration or modification of plant/pathogene interaction. The term "pathogene" includes, for example, bacteria, viruses and fungi as well as protozoa.

These and other embodiments are disclosed and encompassed by the description and examples of the present invention. Further literature concerning any one of the methods, uses and compounds to be employed in accordance with the present invention may be retrieved from public libraries, using for example electronic devices. For example the public database "Medline" may be utilized which is available on the Internet, for example under <http://www.ncbi.nlm.nih.gov/PubMed/medline.html>. Further databases and addresses, such as <http://www.ncbi.nlm.nih.gov/>, <http://www.infobiogen.fr/>, [http://www.fmi.ch/biology/research\\_tools.html](http://www.fmi.ch/biology/research_tools.html), <http://www.tigr.org/>, are known to the person skilled in the art and can also be obtained using, e.g., <http://www.lycos.com>. An overview of patent information in biotechnology and a survey of relevant sources of patent information useful for retrospective searching and for current awareness is given in Berks, TIBTECH 12 (1994), 352-364.

The present invention is further illustrated by reference to the following non-limiting examples.

Unless stated otherwise in the examples, all recombinant DNA techniques are performed according to protocols as described in Sambrook et al. (1989), Molecular Cloning : A Laboratory Manual. Cold Spring Harbor Laboratory Press, NY or in Volumes

1 and 2 of Ausubel et al. (1994), Current Protocols in Molecular Biology, Current Protocols. Standard materials and methods for plant molecular work are described in Plant Molecular Biology Labfase (1993) by R.D.D. Croy, jointly published by BIOS Scientific Publications Ltd (UK) and Blackwell Scientific Publications (UK).

### **Example 1: Identification of cell cycle interacting proteins using the two hybrid system with CDC2b as a bait**

A two-hybrid screening was performed using as bait a fusion between the GAL4 DNA-binding domain and *CDC2bAt*. Vectors and strains used were provided with the Matchmaker Two-Hybrid System (Clontech, Palo Alto, CA). The bait was constructed by inserting the *CDC2bAt* PCR fragment into the pGBT9 vector. The PCR fragment was created from the cDNA using primers to incorporate *EcoRI* restriction enzyme sites (5'-CGGATCCGAATTCATGGAGAACGAG-3' (SEQ ID NO: 15) and 5'-CGGATCCGAATTCTCAGAACTGAGA-3') (SEQ ID NO: 16). The PCR fragment was cut with *EcoRI* and cloned into the *EcoRI* site of pGBT9, resulting in the plasmid pGBTCDC2B. The GAL4 activation domain cDNA fusion library was obtained from Clontech from mRNA of *Arabidopsis thaliana* cell suspensions harvested at various growing stages: early exponential, exponential, early stationary, and stationary phase. For the screening a 1-liter culture of the *Saccharomyces cerevisiae* strain HF7c (*MATa ura3-52 his3-200 ade2-101 lys2-801 trp1-901 leu2-3,112 gal4-542 gal80-538 LYS2::GAL1<sub>UAS</sub>-GAL1<sub>TATA</sub>-HIS3 URA3::GAL4<sub>17mers(3x)</sub>-CyC1<sub>TATA</sub>-LacZ*) was cotransformed with 590 µg pGBTCDC2B, 1100 µg DNA of the library, and 40 mg salmon sperm carrier DNA using the lithium acetate method (Gietz *et al.*, 1992). To estimate the number of independent cotransformants, 1/1000 of the transformation mix was plated on Leu- and Trp- medium. The rest of the transformation mix was plated on medium to select for histidine prototrophy (Trp-, Leu-, His-). After 5 days of growth at 30°C, the colonies larger than 2 mm were streaked on histidine-lacking medium. A total of 10<sup>7</sup> independent cotransformants were screened for there ability to grow on histidine free medium. A 5-day incubation at 30°C yielded 352 colonies. Of the His<sup>+</sup> colonies the activation domain

plasmids were isolated as described (Hoffman and Winston, 1987, Gene 57, 267-272). The *hybriZAP*<sup>TM</sup> inserts were PCR amplified using the primers 5'-AGGGATGTTTAATACCACTAC-3' (SEQ ID NO: 17) and 5'-GCACAGTTGAAGTGAAGTTGC-3' (SEQ ID NO: 18). PCR fragments were digested with *A**lu*I and fractionized on a 2% agarose gel. Plasmid DNA of which the inserts gave rise to different restriction patterns were electroporated into *Escherichia coli* XL1-Blue, and the DNA sequence of the inserts was determined. Extracted DNA was also used to retransform HF7c to test the specificity of the interaction.

**Example 2: Identification of cell cycle interacting proteins using the two hybrid system with CDC2a as a bait**

For the identification of cell cycle interacting proteins also a two hybrid system based on GAL4 recognition sites to regulate the expression of both *his3* and *lacZ* reporter genes was used to identify CDC2aAt-interacting of proteins. The bait used for the two-hybrid screening was constructed by inserting the *CDC2aAt* coding region into the pGBT9 vector (Clontech). The insert was created by PCR using the *CDC2aAt* cDNA as template. Primers were designed to incorporate *Eco*RI restriction enzyme sites. The primers used were 5'-CGAGATCTGAATTCATGGATCAGTA-3' (SEQ ID NO: 19) and 5'-CGAGATCTGAATTCCTAAGGCATGCC-3' (SEQ ID NO: 20). The PCR fragment was cut with *Eco*RI and cloned into the *Eco*RI site of pGBT9, resulting in the pGBTCDC2A plasmid. For the screening a GAL4 activation domain cDNA fusion library was used constructed from *Arabidopsis thaliana* cell suspension cultures. This library was constructed using RNA isolated from cells harvested at 20 hours, 3, 7 and 10 days after dilution of the culture in new medium. These time point correspondent to cells from the early exponential growth phase to the late stationary phase. mRNA was prepared using Dynabeads oligo(dT)<sub>25</sub> according to the manufacturer's instructions (Dyna). The GAL4 activation domain cDNA fusion library was generated using the *HybriZAP*<sup>TM</sup> vector purchased with the *HybriZAP*<sup>TM</sup> Two-Hybrid cDNA Gigapack cloning Kit (Stratagene) following the manufacturer's instructions. The resulting library contained approximately 3.10<sup>6</sup> independent plaque-forming units, with an average insert size of 1 Kb.

For the screening a 1-liter culture of the *Saccharomyces cerevisiae* strain HF7c (*MAT<sub>a</sub>*, *ura3-52 his3-200 ade2-101 lys2-801 trp1-901 leu2-3,112 gal4-542 gal80-538 LYS2::GAL1<sub>UAS</sub>-GAL1<sub>TATA</sub>-HIS3 URA3::GAL4<sub>17mers(3x)</sub>-CyC1<sub>TATA</sub>-LacZ*) was cotransformed with 400 µg pGBTCDC2A, 500 µg DNA of the library, and 40 mg salmon sperm carrier DNA using the lithium acetate method (Gietz *et al.* 1992, *Nucleic Acids Res.* 20, 1425). To estimate the number of independent cotransformants, 1/1000 of the transformation mix was plated on Leu<sup>-</sup> and Trp<sup>-</sup> medium. The rest of the transformation mix was plated on medium to select for histidine prototrophy (Trp<sup>-</sup>, Leu<sup>-</sup>, His<sup>-</sup>). Of a total of approximately 1.2 x 10<sup>7</sup> independent transformants 1200 colonies grew after 3 days of incubation at 30°C. The colonies larger than 2 mm were streaked on histidine-lacking medium supplemented with 10 mM 3-amino-1,2,4-triazole (Sigma). Two-hundred-fifty colonies capable of growing under these conditions were tested for β-galactosidase activity as described (Breedon and Nasmyth 1995, *Cold Spring Harbor Symp. Quant. Biol.* 50, p643-650), and 153 turned out to be His<sup>+</sup> and LacZ<sup>+</sup>. Plasmid DNA was prepared from the positive clones and sequenced.

### Example 3: Cell cycle interacting proteins associating with Cdc2aAt or Cdc2bAt

Nine cDNA clones were obtained by the method described in Example 1 and 2, which are further described below. The specificity of the interaction those clones was verified by the retransformation of yeast with pGBTCDC2A or pGBTCDC2B and the corresponding cDNA clones. As controls, pGBTCDC2A or pGBTCDC2B was cotransformed with a vector containing only the GAL4 activation domain (pGAD424); and the nine cDNA vectors were each cotransformed with a plasmid containing only the GAL4 DNA binding domain (pGBT9). Transformants were plated on medium with or without histidine. Only transformants containing both pGBTCDC2A or pGBTCDC2B and one of the nine cDNA clones were able to grow in the absence of histidine.

#### 1. Vb89 (SEQ ID NO: 7)

A BLAST data base search revealed that the Vb89 clone encode the *Arabidopsis*

*thaliana HAL3* homologue, isolated recently and of which the function was unknown. Unexpectedly, the Vb89 clone interacts with CDC2bAt, but not with CDC2aAt in the two-hybrid system. The interaction of Vb89 with CDC2bAt highlights an important role of Vb 89 in cell cycle control. The publicly available databases were screened with the cDNA VB89. An overall perfect homology with HAL3, already known in the databases was found. With the help of BLASTX U80192 (score 1.9e-106) was found as the best homologue. This sequence is a partial cDNA from *A.thaliana* (entered in the databank: 28-APR-1997)(with publ.:Culianez-Macia,F.A., Espinosa-Ruiz,A. and Serrano,R, *Arabidopsis thaliana HAL3* homolog gene, Unpublished). Except that VB89 is longer, there are no major differences with this cDNA.

*HAL3* is a halotolerant gene isolated in *Saccharomyces cerevisiae* (Ferrando, 1995). Hal3p can inhibit the Ppz1 protein phosphatase resulting in an increased resistance to sodium and lithium. These effect is largely a result of the increased expression of the *ENA/PMR2A* gene. This gene codes for a P-type ATPase responsible for sodium efflux (De Nadal et al., 1998). The *HAL3* gene has also been isolated independently (as *SIS2*) and characterized on the basis of its ability to increase, when present in high copy number, the growth rate of *sit4* mutants (Di Como et al, 1995). The *SIT4/PPH1* gene encodes a type 2A-related Ser/Thr protein phosphatase that is required in late G1 for normal G1 cyclin expression and for bud formation. Interestingly, overexpression of *HAL3/SIS2* stimulates the rate of cyclin accumulation in *sit4* mutants.

De Nadal E., Clotet J., Posas F., Serrano R., gomez N., Arino J. (1998). The yeast halotolerant determinant Hal3p is an inhibitory subunit of the Ppz1p Ser/Thr protein phosphatase. *Proc. Natl. Acad. Sci. USA*, 95: 7357-7362.

Di Como C.J., Bose R., Arndt K.T. (1995). Overexpression of SIS2, Which contains an extremely acidic region, increases the expression of *SWI4*, *CLN1* and *CLN2* in *sit4* mutants. *Genetics*, 139: 95-107.

Ferrando A., Kron S.J., Rios G., Fink G.R., Serrano R. (1995). Regulation of cation transport in *Saccharomyces cerevisiae* by the salt tolerance gene *HAL3*. *Molecular and cellular Biology*, 15:5470-5481.

## 2. VbDAHP (SEQ ID NO: 9)

When a BLAST data base was used it was found that the VbDAHP clone encode a 3-deoxy-D-arabino-heptulosonate 7-phosphate (DAHP) synthase with a high similarity to the DHS2 gene. The VbDAHP clone interacts with CDC2bAt, but not with CDC2aAt in the two-hybrid system. The publicly available databases were screened with the cDNA VBDAHP (SEQ ID NO: 9). An overall perfect homology was found with DAHP (AROG\_ARATH 3-deoxy-D-arabino-heptulosonate 7-phosphate synthase [*Arabidopsis thaliana*]), already known in the databases. With the BLASTX as best homologue Q00218 (score 1.9e-49, C-term; 5.6e-86, N-term) was found. This sequence is a complete mRNA from *A.thaliana* (entered in the databank: 01-NOV-1997) (with publ.:Keith, Proc. Natl. Acad. Sci. U.S.A. 88 (19), 8821-8825 (1991)). With the BLASTN/nr we found the same DAHP.

In *Arabidopsis thaliana*, two genes has been isolated encoding 3-deoxy-D-arabino-heptulosonate 7-phosphate (DAHP) synthase, an enzyme catalyzing the first committed step in aromatic amino acid biosynthesis (Keith et al., 1991). Both genes, DHS1 and DHS2, may have distinct physiological roles, as there are differentially expressed in plants subjected either to physical wounding or to infiltration by virulent and avirulent strains of *Pseudomonas syringae*. Other enzymes in the *Arabidopsis* aromatic pathway are also encoded by duplicated genes, an arrangement that may allow independent regulation of aromatic amino acid biosynthesis by distinct physiological requirements such as protein synthesis and secondary metabolism.

Keith B., Dong X., Ausubel F.M., Fink G.R. (1991) Differential induction of 3-deoxy-D-arabino-heptulosonate 7-phosphate synthase genes in *Arabidopsis thaliana* by wounding and pathogenic attack. Proc. Natl. Acad. Sci. USA, 88: 8821-8825.

### 3. VbHSF (SEQ ID NO: 13)

A BLAST data base search revealed that the VbHSF clone is very similar to the *Arabidopsis thaliana* Heat-Shock Transcription Factor HSF3. The VbHSF clone interacts with CDC2bAt, but not with CDC2aAt with the two-hybrid system. Organisms synthesize heat shock proteins (HSPs) in response to sublethal heat stress and concomitantly acquire increased tolerance against a subsequent, otherwise lethal, heat shock. Heat shock factor (HSF) is essential for the transcription of many HSP genes. Recently two HSF genes, HSF3 and HSF4, were isolated from an *Arabidopsis* cDNA library (Prandl *et al.*, 1998). Transgenic *Arabidopsis* plants were generated containing constructs that allow expression of HSF3 and HSF4 or the respective translational beta-glucuronidase (GUS) fusions. Overexpression of HSF3 or HSF3-GUS, but not of HSF4 or HSF4-GUS, causes HSP synthesis at the non-heat-shock temperature of 25 degrees C in transgenic *Arabidopsis*. In transgenic plants bearing HSF3/HSF3-GUS, transcription of several heat shock genes is derepressed. Electrophoretic mobility shift assays suggest that derepression of the heat shock response is mediated by HSF3/HSF3-GUS functioning as transcription factor. HSF3/HSF3-GUS-overexpressing *Arabidopsis* plants show an increase in basal thermotolerance, indicating the importance of HSFs and HSF-regulated genes as determinants of thermoprotective processes. Plants transgenic for HSF3/HSF3-GUS exhibit no other obvious phenotypic alterations.

Derepression of HSF activity upon overexpression suggests the titration of a negative regulator of HSF3 or an intrinsic constitutive activity of HSF3. Stable overexpression of HSFs may be applied to other organisms as a means of derepressing the heat shock response.

A possible regulatory interaction between heat shock response and cell cycle control in plants has already been suggested. Reindl *et al.* (1997) reported the phosphorylation of the *Arabidopsis* heat-shock transcription factor HSF1 by a cyclin-dependent kinase. The HSF1 kinase forms a stable complex with AtHSF1. The HSF1 kinase interacts with the cell-cycle control protein Suc1p and is immunoprecipitated by an antibody specific for

the *Arabidopsis* cyclin-dependent CDC2a kinase. Phosphorylation by CDC2a in vitro inhibits DNA binding of AtHSF1 to the cognate heat-shock elements.

Different studies have shown that Heat shock factors can serve as auxillary proteins in formation of CDK/cyclin complexes. For example during meiosis I of mouse spermatocytes it is proposed that HSP70-2 assists in CDC2/cylinB1 complex formation through interaction with CDC2 and that this interaction establishes and/or maintains the CDC2 protein in a conformation that is competent for cyclin B1 binding (Zhu et al, 1997).

Prandl R, Hinderhofer K, Eggers-Schumacher G, Schoffl F. (1998). HSF3, a new heat shock factor from *Arabidopsis thaliana*, derepresses the heat shock response and confers thermotolerance when overexpressed in transgenic plants. Mol Gen Genet 1998 May;258(3):269-78.

Reindl A, Schoffl F, Schell J, Koncz C, Bako L (1997). Phosphorylation by a cyclin-dependent kinase modulates DNA binding of the *Arabidopsis* heat-shock transcription factor HSF1 in vitro: Plant Physiol 1997 Sep;115(1):93-100.

Zhu D., Dix D.J., Eddy E.M. (1997). HSP70-2 is required for CDC2 kinase activity in meiosis I of mouse spermatocytes. Development 124: 3007-3014.

#### **4. VbDBP(SEQ ID NO: 11)**

When a BLAST data base was used it was found that the VbDBP clone is very similar to the putative DNA binding protein (*Arabidopsis thaliana*) and also contains a lot of homologies with PCF2 (*Oryza sativa*). VbDBP interacts with CDC2b but not with CDC2a. The publicly available databases were screened with the cDNA VBDPBP (N-term). With the help of BLASTX gene21 from AC003680 (score 1.0e-27) was found as best homologue. This is a genomic sequence from A.thaliana (entered in the databank:20-MAR-1998), chromosome II. The prediction made here gives 1 big exon, but the new predictions made in accordance with the present invention gave two exons



(the big one, followed by a small one). The cDNA VBDPBP shows not so high homology (gene 21 might only be from the same family as VBDPBP) with the big exon, so completion of the cDNA will confirm one or the other annotation and might give a new sequence. Other homologues are D87261) PCF2 [*Oryza sativa*] (score  $9.2e-27$ ) and D87260) PCF1 [*Oryza sativa*] (score  $8.5e-24$ ) both with publication: Kosugi, S. and Ohashi, Y. PCF1 and PCF2 specifically bind to cis elements in the rice proliferating cell nuclear antigen gene. *Plant Cell* 9 (9), 1607-1619 (1997). With the help of BLASTN/nr an other genomic sequence from chromosome V, AB010072 ( $2e-12$ ) (08-JAN-1998) sequenced by the KAOS-people (P1 clone: MEE6) was found. The region with homology is located between (18754..18848) has no annotations at all. The publicly available databases was screened with the cDNA VBDPBP (C-term (SEQ ID NO: 15)) but nothing was found with BLASTX.

PCF1 and PCF2 are proteins isolated in rice that specifically bind to sites IIa and IIb in the promotor region of the rice PCNA gene (Kosugi et al., 1997). The rice proliferating cell nuclear antigen (PCNA) protein is an auxiliary protein of DNA polymerase (that participates in a variety of processes, such as DNA replication, DNA repair synthesis, and cell cycle control through reactions with the CDK-cyclin-CKI complex. The PCNA gene is induced at the G1-to-S phase boundary and is well conserved in eukaryotes. The expression of the rice PCNA gene is restricted exclusively to meristematic regions and is controlled at the transcriptional phase. PCNA protein is also present in proliferating cells but absent from nondividing cells and terminally differentiated plant tissues.

Loss-of-function analysis of the rice PCNA promoter using transgenic plants has demonstrated that two elements (sites IIa and IIb) in the proximal region are essential for the proliferating cell-specific transcriptional activity. On the other hand, two repeated site IIa sequences located upstream of the cauliflower mosaic virus 35S minimal promoter confer transcriptional activation in tobacco protoplast. This suggests that sites IIa and IIb most probably function as positive cis-acting elements in proliferating cells.

The proteins PCF1 and PCF2 specifically bind to sites IIa and IIb in the promoter region of the rice PCNA gene and may act as transcription factors to control DNA synthesis-

related genes in plants. In particular, PCF2, with a high level of DNA binding activity in meristematic tissues, may act as transcriptional activator for these genes. These proteins have a deduced basic helix-loop-helix (bHLH) motif that is responsible for DNA binding and dimerization. PCF1 and PCF2 are novel types of bHLH proteins that are distinct from other known bHLH transcriptional factors.

Kosugi, S., and Ohasi Y. (1997) PCF1 and PCF2 specifically bind to cis elements in the Rice proliferating cell nuclear antigen gene. *The Plant Cell*, 9, 1607-1619.

#### **5. Vb33 (SEQ ID NO: 5)**

The Vb33 clone encodes a protein interacting with CDC2b but not with CDC2a. The publicly available databases were screened with the cDNA VB33. With the BLASTX as best homologue a predicted gene on the Z49937 sequence having a similarity with an ankyrin motif (score 0.62) was found. This sequence comes from *C.elegans* cosmid and the gene F14F3.2 was predicted based on a *C.elegans* EST (yk192g4.5).

#### **6. LDV24 (SEQ ID NO: 3)**

The LDV24 gene encodes a protein interacting with CDC2a and being highly similar to the PREG1 and PHO80 proteins of *Neurospora crassa* and *Saccharomyces cerevisiae*, respectively. The publicly available databases were screened with the cDNA LDV24. With the BLASTX as best homologue the PREG(AF051226) protein from *Picea mariana* (score: 1.5e-35) and PREG(AC003672) protein from *Arabidopsis* (score: 3.1e-35) were found. But there is homology with (P20052|PH80\_YEAST) PHOSPHATE SYSTEM CYCLIN PHO80 (score: 2.1e-10). With the BLASTN/nr we found AF051226 *Picea mariana* PREG-like protein (score: 3.9e-12). Functional domains are predicted at amino acid positions 61-168 and 73-171 as comprising putative cyclin like interacting domains.

PHO80 itself shows similarity to the *Saccharomyces cerevisiae* G1-specific cyclins HCS26 and OrfD (Kaffman Science 263 (1994) 1153-1155). The catalytic CDK subunit binding to PHO80 is PHO85, a CDK with roles in both the cell cycle and metabolic controls (Lenburg and O'Shea 1996, TIBS 21, p383-387). PHO80 in complex with PHO85 regulates phosphatase gene expression. When inorganic phosphate in the medium is abundant the PHO80-PHO85 complex phosphorylates the PHO4 transcription factor. Phosphorylated PHO4 remains mainly cytoplasmic, resulting in the repression of expression of the PHO5 phosphatase gene (O'Neill et al. 1996, Science 271, p209-212). When cell are starved for phosphate, the PHO80-PHO85 complex is inhibited by the CDK inhibitor PHO81, and transcription of PHO5 is activated.

The levels of PHO5 expression are sensitive to the levels of PHO80. Overexpression of PHO80 results in a partial defect of PHO5 activation when phosphate is limiting (Yoshida et al. 1989, MGG 217, p40-46; Madden et al. 1988, Nucleic Acids Res. 16, p2625-2637). At the other hand, deletion of PHO80 results in the presence of high levels of inorganic phosphate (Madden et al. 1988, Nucleic Acids Res. 16, p2625-2637). Similar effects can be expected for plants when the *LDV24* genes is deleted or overexpressed. This might result in an adapted growth in conditions where organic phosphate is present at limiting or exceeding levels. More phosphate accumulation might positively affect the rate of plant growth and biomass production.

#### **7. LDV115 (SEQ ID NO: 1)**

The *LDV115* gene encodes a protein interacting with CDC2a but not with CDC2b and showing limited similarity to the *Saccharomyces cerevisiae* WEB1 protein. The publicly available databases were screened with the cDNA LDV115. With the BLASTX it was found as best homologue the WEB1 protein from *S.pombe* (AB004537)(score 6.7e-17). This protein as well as the other hits were mainly due to proline-richness of the LDV115 translation. The homology is low but spread over about 50% of the *S.pombe* protein, which might indicate that LDV115 is at least a member of the family. The *WEB1* gene was isolated as a yeast homologue of the adenoviral E1A gene (Zieler et al., 1995, MCB

15, p3227-3237). The protein products of the E1A gene are implicated in a variety of transcriptional and cell cycle events, involving interactions with several proteins present in the human cells, including parts of the transcriptional machinery and negative regulators of cell division such as the Rb gene product and p107. WEB1 is identical to SEC31, a protein involved in budding of transport vesicles from the endoplasmic reticulum (Pryer et al. 1993, J. Cell. Biol. 120, p865-875). The protein similarity between WEB1 and LDV115 is almost completely due to the presence of a proline-rich region found in both proteins. Proline-rich regions are not restricted to the WEB1 protein, but can also be found in many structural proteins such as hydroxyproline-rich glycoproteins and extensins. Therefore, LDV115 might not be a true homologue of WEB1.

**Example 4: Extension of cell cycle interacting protein encoding polynucleotides to full length or to recover regulatory elements**

The cell cycle interacting protein encoding nucleic acid sequences (SEQ ID NOS: 1, 3, 5, 7, 9, 11 and 13) are used to design oligonucleotide primers for extending a partial nucleotide sequence to full length or for obtaining 5' sequences from genomic libraries. One primer is synthesized to initiate extension in the antisense direction (XLR) and the other is synthesized to extend sequence in the sense direction (XLF). Primers allow the extension of the known cell cycle interacting protein encoding sequence "outward" generating amplicons containing new, unknown nucleotide sequence for the region of interest. The initial primers are designed from the cDNA using OLIGO® 4.06 Primer Analysis Software (National Biosciences), or another appropriate program, to be preferably 22-30 nucleotides in length, to have a GC content of preferably 50% or more, and to anneal to the target sequence at temperatures preferably about 68°-72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations is avoided. The original, selected cDNA libraries, prepared from mRNA isolated from actively dividing cells or a plant genomic library are used to extend the sequence; the latter is most useful to obtain 5' upstream regions. If more extension is necessary or desired, additional sets of primers are designed to further extend the

known region. By following the instructions for the XL-PCR kit (Perkin Elmer) and thoroughly mixing the enzyme and reaction mix, high fidelity amplification is obtained. Beginning with 40 pmol of each primer and the recommended concentrations of all other components of the kit, PCR is performed using the Peltier Thermal Cycle (PTC200; MJ Research, Watertown MA) and the following parameters:

- |         |   |
|---------|---|
| Step 1  | 94°C for 1 min (initial denaturation)     |
| Step 2  | 65°C for 1 min                            |
| Step 3  | 68°C for 6 min                            |
| Step 4  | 94° for 15 sec                            |
| Step 5  | 65°C for 1 min                            |
| Step 6  | 68°C for 7 min                            |
| Step 7  | Repeat steps 4-6 for 15 additional cycles |
| Step 8  | 94°C for 15 sec                           |
| Step 9  | 65°C for 1 min                            |
| Step 10 | 68°C for 7:15 min                         |
| Step 11 | Repeat step 8-10 for 12 cycles            |
| Step 12 | 72°C for 8 min                            |
| Step 13 | 4°C (and holding)                         |

A 5-10 µl aliquot of the reaction mixture is analyzed by electrophoresis on a low concentration (about 0.6-0.8%) agarose mini-gel to determine which reactions were successful in extending the sequence. Bands thought to contain the largest products were selected and cut out of the gel. Further purification involves using a commercial gel extraction method such as QIAQuick™ (QIAGEN Inc). After recovery of the DNA, Klenow enzyme was used to trim single-stranded, nucleotide overhangs creating blunt ends which facilitate religation and cloning. After ethanol precipitation, the products are redissolved in 13 µl of ligation buffer, 1µl T4-DNA ligase (15 units) and 1 µl T4 polynucleotide kinase are added, and the mixture is incubated at room temperature for 2-3 hours or overnight at 16°C. Competent *E. coli* cells (in 40 µl of appropriate media) are transformed with 3 µl of ligation mixture and cultured in 80 µl of SOC medium

(Sambrook, supra). After incubation for one hour at 37°C, the whole transformation mixture is plated on Luria Bertani (LB)-agar (Sambrook, supra) containing 2xCarb. The following day, several colonies are randomly picked from each plate and cultured in 150 µl of liquid LB/2xCarb medium placed in an individual well of an appropriate, commercially-available, sterile 96-well microtiter plate. The following day, 5 µl of each overnight culture is transferred into a non-sterile 96-well plate and after dilution 1:10 with water, 5 µl of each sample is transferred into a PCR array. For PCR amplification, 18 µl of concentrated PCR reaction mix (3.3x) containing 4 units of 4Tth DNA polymerase, a vector primer and both of the gene specific primers used for the extension reaction are added to each well. Amplification is performed using the following conditions:

Step 1	94°C for 60 sec
Step 2	94°C for 20 sec
Step 3	55°C for 30 sec
Step 4	72°C for 90 sec
Step 5	Repeat steps 2-4 for an additional 29 cycles
Step 6	72°C for 180 sec
Step 7	4°C (and holding)

Aliquots of the PCR reactions are run on agarose gels together with molecular weight markers. The sizes of the PCR products are compared to the original partial cDNAs, and appropriate clones are selected, ligated into plasmid and sequenced.

75

## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

## (i) APPLICANT:

(A) NAME: CropDesign N.V.  
(B) STREET: Technologiepark 3  
(C) CITY: Zwijnaarde-Gent  
(D) STATE: none  
(E) COUNTRY: Belgium  
(F) POSTAL CODE (ZIP): 9052

(ii) TITLE OF INVENTION: Novel cell cycle genes and uses thereof

(iii) NUMBER OF SEQUENCES: 20

## (iv) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk  
(B) COMPUTER: IBM PC compatible  
(C) OPERATING SYSTEM: PC-DOS/MS-DOS  
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25 (EPO)

## (2) INFORMATION FOR SEQ ID NO: 1:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1989 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

## (ix) FEATURE:

(A) NAME/KEY: CDS  
(B) LOCATION: 2..1672

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

A ACG CAA GAA ATG CAA GAA GAA GAG GAA GAA AGT TCT GAC CCA GTT  
Thr Gln Glu Met Gln Glu Glu Glu Glu Ser Ser Asp Pro Val  
1 5 10 15

46

76

TTT GAT AAT GCC ATC CAG CGA GCG TTG ATT GTT GGA GAT TAC AAG GAG Phe Asp Asn Ala Ile Gln Arg Ala Leu Ile Val Gly Asp Tyr Lys Glu	94
20 25 30	
GCG GTG GAT CAG TGT ATA ACT GCA AAT AAG ATG GCC GAT GCT TTA GTT Ala Val Asp Gln Cys Ile Thr Ala Asn Lys Met Ala Asp Ala Leu Val	142
35 40 45	
ATT GCT CAT GTT GGT GGT ACA GCG TTG TGG GAG AGT ACT CGT GAG AAA Ile Ala His Val Gly Gly Thr Ala Leu Trp Glu Ser Thr Arg Glu Lys	190
50 55 60	
TAT TTG AAG ACG AAC AGT GCG CCA TAC ATG AAG GTT GTT TCT GCG ATG Tyr Leu Lys Thr Asn Ser Ala Pro Tyr Met Lys Val Val Ser Ala Met	238
65 70 75	
GTG AAC AAT GAT CTC AGG AGC CTT ATC TAT ACA AGG TCA CAT AAG TTC Val Asn Asn Asp Leu Arg Ser Leu Ile Tyr Thr Arg Ser His Lys Phe	286
80 85 90 95	
TGG AAA GAG ACT CTT GCT CTC CTC TGT ACT TTT GCA CAA GGA GAA CAA Trp Lys Glu Thr Leu Ala Leu Leu Cys Thr Phe Ala Gln Gly Glu Gln	334
100 105 110	
TGG ACA ACC CTG TGT GAT GCC CTT GCC TCG AAG TTG ATG GCT GCT GGT Trp Thr Thr Leu Cys Asp Ala Leu Ala Ser Lys Leu Met Ala Ala Gly	382
115 120 125	
AAC ACT TTG GCT GCA GTT CTC TGC TAC ATT TGC GCA GGC AAT GTT GAC Asn Thr Leu Ala Ala Val Leu Cys Tyr Ile Cys Ala Gly Asn Val Asp	430
130 135 140	
AGA ACA GTA GAA ATT TGG TCG AGG AGC CTT GCA AAT GAG CGG GAT GGA Arg Thr Val Glu Ile Trp Ser Arg Ser Leu Ala Asn Glu Arg Asp Gly	478
145 150 155	
AGA TCT TAT GCT GAG CTT CTT CAA GAT CTT ATG GAG AAG ACT CTT GTC Arg Ser Tyr Ala Glu Leu Leu Gln Asp Leu Met Glu Lys Thr Leu Val	526
160 165 170 175	
CTT GCT TTG GCA ACT GGC AAC AAA AAG TTC AGC GCA TCT CTG TGT AAA Leu Ala Leu Ala Thr Gly Asn Lys Lys Phe Ser Ala Ser Leu Cys Lys	574
180 185 190	
CTC TTT GAG AGT TAT GCT GAG ATA CTG GCC AGC CAA GGG CTT CTT ACA Leu Phe Glu Ser Tyr Ala Glu Ile Leu Ala Ser Gln Gly Leu Leu Thr	622
195 200 205	
ACG GCA ATG AAG TAC TTG AAA GTT CTG GAT TCT GGT GGC TTG TCA CCT Thr Ala Met Lys Tyr Leu Lys Val Leu Asp Ser Gly Gly Leu Ser Pro	670
210 215 220	
GAA CTT TCA ATA TTA CGT GAT CGT ATT TCT CTA TCT GCA GAA CCT GAG Glu Leu Ser Ile Leu Arg Asp Arg Ile Ser Leu Ser Ala Glu Pro Glu	718



77

225	230	235	
ACT AAC ACT ACA GCT TCA GGA AAC ACT CAG CCT CAA AGC ACC ATG CCA			766
Thr Asn Thr Thr Ala Ser Gly Asn Thr Gln Pro Gln Ser Thr Met Pro			
240	245	250	255
TAT AAT CAG GAG CCA ACT CAG GCG CAA CCA AAC GTT CTT GCT AAC CCA			814
Tyr Asn Gln Glu Pro Thr Gln Ala Gln Pro Asn Val Leu Ala Asn Pro			
260	265	270	
TAT GAT AAT CAG TAT CAG CAA CCG TAC ACT GAT TCT TAT TAT GTC CCT			862
Tyr Asp Asn Gln Tyr Gln Gln Pro Tyr Thr Asp Ser Tyr Tyr Val Pro			
275	280	285	
CAA GTT TCA CAT CCA CCC ATG CAG CAA CCA ACC ATG TTT ATG CCA CAC			910
Gln Val Ser His Pro Pro Met Gln Gln Pro Thr Met Phe Met Pro His			
290	295	300	
CAA GCT CAG CCA GCT CCG CAG CCA TCT TTT ACT CCA GCT CCT ACA AGC			958
Gln Ala Gln Pro Ala Pro Gln Pro Ser Phe Thr Pro Ala Pro Thr Ser			
305	310	315	
AAT GCT CAG CCA TCC ATG AGA ACT ACA TTT GTT CCT TCA ACT CCC CCT			1006
Asn Ala Gln Pro Ser Met Arg Thr Thr Phe Val Pro Ser Thr Pro Pro			
320	325	330	335
GCA CTG AAG AAT GCA GAT CAA TAT CAG CAG CCA ACC ATG AGT TCT CAT			1054
Ala Leu Lys Asn Ala Asp Gln Tyr Gln Gln Pro Thr Met Ser Ser His			
340	345	350	
TCA TTC ACG GGA CCA TCT AAC AAT GCA TAC CCT GTT CCC CCG GGT CCT			1102
Ser Phe Thr Gly Pro Ser Asn Asn Ala Tyr Pro Val Pro Pro Gly Pro			
355	360	365	
GGT CAA TAT GCA CCT TCT GGC CCT TCA CAA CTT GGG CAA TAT CCT AAC			1150
Gly Gln Tyr Ala Pro Ser Gly Pro Ser Gln Leu Gly Gln Tyr Pro Asn			
370	375	380	
CCT AAG ATG CCC CAA GTT GTT GCT CCA GCA GCT GGA CCC ATA GGA TTT			1198
Pro Lys Met Pro Gln Val Val Ala Pro Ala Ala Gly Pro Ile Gly Phe			
385	390	395	
ACG CCC ATG GCA ACT CCA GGA GTT GCT CCA AGA TCT GTG CAA CCA GCA			1246
Thr Pro Met Ala Thr Pro Gly Val Ala Pro Arg Ser Val Gln Pro Ala			
400	405	410	415
AGT CCT CCA ACA CAG CAG GCA GCT GCA CAG GCA GCC CCT GCG CCT GCA			1294
Ser Pro Pro Thr Gln Gln Ala Ala Ala Gln Ala Ala Pro Ala Pro Ala			
420	425	430	
ACT CCG CCA CCA ACT GTT CAG ACT GCA GAT ACT TCC AAC GTT CCA GCC			1342
Thr Pro Pro Pro Thr Val Gln Thr Ala Asp Thr Ser Asn Val Pro Ala			
435	440	445	
CAC CAG AAA CCT GTG ATA GCA ACG TTG ACA AGG CTT TTC AAT GAG ACA			1390

78

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His Gln Lys Pro Val Ile Ala Thr Leu Thr Arg Leu Phe Asn Glu Thr
    450                                455                                460

TCG GAA GCA CTG GGA GGC GCA CGT GCG AAT ACT ACT AAG AAG CGT GAG      1438
Ser Glu Ala Leu Gly Gly Ala Arg Ala Asn Thr Thr Lys Lys Arg Glu
    465                                470                                475

ATA GAA GAC AAC TCG AGA AAA TTA GGT GCT CTG TTT GTG AAA CTC AAC      1486
Ile Glu Asp Asn Ser Arg Lys Leu Gly Ala Leu Phe Val Lys Leu Asn
    480                                485                                490                                495

AGC GGA GAC ATC TCC AAG AAT GCT GCG GAC AAA CTC GCA CAG CTA TGC      1534
Ser Gly Asp Ile Ser Lys Asn Ala Ala Asp Lys Leu Ala Gln Leu Cys
    500                                505                                510

CAA GCT CTG GAC AAC AAT GAC TTC AGC ACA GCC CTT CAA ATA CAG GTA      1582
Gln Ala Leu Asp Asn Asn Asp Phe Ser Thr Ala Leu Gln Ile Gln Val
    515                                520                                525

CTT CTG ACT ACC AGC GAA TGG GAC GAA TGC AAC TTC TGG CTG GCA ACA      1630
Leu Leu Thr Thr Ser Glu Trp Asp Glu Cys Asn Phe Trp Leu Ala Thr
    530                                535                                540

CTA AAG CGG ATG ATG GTC AAG GCC AGG CAA AAT GTG CCG TGAT TAT TTA      1679
Leu Lys Arg Met Met Val Lys Ala Arg Gln Asn Val Arg
    545                                550                                555

TTTTCTGGTT CATGGATTTT TTTTATTATA AATTTTAAGG AGGGACGTGT GTATCAAACT      1739

CCTTTTGCTT TCTTAATTTT UGTTTTTTTA AAACGCCGTT GCTGCTCTAA TTTTTTTTTT      1799

TTTTTTTTTT TGTCATTTAT GAAGCTCATC TGCTACTTCC AGTTACTTTT TTGTTTAGAT      1859

AGTATAGAGA TCATCAGATT GCTGAAACAT TTTCATGTTT TTGGATGTTA CTTTACCGGA      1919

CAAGTTTGTT CGTCTTTCTT TCTAAAAAAA AAAAAAAAAA AAAAAAAAAA ACAAAAAAAAAA      1979

AAAAAAAAAA                                                                1989

```

## (2) INFORMATION FOR SEQ ID NO: 2:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 556 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE-TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

```

Thr Gln Glu Met Gln Glu Glu Glu Ser Ser Asp Pro Val Phe
  1           5           10           15

Asp Asn Ala Ile Gln Arg Ala Leu Ile Val Gly Asp Tyr Lys Glu Ala

```

79

				20					25							30	
Val	Asp	Gln	Cys	Ile	Thr	Ala	Asn	Lys	Met	Ala	Asp	Ala	Leu	Val	Ile		
		35					40					45					
Ala	His	Val	Gly	Gly	Thr	Ala	Leu	Trp	Glu	Ser	Thr	Arg	Glu	Lys	Tyr		
	50					55					60						
Leu	Lys	Thr	Asn	Ser	Ala	Pro	Tyr	Met	Lys	Val	Val	Ser	Ala	Met	Val		
65					70					75					80		
Asn	Asn	Asp	Leu	Arg	Ser	Leu	Ile	Tyr	Thr	Arg	Ser	His	Lys	Phe	Trp		
				85					90					95			
Lys	Glu	Thr	Leu	Ala	Leu	Leu	Cys	Thr	Phe	Ala	Gln	Gly	Glu	Gln	Trp		
			100					105					110				
Thr	Thr	Leu	Cys	Asp	Ala	Leu	Ala	Ser	Lys	Leu	Met	Ala	Ala	Gly	Asn		
		115					120					125					
Thr	Leu	Ala	Ala	Val	Leu	Cys	Tyr	Ile	Cys	Ala	Gly	Asn	Val	Asp	Arg		
	130					135					140						
Thr	Val	Glu	Ile	Trp	Ser	Arg	Ser	Leu	Ala	Asn	Glu	Arg	Asp	Gly	Arg		
145					150					155					160		
Ser	Tyr	Ala	Glu	Leu	Leu	Gln	Asp	Leu	Met	Glu	Lys	Thr	Leu	Val	Leu		
				165					170					175			
Ala	Leu	Ala	Thr	Gly	Asn	Lys	Lys	Phe	Ser	Ala	Ser	Leu	Cys	Lys	Leu		
			180					185					190				
Phe	Glu	Ser	Tyr	Ala	Glu	Ile	Leu	Ala	Ser	Gln	Gly	Leu	Leu	Thr	Thr		
		195					200					205					
Ala	Met	Lys	Tyr	Leu	Lys	Val	Leu	Asp	Ser	Gly	Gly	Leu	Ser	Pro	Glu		
	210					215					220						
Leu	Ser	Ile	Leu	Arg	Asp	Arg	Ile	Ser	Leu	Ser	Ala	Glu	Pro	Glu	Thr		
225					230					235					240		
Asn	Thr	Thr	Ala	Ser	Gly	Asn	Thr	Gln	Pro	Gln	Ser	Thr	Met	Pro	Tyr		
				245					250					255			
Asn	Gln	Glu	Pro	Thr	Gln	Ala	Gln	Pro	Asn	Val	Leu	Ala	Asn	Pro	Tyr		
			260					265					270				
Asp	Asn	Gln	Tyr	Gln	Gln	Pro	Tyr	Thr	Asp	Ser	Tyr	Tyr	Val	Pro	Gln		
		275					280					285					
Val	Ser	His	Pro	Pro	Met	Gln	Gln	Pro	Thr	Met	Phe	Met	Pro	His	Gln		
	290					295					300						
Ala	Gln	Pro	Ala	Pro	Gln	Pro	Ser	Phe	Thr	Pro	Ala	Pro	Thr	Ser	Asn		
305					310						315				320		

Ala Gln Pro Ser Met Arg Thr Thr Phe Val Pro Ser Thr Pro Pro Ala  
 325 330 335  
 Leu Lys Asn Ala Asp Gln Tyr Gln Gln Pro Thr Met Ser Ser His Ser  
 340 345 350  
 Phe Thr Gly Pro Ser Asn Asn Ala Tyr Pro Val Pro Pro Gly Pro Gly  
 355 360 365  
 Gln Tyr Ala Pro Ser Gly Pro Ser Gln Leu Gly Gln Tyr Pro Asn Pro  
 370 375 380  
 Lys Met Pro Gln Val Val Ala Pro Ala Ala Gly Pro Ile Gly Phe Thr  
 385 390 395 400  
 Pro Met Ala Thr Pro Gly Val Ala Pro Arg Ser Val Gln Pro Ala Ser  
 405 410 415  
 Pro Pro Thr Gln Gln Ala Ala Ala Gln Ala Ala Pro Ala Pro Ala Thr  
 420 425 430  
 Pro Pro Pro Thr Val Gln Thr Ala Asp Thr Ser Asn Val Pro Ala His  
 435 440 445  
 Gln Lys Pro Val Ile Ala Thr Leu Thr Arg Leu Phe Asn Glu Thr Ser  
 450 455 460  
 Glu Ala Leu Gly Gly Ala Arg Ala Asn Thr Thr Lys Lys Arg Glu Ile  
 465 470 475 480  
 Glu Asp Asn Ser Arg Lys Leu Gly Ala Leu Phe Val Lys Leu Asn Ser  
 485 490 495  
 Gly Asp Ile Ser Lys Asn Ala Ala Asp Lys Leu Ala Gln Leu Cys Gln  
 500 505 510  
 Ala Leu Asp Asn Asn Asp Phe Ser Thr Ala Leu Gln Ile Gln Val Leu  
 515 520 525  
 Leu Thr Thr Ser Glu Trp Asp Glu Cys Asn Phe Trp Leu Ala Thr Leu  
 530 535 540  
 Lys Arg Met Met Val Lys Ala Arg Gln Asn Val Arg  
 545 550 555

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 814 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 3..665

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

GG GAC TCT CTC GCA ACC GAT CCA GCT TTC ATT GAT TCG GAT GTA TAC	47
Asp Ser Leu Ala Thr Asp Pro Ala Phe Ile Asp Ser Asp Val Tyr	
1 5 10 15	
CTC AGG TTA GGA CTT ATT ATT GAG GGC AAA CGA TTG AAA AAG CCA CCG	95
Leu Arg Leu Gly Leu Ile Ile Glu Gly Lys Arg Leu Lys Lys Pro Pro	
20 25 30	
ACT GTT CTC TCA CGC CTC TCT TCT TCT CTG GAG AGA TCT CTG TTA CTC	143
Thr Val Leu Ser Arg Leu Ser Ser Ser Leu Glu Arg Ser Leu Leu Leu	
35 40 45	
AAT CAT GAT GAC AAG ATT CTG CTT GGA TCG CCA GAC TCT GTT ACC GTG	191
Asn His Asp Asp Lys Ile Leu Leu Gly Ser Pro Asp Ser Val Thr Val	
50 55 60	
TTT GAC GGG AGA TCT CCC CCT GAG ATC AGT ATT GCA CAC TAC TTG GAT	239
Phe Asp Gly Arg Ser Pro Pro Glu Ile Ser Ile Ala His Tyr Leu Asp	
65 70 75	
CGC ATT TTC AAG TAC TCT TGC TGC AGT CCC TCC TGC TTC GTC ATT GCG	287
Arg Ile Phe Lys Tyr Ser Cys Cys Ser Pro Ser Cys Phe Val Ile Ala	
80 85 90 95	
CAT ATC TAC ATT GAT CAC TTT CTC CAT AAG ACC CGA GCC CTT CTC AAA	335
His Ile Tyr Ile Asp His Phe Leu His Lys Thr Arg Ala Leu Leu Lys	
100 105 110	
CCC CTT AAT GTC CAC CGC CTT ATC ATT ACA ACT GTC ATG TTA GCT GCT	383
Pro Leu Asn Val His Arg Leu Ile Ile Thr Thr Val Met Leu Ala Ala	
115 120 125	
AAA GTC TTC GAT GAT AGG TAT TTC AAC AAT GCA TAC TAC GCA AGA GTG	431
Lys Val Phe Asp Asp Arg Tyr Phe Asn Asn Ala Tyr Tyr Ala Arg Val	
130 135 140	
GGA GGT GTG ACT ACG AGA GAG TTA AAC AGA TTG GAG ATG GAG TTG TTG	479
Gly Gly Val Thr Thr Arg Glu Leu Asn Arg Leu Glu Met Glu Leu Leu	
145 150 155	
TTT ACC CTT GAC TTC AAG CTT CAG GTA GAT CCT CAG ACG TTT CAC ACA	527
Phe Thr Leu Asp Phe Lys Leu Gln Val Asp Pro Gln Thr Phe His Thr	
160 165 170 175	

CAC TGT TGT CAG TTA GAA AAG CAG AAC AGA GAC GGC TTC CAG ATC GAG 575  
 His Cys Cys Gln Leu Glu Lys Gln Asn Arg Asp Gly Phe Gln Ile Glu  
 180 185 190  
 TGG CCC ATA AAA GAA GCA TGC CGA GCC AAC AAA GAG ACT TGG CAG AAG 623  
 Trp Pro Ile Lys Glu Ala Cys Arg Ala Asn Lys Glu Thr Trp Gln Lys  
 195 200 205  
 AGG ACA CCC GAC TCA TTC TGC TCT CAA ACC ACA GCA CGC TGATCGGCAA 672  
 Arg Thr Pro Asp Ser Phe Cys Ser Gln Thr Thr Ala Arg  
 210 215 220  
 GGGTAAGATA GGATTATTTT GTGTTT TAGT AGTGATGATT CTTTTCATG ATTGATTGTT 732  
 TGTGACAATT GTGTGTAGTA GAAAATCTGA AAATTTCTAC CAACTCATT CTTAAGAAGT 792  
 TGCTAAAAAA AAAAAAAAAA AA 814

## (2) INFORMATION FOR SEQ ID NO: 4:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 220 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Asp Ser Leu Ala Thr Asp Pro Ala Phe Ile Asp Ser Asp Val Tyr Leu  
 1 5 10 15  
 Arg Leu Gly Leu Ile Ile Glu Gly Lys Arg Leu Lys Lys Pro Pro Thr  
 20 25 30  
 Val Leu Ser Arg Leu Ser Ser Ser Leu Glu Arg Ser Leu Leu Asn  
 35 40 45  
 His Asp Asp Lys Ile Leu Leu Gly Ser Pro Asp Ser Val Thr Val Phe  
 50 55 60  
 Asp Gly Arg Ser Pro Pro Glu Ile Ser Ile Ala His Tyr Leu Asp Arg  
 65 70 75 80  
 Ile Phe Lys Tyr Ser Cys Cys Ser Pro Ser Cys Phe Val Ile Ala His  
 85 90 95  
 Ile Tyr Ile Asp His Phe Leu His Lys Thr Arg Ala Leu Leu Lys Pro  
 100 105 110  
 Leu Asn Val His Arg Leu Ile Ile Thr Thr Val Met Leu Ala Ala Lys  
 115 120 125

83

Val Phe Asp Asp Arg Tyr Phe Asn Asn Ala Tyr Tyr Ala Arg Val Gly  
 130 135 140

Gly Val Thr Thr Arg Glu Leu Asn Arg Leu Glu Met Glu Leu Leu Phe  
 145 150 155 160

Thr Leu Asp Phe Lys Leu Gln Val Asp Pro Gln Thr Phe His Thr His  
 165 170 175

Cys Cys Gln Leu Glu Lys Gln Asn Arg Asp Gly Phe Gln Ile Glu Trp  
 180 185 190

Pro Ile Lys Glu Ala Cys Arg Ala Asn Lys Glu Thr Trp Gln Lys Arg  
 195 200 205

Thr Pro Asp Ser Phe Cys Ser Gln Thr Thr Ala Arg  
 210 215 220

## (2) INFORMATION FOR SEQ ID NO: 5:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1269 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA

## (iii) HYPOTHETICAL: NO

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1269

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

AAT TCG GCA CGA GGC CTT CTC CAG CTT CAT CCT TGC AAC AAG GTG GTA	48
Asn Ser Ala Arg Gly Leu Leu Gln Leu His Pro Cys Asn Lys Val Val	
1 5 10 15	
CTC TGG GGT CTT TCT CAT CAG ATA TTT GTC GGC TGC TGC AGC TCT GTG	96
Leu Trp Gly Leu Ser His Gln Ile Phe Val Gly Cys Cys Ser Ser Val	
20 25 30	
ATG GAA GAT GAT GCT ACT AGC AAA TTA GCT GCC CCC AAG CCC GAG CCT	144
Met Glu Asp Asp Ala Thr Ser Lys Leu Ala Ala Pro Lys Pro Glu Pro	
35 40 45	
GCT GAT CAG AAT CTC GAA GCT GGC AAA GCT GCT GTC TTC CAA AGG GGA	192
Ala Asp Gln Asn Leu Glu Ala Gly Lys Ala Ala Val Phe Gln Arg Gly	
50 55 60	

84

TAC AAT TTG GTT CAG GGG AAG TCA GAA CAT GGA TTA CCA TTG GTT GAT	240
Tyr Asn Leu Val Gln Gly Lys Ser Glu His Gly Leu Pro Leu Val Asp	
65 70 75 80	
AAT TGC AAA GAT TTG TCC TTA GCA GCT GGT AAC AAT TTC GAT GGA ACG	288
Asn Cys Lys Asp Leu Ser Leu Ala Ala Gly Asn Asn Phe Asp Gly Thr	
85 90 95	
GCT CCT TTG GAG TAT CAT CAG CAG TAT GAT CTG CAA CAA GAG TTT GAA	336
Ala Pro Leu Glu Tyr His Gln Gln Tyr Asp Leu Gln Gln Glu Phe Glu	
100 105 110	
CCA AAC TTC AAT GGT GGT TTC AAC AAT TGT CCC AGT TAT GGT GTA GTA	384
Pro Asn Phe Asn Gly Gly Phe Asn Asn Cys Pro Ser Tyr Gly Val Val	
115 120 125	
GAG GGT CCT ATA CAT ATC TCT AAT TTT ATC CCG ACT ATT TGT CCT CAC	432
Glu Gly Pro Ile His Ile Ser Asn Phe Ile Pro Thr Ile Cys Pro His	
130 135 140	
CCT CTG CAT TCT TGG GTC CAA AAA TGT GCT CTT TGG GAT TGC CCT AGC	480
Pro Leu His Ser Trp Val Gln Lys Cys Ala Leu Trp Asp Cys Pro Ser	
145 150 155 160	
CAG CTC AGG GAT TTG ATT GGG TCC AGG ATT ACT GCA GCA GCT TCC ACG	528
Gln Leu Arg Asp Leu Ile Gly Ser Arg Ile Thr Ala Ala Ala Ser Thr	
165 170 175	
CTG CAC TGG CTT TCA AAT GAA AGG GCC ACC AGG TAT GAA TCC GGT GTG	576
Leu His Trp Leu Ser Asn Glu Arg Ala Thr Arg Tyr Glu Ser Gly Val	
180 185 190	
CGT CCT GGA AGT ATC GGC CTA AAA GAC GGT CTG CTT TTT GCT GCT CTT	624
Arg Pro Gly Ser Ile Glu Leu Lys Asp Gly Leu Leu Phe Ala Ala Leu	
195 200 205	
AGT GCA AAG GCT GGA GGG AAA GAT GTT GGT ATT CCC GAA TGT GAA GGA	672
Ser Ala Lys Ala Gly Gly Lys Asp Val Gly Ile Pro Glu Cys Glu Gly	
210 215 220	
GCT GCA ACT GCT AAA TCT CCA TGG AAT GCT CCA GAG CTC TTT GAT CTC	720
Ala Ala Thr Ala Lys Ser Pro Trp Asn Ala Pro Glu Leu Phe Asp Leu	
225 230 235 240	
ACG GTT CTG GAG AGT GAG ACA CTA AGG GAG TGG CTA TTC TTT GAC AAG	768
Thr Val Leu Glu Ser Glu Thr Leu Arg Glu Trp Leu Phe Phe Asp Lys	
245 250 255	
CCA AGG AGG GCC TTT GAG AGC GGG AAC AGA AAG CAA AGA TCT TTA CCA	816
Pro Arg Arg Ala Phe Glu Ser Gly Asn Arg Lys Gln Arg Ser Leu Pro	
260 265 270	
GAC TAC AAT GGT CGT GGT TGG CAC GAG TCA CGT AAA CAG ATC ATG GTC	864
Asp Tyr Asn Gly Arg Gly Trp His Glu Ser Arg Lys Gln Ile Met Val	
275 280 285	



85

GAG TTT GGA GGG CTG AAG AGA TCT TAC TAC ATG GAT CCA CAG CCT CTG	912
Glu Phe Gly Gly Leu Lys Arg Ser Tyr Tyr Met Asp Pro Gln Pro Leu	
290 295 300	
CAC CAT TTC GAA TGG CAT CTT TAC GAA TAT GAG ATC AAC AAG TGT GAT	960
His His Phe Glu Trp His Leu Tyr Glu Tyr Glu Ile Asn Lys Cys Asp	
305 310 315 320	
GCT TGT GCC TTG TAC AGG CTC GAG CTC AAG CTT GTT GAC GGG AAG AAG	1008
Ala Cys Ala Leu Tyr Arg Leu Glu Leu Lys Leu Val Asp Gly Lys Lys	
325 330 335	
ACT TCA AAA GGC AAA GTC TCA AAC GAC TCA GTG GCT GAT CTG CAG AAG	1056
Thr Ser Lys Gly Lys Val Ser Asn Asp Ser Val Ala Asp Leu Gln Lys	
340 345 350	
CAG ATG GGA AGA CTC ACA GCT GAG TTC CCT CCA GAA AAC AAT ACC ACT	1104
Gln Met Gly Arg Leu Thr Ala Glu Phe Pro Pro Glu Asn Asn Thr Thr	
355 360 365	
AAC ACC ACC AAC AAC AAC AAA CGC TGC ATC AAA GGA AGA CCA AAA GTG	1152
Asn Thr Thr Asn Asn Asn Lys Arg Cys Ile Lys Gly Arg Pro Lys Val	
370 375 380	
AGC ACA AAA GTC GCC ACC GGG AAT GTT CAG AAC ACA GTA GAG CAG GCA	1200
Ser Thr Lys Val Ala Thr Gly Asn Val Gln Asn Thr Val Glu Gln Ala	
385 390 395 400	
AAT GAC TAT GGA GTA GGT GAA GAG TTT AAC TAT CTG GTC GGA AAT CTA	1248
Asn Asp Tyr Gly Val Gly Glu Glu Phe Asn Tyr Leu Val Gly Asn Leu	
405 410 415	
AGC GAC TAT TAT ATC CCC TG	1269
Ser Asp Tyr Tyr Ile Pro	
420	

## (2) INFORMATION FOR SEQ ID NO: 6:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 422 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

Asn Ser Ala Arg Gly Leu Leu Gln Leu His Pro Cys Asn Lys Val Val
1 5 10 15
Leu Trp Gly Leu Ser His Gln Ile Phe Val Gly Cys Cys Ser Ser Val
20 25 30

86

Met Glu Asp Asp Ala Thr Ser Lys Leu Ala Ala Pro Lys Pro Glu Pro  
 35 40 45

Ala Asp Gln Asn Leu Glu Ala Gly Lys Ala Ala Val Phe Gln Arg Gly  
 50 55 60

Tyr Asn Leu Val Gln Gly Lys Ser Glu His Gly Leu Pro Leu Val Asp  
 65 70 75 80

Asn Cys Lys Asp Leu Ser Leu Ala Ala Gly Asn Asn Phe Asp Gly Thr  
 85 90 95

Ala Pro Leu Glu Tyr His Gln Gln Tyr Asp Leu Gln Gln Glu Phe Glu  
 100 105 110

Pro Asn Phe Asn Gly Gly Phe Asn Asn Cys Pro Ser Tyr Gly Val Val  
 115 120 125

Glu Gly Pro Ile His Ile Ser Asn Phe Ile Pro Thr Ile Cys Pro His  
 130 135 140

Pro Leu His Ser Trp Val Gln Lys Cys Ala Leu Trp Asp Cys Pro Ser  
 145 150 155 160

Gln Leu Arg Asp Leu Ile Gly Ser Arg Ile Thr Ala Ala Ala Ser Thr  
 165 170 175

Leu His Trp Leu Ser Asn Glu Arg Ala Thr Arg Tyr Glu Ser Gly Val  
 180 185 190

Arg Pro Gly Ser Ile Gly Leu Lys Asp Gly Leu Leu Phe Ala Ala Leu  
 195 200 205

Ser Ala Lys Ala Gly Gly Lys Asp Val Gly Ile Pro Glu Cys Glu Gly  
 210 215 220

Ala Ala Thr Ala Lys Ser Pro Trp Asn Ala Pro Glu Leu Phe Asp Leu  
 225 230 235 240

Thr Val Leu Glu Ser Glu Thr Leu Arg Glu Trp Leu Phe Phe Asp Lys  
 245 250 255

Pro Arg Arg Ala Phe Glu Ser Gly Asn Arg Lys Gln Arg Ser Leu Pro  
 260 265 270

Asp Tyr Asn Gly Arg Gly Trp His Glu Ser Arg Lys Gln Ile Met Val  
 275 280 285

Glu Phe Gly Gly Leu Lys Arg Ser Tyr Tyr Met Asp Pro Gln Pro Leu  
 290 295 300

His His Phe Glu Trp His Leu Tyr Glu Tyr Glu Ile Asn Lys Cys Asp  
 305 310 315 320

Ala Cys Ala Leu Tyr Arg Leu Glu Leu Lys Leu Val Asp Gly Lys Lys

87

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          325              330              335
Thr Ser Lys Gly Lys Val Ser Asn Asp Ser Val Ala Asp Leu Gln Lys
      340              345              350
Gln Met Gly Arg Leu Thr Ala Glu Phe Pro Pro Glu Asn Asn Thr Thr
      355              360              365
Asn Thr Thr Asn Asn Asn Lys Arg Cys Ile Lys Gly Arg Pro Lys Val
      370              375              380
Ser Thr Lys Val Ala Thr Gly Asn Val Gln Asn Thr Val Glu Gln Ala
      385              390              395              400
Asn Asp Tyr Gly Val Gly Glu Glu Phe Asn Tyr Leu Val Gly Asn Leu
      405              410              415
Ser Asp Tyr Tyr Ile Pro
      420

```

## (2) INFORMATION FOR SEQ ID NO: 7:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 654 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA

## (iii) HYPOTHETICAL: NO

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..654

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

```

GAA TTC GGC ACG AGC TCC TTC CTC GGC TGT AAC AAG ATA GAG AAG AAG      48
Glu Phe Gly Thr Ser Ser Phe Leu Gly Cys Asn Lys Ile Glu Lys Lys
  1              5              10              15
ATG AAT ATG GAA GTG GAT ACA GTA ACA AGG AAG CCT CGT ATC TTA CTA      96
Met Asn Met Glu Val Asp Thr Val Thr Arg Lys Pro Arg Ile Leu Leu
      20              25              30
GCT GCA AGT GGA AGT GTG GCT TCA ATT AAG TTC AGT AAT CTC TGC CAT      144
Ala Ala Ser Gly Ser Val Ala Ser Ile Lys Phe Ser Asn Leu Cys His
      35              40              45
TGT TTC TCA GAA TGG GCT GAA GTC AAA GCC GTC GCT TCA AAA TCA TCT      192
Cys Phe Ser Glu Trp Ala Glu Val Lys Ala Val Ala Ser Lys Ser Ser

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88

50	55	60	
CTC AAT TTC GTT GAT AAA CCT TCT CTA CCT CAG AAT GTG ACT CTC TAT			240
Leu Asn Phe Val Asp Lys Pro Ser Leu Pro Gln Asn Val Thr Leu Tyr			
65	70	75	80
ACA GAT GAA GAT GAA TGG TCT AGC TGG AAC AAG ATT GGT GAT CCC GTT			288
Thr Asp Glu Asp Glu Trp Ser Ser Trp Asn Lys Ile Gly Asp Pro Val			
	85	90	95
CTT CAT ATC GAG CTC AGA CGC TGG GCT GAT GTT ATG ATC ATT GCT CCT			336
Leu His Ile Glu Leu Arg Arg Trp Ala Asp Val Met Ile Ile Ala Pro			
	100	105	110
TTG TCT GCT AAC ACA TTA GCC AAG ATT GCT GGT GGG TTA TGT GAT AAT			384
Leu Ser Ala Asn Thr Leu Ala Lys Ile Ala Gly Gly Leu Cys Asp Asn			
	115	120	125
CTA TTG ACA TGT ATA GTA AGA GCA TGG GAT TAT AGC AAA CCG TTG TTT			432
Leu Leu Thr Cys Ile Val Arg Ala Trp Asp Tyr Ser Lys Pro Leu Phe			
	130	135	140
GTT GCA CCG GCG ATG AAG ACT TTG ATG TGG AAC AAT CCT TTC ACA GAA			480
Val Ala Pro Ala Met Asn Thr Leu Met Trp Asn Asn Pro Phe Thr Glu			
	145	150	155
CGG CAC CTT GTC TTG CTT GAT GAA CTT GGA ATC ACC CTA ATT CCT CCG			528
Arg His Leu Val Leu Leu Asp Glu Leu Gly Ile Thr Leu Ile Pro Pro			
	165	170	175
ATC AAG AAG AAA CTG GCC TGT GGA GAG TAG GGT AAT GGC GCA ATG GCT			576
Ile Lys Lys Lys Leu Ala Cys Gly Asp Tyr Gly Asn Gly Ala Met Ala			
	180	185	190
GAG CCT TCT CTG ATT TAT TCC ACT GTT AGA CTG TTC TGG GAG TCA CAA			624
Glu Pro Ser Leu Ile Tyr Ser Thr Val Arg Leu Phe Trp Glu Ser Gln			
	195	200	205
GCT CGT AAA CAA AGA GAT GGA ACC AGT TG			654
Ala Arg Lys Gln Arg Asp Gly Thr Ser			
	210	215	

## (2) INFORMATION FOR SEQ ID NO: 8:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 217 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

Glu Phe Gly Thr Ser Ser Phe Leu Gly Cys Asn Lys Ile Glu Lys Lys

89

1	5	10	15
Met Asn Met Glu Val Asp Thr Val Thr Arg Lys Pro Arg Ile Leu Leu	20	25	30
Ala Ala Ser Gly Ser Val Ala Ser Ile Lys Phe Ser Asn Leu Cys His	35	40	45
Cys Phe Ser Glu Trp Ala Glu Val Lys Ala Val Ala Ser Lys Ser Ser	50	55	60
Leu Asn Phe Val Asp Lys Pro Ser Leu Pro Gln Asn Val Thr Leu Tyr	65	70	75
Thr Asp Glu Asp Glu Trp Ser Ser Trp Asn Lys Ile Gly Asp Pro Val	85	90	95
Leu His Ile Glu Leu Arg Arg Trp Ala Asp Val Met Ile Ile Ala Pro	100	105	110
Leu Ser Ala Asn Thr Leu Ala Lys Ile Ala Gly Gly Leu Cys Asp Asn	115	120	125
Leu Leu Thr Cys Ile Val Arg Ala Trp Asp Tyr Ser Lys Pro Leu Phe	130	135	140
Val Ala Pro Ala Met Asn Thr Leu Met Trp Asn Asn Pro Phe Thr Glu	145	150	155
Arg His Leu Val Leu Leu Asp Glu Leu Gly Ile Thr Leu Ile Pro Pro	165	170	175
Ile Lys Lys Lys Leu Ala Cys Gly Asp Tyr Gly Asn Gly Ala Met Ala	180	185	190
Glu Pro Ser Leu Ile Tyr Ser Thr Val Arg Leu Phe Trp Glu Ser Gln	195	200	205
Ala Arg Lys Gln Arg Asp Gly Thr Ser	210	215	

## (2) INFORMATION FOR SEQ ID NO: 9:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1856 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

## (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION:63..1583

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

GAATTCCTCG AGCTACGTCA GGGCCCTGAC GTAGCCGTCA ATCGAAATCC CAAAGATCAG	60
CG ATG GTG ACT CTA AAC GCT TCT TCT CCT CTC ACG ACC AAG TCG TTC	107
Met Val Thr Leu Asn Ala Ser Ser Pro Leu Thr Thr Lys Ser Phe	
1 5 10 15	
CTC CCC TAC CGT CAC GCT CCT CGC CGT CCA ATC TCT TTC TCC CCT GTC	155
Leu Pro Tyr Arg His Ala Pro Arg Arg Pro Ile Ser Phe Ser Pro Val	
20 25 30	
TTC GCC GTT CAT TCG ACT GAC CCC AAG AAA TCT ACC CAA TCA GCC TCC	203
Phe Ala Val His Ser Thr Asp Pro Lys Lys Ser Thr Gln Ser Ala Ser	
35 40 45	
GCT TCG GTT AAA TGG AGT CTA GAG AGT TGG AAG TCG AAG AAA GCT TTG	251
Ala Ser Val Lys Trp Ser Leu Glu Ser Trp Lys Ser Lys Lys Ala Leu	
50 55 60	
CAA TTG CCG GAT TAT CCT GAT CAG AAG GAT GTT GAT TCG GTT CTA CAG	299
Gln Leu Pro Asp Tyr Pro Asp Gln Lys Asp Val Asp Ser Val Leu Gln	
65 70 75	
ACG CTT TCT TCT TTT CCT CCT ATA GTT TTC GCT GGT GAG GCT AGG AAA	347
Thr Leu Ser Ser Phe Pro Pro Ile Val Phe Ala Gly Glu Ala Arg Lys	
80 85 90 95	
CTA GAG GAT AAG CTT GGT CAA GCG GCT ATG GGT CAA GCC TTT ATG CTT	395
Leu Glu Asp Lys Leu Gly Gln Ala Ala Met Gly Gln Ala Phe Met Leu	
100 105 110	
CAA GGT GGT GAT TGT GCT GAG AGT TTC AAG GAA TTT AAC GCT AAT AAC	443
Gln Gly Gly Asp Cys Ala Glu Ser Phe Lys Glu Phe Asn Ala Asn Asn	
115 120 125	
ATT AGA GAC ACC TTT AGG GTT CTT CTT CAG ATG GGT GTT GTT CTC ATG	491
Ile Arg Asp Thr Phe Arg Val Leu Leu Gln Met Gly Val Val Leu Met	
130 135 140	
TTC GGT GGC CAG TTA CCA GTT ATC AAG GTG GGA AGA ATG GCT GGT CAG	539
Phe Gly Gly Gln Leu Pro Val Ile Lys Val Gly Arg Met Ala Gly Gln	
145 150 155	
TTT GCG AAG CCG AGA TTA GAC CCA TTT GAG GAG AAA GAT GGT GTG AAG	587
Phe Ala Lys Pro Arg Leu Asp Pro Phe Glu Glu Lys Asp Gly Val Lys	
160 165 170 175	
CTG CCG AGT TAC AGA GGA GAT AAC ATA AAT GGT GAT GCT TTT GAT GAG	635

Leu Pro Ser Tyr Arg Gly Asp Asn Ile Asn Gly Asp Ala Phe Asp Glu	
180 185 190	
AAA TCG AGG ATT CCT GAT CCT CAT AGG ATG GTT AGA GCG TAC ACA CAG	683
Lys Ser Arg Ile Pro Asp Pro His Arg Met Val Arg Ala Tyr Thr Gln	
195 200 205	
TCT GTG GCT ACG TTG AAT CTC TTG AGA GCA TTT GCT ACT GGA GGT TAT	731
Ser Val Ala Thr Leu Asn Leu Leu Arg Ala Phe Ala Thr Gly Gly Tyr	
210 215 220	
GCA GCT ATG CAG AGA GTT AGC CAG TGG AAC CTT GAT TTC ACG CAA CAT	779
Ala Ala Met Gln Arg Val Ser Gln Trp Asn Leu Asp Phe Thr Gln His	
225 230 235	
AGT GAA CAG GGT GAC AGG TAC CGT GAA TTG GCT AAT AGA GTT GAT GAG	827
Ser Glu Gln Gly Asp Arg Tyr Arg Glu Leu Ala Asn Arg Val Asp Glu	
240 245 250 255	
GCT TTG GGA TTC ATG GGT GCA GCT GGA CTT ACT AGT GCT CAC CCG ATC	875
Ala Leu Gly Phe Met Gly Ala Ala Gly Leu Thr Ser Ala His Pro Ile	
260 265 270	
ATG ACT ACT ACT GAG TTT TGG ACA TCC CAT GAG TGT TTG TTA TTG CCT	923
Met Thr Thr Thr Glu Phe Trp Thr Ser His Glu Cys Leu Leu Leu Pro	
275 280 285	
TAT GAG CAA GCA CTC ACA AGA GAG GAT TCA ACA TCT GGA CTT TAC TAT	971
Tyr Glu Gln Ala Leu Thr Arg Glu Asp Ser Thr Ser Gly Leu Tyr Tyr	
290 295 300	
GAT TGC TCT GCG CAC ATG CTT TGG GTT GGA GAA CGA ACT CGC CAA CTT	1019
Asp Cys Ser Ala His Met Leu Trp Val Gly Glu Arg Thr Arg Gln Leu	
305 310 315	
GAT GGT GCT CAT GTT GAG TTT CTG AGG GGA ATC GCT AAC CCC CTC GGA	1067
Asp Gly Ala His Val Glu Phe Leu Arg Gly Ile Ala Asn Pro Leu Gly	
320 325 330 335	
ATC AAG GTG AGT GAT AAA ATG GTC CCT AGT GAA CTG GTT AAG CTG ATA	1115
Ile Lys Val Ser Asp Lys Met Val Pro Ser Glu Leu Val Lys Leu Ile	
340 345 350	
GAG ATA CTA AAC CCT CAG AAC AAG CCT GGA AGG ATT ACG GTT ATA GTG	1163
Glu Ile Leu Asn Pro Gln Asn Lys Pro Gly Arg Ile Thr Val Ile Val	
355 360 365	
AGA ATG GGA GCT GAG AAT ATG CGG GTC AAG CTT CCT AAT TTG ATC AGA	1211
Arg Met Gly Ala Glu Asn Met Arg Val Lys Leu Pro Asn Leu Ile Arg	
370 375 380	
GCA GTC CGT GGA GCC GGT CAG ATT GTG ACT TGG GTT AGT GAT CCA ATG	1259
Ala Val Arg Gly Ala Gly Gln Ile Val Thr Trp Val Ser Asp Pro Met	
385 390 395	

CAC GGA AAC ACA ATC ATG GCT CCT GGT GGG CTA AAA ACT CGT TCT TTC	1307
His Gly Asn Thr Ile Met Ala Pro Gly Gly Leu Lys Thr Arg Ser Phe	
400 405 410 415	
GAT GCA ATC AGG GCG GAA TTG AGA GCG TTC TTC GAC GTC CAT GAT CAA	1355
Asp Ala Ile Arg Ala Glu Leu Arg Ala Phe Phe Asp Val His Asp Gln	
420 425 430	
GAA GGG AGT TTC CCT GGC GGG GTT CAT TTA GAA ATG ACT GGT CAA AAC	1403
Glu Gly Ser Phe Pro Gly Gly Val His Leu Glu Met Thr Gly Gln Asn	
435 440 445	
GTG ACT GAA TGT GTC GGA GGG TCA CGC ACC ATC ACT TAC AAC GAT CTA	1451
Val Thr Glu Cys Val Gly Gly Ser Arg Thr Ile Thr Tyr Asn Asp Leu	
450 455 460	
AGC TCA CGC TAC CAC ACT CAC TGT GAC CCA AGA CTC AAC GCA TCT CAG	1499
Ser Ser Arg Tyr His Thr His Cys Asp Pro Arg Leu Asn Ala Ser Gln	
465 470 475	
TCT CTG GAG CTT GCA TTC ATC ATT GCA GAG CGT CTG CGA AAG AGA AGG	1547
Ser Leu Glu Leu Ala Phe Ile Ile Ala Glu Arg Leu Arg Lys Arg Arg	
480 485 490 495	
CTT GGT TCC GGG AAT CTT CCG TCA TCT ATT GGA GTC TAGAGAACAA	1593
Leu Gly Ser Gly Asn Leu Pro Ser Ser Ile Gly Val	
500 505	
GAAAATACTT ATCCGAGCTA GGATGTGTGT GTATAGAGGC TGATCTCTAG TTATTAAGTT	1653
GCCAAGTTAA ATGAGCTTGT GTACTGTAA AAGTAAGATA TTGTTGTTTT TGTGTGTTGG	1713
GTTATGATTT TGTCTGAAAT AAGTGGCTGA CTTTATAACC CGTAAATCTC TACGTCACGC	1773
TTGCAACAAA AATTCGATAT TTGATTCAAT CACAGAAAAG TCCTCCCATT AAGGTGTAAA	1833
CCCTGACGTA GCTCGAGGAA TTC	1856

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 507 amino acids  
(B) TYPE: amino acid  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

Met Val Thr Leu Asn Ala Ser Ser Pro Leu Thr Thr Lys Ser Phe Leu  
1 5 10 15  
Pro Tyr Arg His Ala Pro Arg Arg Pro Ile Ser Phe Ser Pro Val Phe  
20 25 30



93

Ala Val His Ser Thr Asp Pro Lys Lys Ser Thr Gln Ser Ala Ser Ala  
35 40 45

Ser Val Lys Trp Ser Leu Glu Ser Trp Lys Ser Lys Lys Ala Leu Gln  
50 55 60

Leu Pro Asp Tyr Pro Asp Gln Lys Asp Val Asp Ser Val Leu Gln Thr  
65 70 75 80

Leu Ser Ser Phe Pro Pro Ile Val Phe Ala Gly Glu Ala Arg Lys Leu  
85 90 95

Glu Asp Lys Leu Gly Gln Ala Ala Met Gly Gln Ala Phe Met Leu Gln  
100 105 110

Gly Gly Asp Cys Ala Glu Ser Phe Lys Glu Phe Asn Ala Asn Asn Ile  
115 120 125

Arg Asp Thr Phe Arg Val Leu Leu Gln Met Gly Val Val Leu Met Phe  
130 135 140

Gly Gly Gln Leu Pro Val Ile Lys Val Gly Arg Met Ala Gly Gln Phe  
145 150 155 160

Ala Lys Pro Arg Leu Asp Pro Phe Glu Glu Lys Asp Gly Val Lys Leu  
165 170 175

Pro Ser Tyr Arg Gly Asp Asn Ile Asn Gly Asp Ala Phe Asp Glu Lys  
180 185 190

Ser Arg Ile Pro Asp Pro His Arg Met Val Arg Ala Tyr Thr Gln Ser  
195 200 205

Val Ala Thr Leu Asn Leu Leu Arg Ala Phe Ala Thr Gly Gly Tyr Ala  
210 215 220

Ala Met Gln Arg Val Ser Gln Trp Asn Leu Asp Phe Thr Gln His Ser  
225 230 235 240

Glu Gln Gly Asp Arg Tyr Arg Glu Leu Ala Asn Arg Val Asp Glu Ala  
245 250 255

Leu Gly Phe Met Gly Ala Ala Gly Leu Thr Ser Ala His Pro Ile Met  
260 265 270

Thr Thr Thr Glu Phe Trp Thr Ser His Glu Cys Leu Leu Leu Pro Tyr  
275 280 285

Glu Gln Ala Leu Thr Arg Glu Asp Ser Thr Ser Gly Leu Tyr Tyr Asp  
290 295 300

Cys Ser Ala His Met Leu Trp Val Gly Glu Arg Thr Arg Gln Leu Asp  
305 310 315 320

Gly Ala His Val Glu Phe Leu Arg Gly Ile Ala Asn Pro Leu Gly Ile

94

325	330	335
Lys Val Ser Asp Lys Met Val Pro Ser Glu Leu Val Lys Leu Ile Glu		
340	345	350
Ile Leu Asn Pro Gln Asn Lys Pro Gly Arg Ile Thr Val Ile Val Arg		
355	360	365
Met Gly Ala Glu Asn Met Arg Val Lys Leu Pro Asn Leu Ile Arg Ala		
370	375	380
Val Arg Gly Ala Gly Gln Ile Val Thr Trp Val Ser Asp Pro Met His		
385	390	395
Gly Asn Thr Ile Met Ala Pro Gly Gly Leu Lys Thr Arg Ser Phe Asp		
405	410	415
Ala Ile Arg Ala Glu Leu Arg Ala Phe Phe Asp Val His Asp Gln Glu		
420	425	430
Gly Ser Phe Pro Gly Gly Val His Leu Glu Met Thr Gly Gln Asn Val		
435	440	445
Thr Glu Cys Val Gly Gly Ser Arg Thr Ile Thr Tyr Asn Asp Leu Ser		
450	455	460
Ser Arg Tyr His Thr His Cys Asp Pro Arg Leu Asn Ala Ser Gln Ser		
465	470	475
Leu Glu Leu Ala Phe Ile Ile Ala Glu Arg Leu Arg Lys Arg Arg Leu		
485	490	495
Gly Ser Gly Asn Leu Pro Ser Ser Ile Gly Val		
500	505	

## (2) INFORMATION FOR SEQ ID NO: 11:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1081 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA

## (iii) HYPOTHETICAL: NO

## (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..954

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

GAA TTC GGC ACG AGG GAT CCC AAG AAC CTA AAT CGT CAC CAA GTA CCA Glu Phe Gly Thr Arg Asp Pro Lys Asn Leu Asn Arg His Gln Val Pro 510 515 520	48
AAT TTC TTG AAC CCA CCA CCA CCA CCG CGA AAT CAG GGT TTG GTA GAT Asn Phe Leu Asn Pro Pro Pro Pro Pro Arg Asn Gln Gly Leu Val Asp 525 530 535	96
GAT GAT GCT GCT TCT GCT GTT GTT TCC GAC GAG AAT CGC AAA CCA ACA Asp Asp Ala Ala Ser Ala Val Val Ser Asp Glu Asn Arg Lys Pro Thr 540 545 550 555	144
ACT GAG ATT AAA GAT TTC CAG ATC GTG GTC TCT GCT TCC GAC AAA GAA Thr Glu Ile Lys Asp Phe Gln Ile Val Val Ser Ala Ser Asp Lys Glu 560 565 570	192
CCA AAC AAG AAG AGT CAG AAT CAG AAC CAG CTT GGT CCT AAG AGA AGC Pro Asn Lys Lys Ser Gln Asn Gln Asn Gln Leu Gly Pro Lys Arg Ser 575 580 585	240
TCT AAC AAA GAC AGA CAC ACT AAA GTC GAA GGT AGA GGT CGA CGA ATT Ser Asn Lys Asp Arg His Thr Lys Val Glu Gly Arg Gly Arg Arg Ile 590 595 600	288
CGG ATG CCT GCT CTT TGT GCT GCT AGG ATT TTT CAA TTG ACT AGA GAA Arg Met Pro Ala Leu Cys Ala Ala Arg Ile Phe Gln Leu Thr Arg Glu 605 610 615	336
TTG GGT CAT AAA TCT GAT GGT GAA ACT ATC CAG TGG CTG CTT CAA CAA Leu Gly His Lys Ser Asp Gly Glu Thr Ile Gln Trp Leu Leu Gln Gln 620 625 630 635	384
GCT GAG CCA TCG ATT ATT GCA GCT ACT GGT TCA GGA ACT ATA CCG GCC Ala Glu Pro Ser Ile Ile Ala Ala Thr Gly Ser Gly Thr Ile Pro Ala 640 645 650	432
TCT GCT TTA GCT TCT TCA GCT GCA ACC TCT AAC CAT CAT CAA GGT GGG Ser Ala Leu Ala Ser Ser Ala Ala Thr Ser Asn His His Gln Gly Gly 655 660 665	480
TCT CTT ACT GCT GGT TTA ATG ATC AGT CAT GAC TTA GAT GGT GGG TCT Ser Leu Thr Ala Gly Leu Met Ile Ser His Asp Leu Asp Gly Gly Ser 670 675 680	528
AGT AGT AGT GGT AGA CCA TTA AAT TGG GGG ATT GGT GGC GGT GAA GGA Ser Ser Ser Gly Arg Pro Leu Asn Trp Gly Ile Gly Gly Gly Glu Gly 685 690 695	576
GTT TCT AGG TCA AGT TTA CCA ACT GGG TTA TGG CCA AAT GTA GCT GGG Val Ser Arg Ser Ser Leu Pro Thr Gly Leu Trp Pro Asn Val Ala Gly 700 705 710 715	624
TTT GGT TCT GGT GTG CCA ACC ACT GGT TTA ATG AGT GAA GGA GCT GGT Phe Gly Ser Gly Val Pro Thr Thr Gly Leu Met Ser Glu Gly Ala Gly 720 725 730	672

96

TAT AGA ATT GGG TTT CCT GGT TTT GAT TTT CCT GGT GTT GGT CAT ATG 720  
 Tyr Arg Ile Gly Phe Pro Gly Phe Asp Phe Pro Gly Val Gly His Met  
 735 740 745

AGT TTT GCA TCT ATT TTG GGT GGG AAT CAT AAT CAG ATG CCT GGA CTT 768  
 Ser Phe Ala Ser Ile Leu Gly Gly Asn His Asn Gln Met Pro Gly Leu  
 750 755 760

GAG TTA GGC TTG TCT CAA GAA GGG AAT GTT GGT GTT TTG AAT CCT CAG 816  
 Glu Leu Gly Leu Ser Gln Glu Gly Asn Val Gly Val Leu Asn Pro Gln  
 765 770 775

TCT TTT ACT CAG ATT TAT CAA CAG ATG GGT CAG GCT CAG GCT CAA GCT 864  
 Ser Phe Thr Gln Ile Tyr Gln Gln Met Gly Gln Ala Gln Ala Gln Ala  
 780 785 790 795

CAA GGT AGG GTT CTT CAC CAT ATG CAT CAT AAC CAT GAA GAA CAT CAG 912  
 Gln Gly Arg Val Leu His His Met His His Asn His Glu Glu His Gln  
 800 805 810

CAA GAG AGT GGT GAG AAA GAT GAT TCT CAA GGC TCA GGT CGT 954  
 Gln Glu Ser Gly Glu Lys Asp Asp Ser Gln Gly Ser Gly Arg  
 815 820 825

TAAAAGGATT GGGTTTTTTT TGTATCTTCT GGATTGAAA AAGCTTTTGG CTTTGTGTTT 1014

GTGATAATAT TGTTGTAAT TGTACACCA TGCAGAAGAA AAAGAAAAGG TTATATAAAA 1074

AAAAAAA 1081

## (2) INFORMATION FOR SEQ ID NO: 12:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 318 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

Glu Phe Gly Thr Arg Asp Pro Lys Asn Leu Asn Arg His Gln Val Pro  
 1 5 10 15

Asn Phe Leu Asn Pro Pro Pro Pro Pro Arg Asn Gln Gly Leu Val Asp  
 20 25 30

Asp Asp Ala Ala Ser Ala Val Val Ser Asp Glu Asn Arg Lys Pro Thr  
 35 40 45

Thr Glu Ile Lys Asp Phe Gln Ile Val Val Ser Ala Ser Asp Lys Glu  
 50 55 60

Pro Asn Lys Lys Ser Gln Asn Gln Asn Gln Leu Gly Pro Lys Arg Ser

97

65	70	75	80
Ser Asn Lys Asp Arg His Thr Lys Val Glu Gly Arg Gly Arg Arg Ile	85	90	95
Arg Met Pro Ala Leu Cys Ala Ala Arg Ile Phe Gln Leu Thr Arg Glu	100	105	110
Leu Gly His Lys Ser Asp Gly Glu Thr Ile Gln Trp Leu Leu Gln Gln	115	120	125
Ala Glu Pro Ser Ile Ile Ala Ala Thr Gly Ser Gly Thr Ile Pro Ala	130	135	140
Ser Ala Leu Ala Ser Ser Ala Ala Thr Ser Asn His His Gln Gly Gly	145	150	155
Ser Leu Thr Ala Gly Leu Met Ile Ser His Asp Leu Asp Gly Gly Ser	165	170	175
Ser Ser Ser Gly Arg Pro Leu Asn Trp Gly Ile Gly Gly Gly Glu Gly	180	185	190
Val Ser Arg Ser Ser Leu Pro Thr Gly Leu Trp Pro Asn Val Ala Gly	195	200	205
Phe Gly Ser Gly Val Pro Thr Thr Gly Leu Met Ser Glu Gly Ala Gly	210	215	220
Tyr Arg Ile Gly Phe Pro Gly Phe Asp Phe Pro Gly Val Gly His Met	225	230	235
Ser Phe Ala Ser Ile Leu Gly Gly Asn His Asn Gln Met Pro Gly Leu	245	250	255
Glu Leu Gly Leu Ser Gln Glu Gly Asn Val Gly Val Leu Asn Pro Gln	260	265	270
Ser Phe Thr Gln Ile Tyr Gln Gln Met Gly Gln Ala Gln Ala Gln Ala	275	280	285
Gln Gly Arg Val Leu His His Met His His Asn His Glu Glu His Gln	290	295	300
Gln Glu Ser Gly Glu Lys Asp Asp Ser Gln Gly Ser Gly Arg	305	310	315

(2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 777 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

98

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION:1..774

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

AAT TCG GCA CGA GGC AAC AAA AAG AGG AGA CTT CCT GTA GAT GAG CAG	48
Asn Ser Ala Arg Gly Asn Lys Lys Arg Arg Leu Pro Val Asp Glu Gln	
320 325 330	
GAG AAT CGT GGT GAC AA GTG GCT AAT GGT CTT AAC CGC CAG ATT GTT	96
Glu Asn Arg Gly Asp Asn Val Ala Asn Gly Leu Asn Arg Gln Ile Val	
335 340 345 350	
AGA TAT CAG CCG TCG ATA AAC GAA GCA GCA CAA AAT ATG CTT CGA CAG	144
Arg Tyr Gln Pro Ser Ile Asn Glu Ala Ala Gln Asn Met Leu Arg Gln	
355 360 365	
TTC TTA AAT ACT AGT ACC TCA CCT CGG TAT GAA TCA GTT TCA AAC AAT	192
Phe Leu Asn Thr Ser Thr Ser Pro Arg Tyr Glu Ser Val Ser Asn Asn	
370 375 380	
CCT GAC AGT TTC CTA TTG GGT GAT GTT CCC AGT TCT ACC TCT GTA GAC	240
Pro Asp Ser Phe Leu Leu Gly Asp Val Pro Ser Ser Thr Ser Val Asp	
385 390 395	
AAT GGG AAC CCT TCA AGT AGA GTT TCT GGA GTA ACA TTG GCC GAG TTT	288
Asn Gly Asn Pro Ser Ser Arg Val Ser Gly Val Thr Leu Ala Glu Phe	
400 405 410	
TCA CCC AAC ACA GTT CAG TCA GCA ACG AAT CAA GTA CCC GAA GCA AGT	336
Ser Pro Asn Thr Val Gln Ser Ala Thr Asn Gln Val Pro Glu Ala Ser	
415 420 425 430	
TTG GCT CAT CAT CCT CAA GCT GGT CTG GTT CAG CCA AAT ATA GGT CAA	384
Leu Ala His His Pro Gln Ala Gly Leu Val Gln Pro Asn Ile Gly Gln	
435 440 445	
AGT CCG GCT CAA GGA GCA GCA CCT GCA GAC TCT TGG AGC CCT GAA TTT	432
Ser Pro Ala Gln Gly Ala Ala Pro Ala Asp Ser Trp Ser Pro Glu Phe	
450 455 460	
GAT TTA GTT GGA TGC GAG ACA GAT AGT GGA GAG TGT TTT GAT CCA ATA	480
Asp Leu Val Gly Cys Glu Thr Asp Ser Gly Glu Cys Phe Asp Pro Ile	
465 470 475	
ATG GCT GTT TTA GAT GAG TCA GAA GGC GAT GCA ATT TCT CCT GAA GGT	528
Met Ala Val Leu Asp Glu Ser Glu Gly Asp Ala Ile Ser Pro Glu Gly	
480 485 490	

99

GAG GGC AAG ATG AAT GAG TTA CTG GAG GGA GTC CCT AAG CTG CCC GGA	576
Glu Gly Lys Met Asn Glu Leu Leu Glu Gly Val Pro Lys Leu Pro Gly	
495 500 505 510	
ATC CAA GAT CCA TTC TGG GAA CAG TTC TTT TCT GTT GAA CTC CCA GCG	624
Ile Gln Asp Pro Phe Trp Glu Gln Phe Phe Ser Val Glu Leu Pro Ala	
515 520 525	
ATT GCA GAT ACA GAC GAT ATT CTA TCA GGA TCA GTG GAG AAT AAT GAC	672
Ile Ala Asp Thr Asp Asp Ile Leu Ser Gly Ser Val Glu Asn Asn Asp	
530 535 540	
TTG GTA TTG GAA CAA GAA CCA AAC GAG TGG ACC CGT AAT GAA CAA CAA	720
Leu Val Leu Glu Gln Glu Pro Asn Glu Trp Thr Arg Asn Glu Gln Gln	
545 550 555	
ATG AAG TAT CTT ACT GAA CAA ATG GGA CTG CTT TCC TCA GAA GCA CAG	768
Met Lys Tyr Leu Thr Glu Gln Met Gly Leu Leu Ser Ser Glu Ala Gln	
560 565 570	
AGG AAA TAA	777
Arg Lys	
575	

## (2) INFORMATION FOR SEQ ID NO: 14:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 258 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

Asn Ser Ala Arg Gly Asn Lys Lys Arg Arg Leu Pro Val Asp Glu Gln	
1 5 10 15	
Glu Asn Arg Gly Asp Asn Val Ala Asn Gly Leu Asn Arg Gln Ile Val	
20 25 30	
Arg Tyr Gln Pro Ser Ile Asn Glu Ala Ala Gln Asn Met Leu Arg Gln	
35 40 45	
Phe Leu Asn Thr Ser Thr Ser Pro Arg Tyr Glu Ser Val Ser Asn Asn	
50 55 60	
Pro Asp Ser Phe Leu Leu Gly Asp Val Pro Ser Ser Thr Ser Val Asp	
65 70 75 80	
Asn Gly Asn Pro Ser Ser Arg Val Ser Gly Val Thr Leu Ala Glu Phe	
85 90 95	
Ser Pro Asn Thr Val Gln Ser Ala Thr Asn Gln Val Pro Glu Ala Ser	

100

100	105	110
Leu Ala His His Pro Gln Ala Gly Leu Val Gln Pro Asn Ile Gly Gln		
115	120	125
Ser Pro Ala Gln Gly Ala Ala Pro Ala Asp Ser Trp Ser Pro Glu Phe		
130	135	140
Asp Leu Val Gly Cys Glu Thr Asp Ser Gly Glu Cys Phe Asp Pro Ile		
145	150	155
Met Ala Val Leu Asp Glu Ser Glu Gly Asp Ala Ile Ser Pro Glu Gly		
165	170	175
Glu Gly Lys Met Asn Glu Leu Leu Glu Gly Val Pro Lys Leu Pro Gly		
180	185	190
Ile Gln Asp Pro Phe Trp Glu Gln Phe Phe Ser Val Glu Leu Pro Ala		
195	200	205
Ile Ala Asp Thr Asp Asp Ile Leu Ser Gly Ser Val Glu Asn Asn Asp		
210	215	220
Leu Val Leu Glu Gln Glu Pro Asn Glu Trp Thr Arg Asn Glu Gln Gln		
225	230	235
Met Lys Tyr Leu Thr Glu Gln Met Gly Leu Leu Ser Ser Glu Ala Gln		
245	250	255

Arg Lys

## (2) INFORMATION FOR SEQ ID NO: 15:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "oligonucleotide"

## (iii) HYPOTHETICAL: YES~

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

CGGATCCGAA TTCATGGAGA ACGAG~

25

## (2) INFORMATION FOR SEQ ID NO: 16:

## (i) SEQUENCE CHARACTERISTICS:



101

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
  - (A) DESCRIPTION: /desc = "oligonucleotide"

- (iii) HYPOTHETICAL: YES

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

CGGATCCGAA TTCTCAGAAC TGAGA

25

- (2) INFORMATION FOR SEQ ID NO: 17:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 21 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
  - (A) DESCRIPTION: /desc = "oligonucleotide"

- (iii) HYPOTHETICAL: YES

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

AGGGATGTTT AATACCACTA C

21

- (2) INFORMATION FOR SEQ ID NO: 18:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 21 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
  - (A) DESCRIPTION: /desc = "oligonucleotide"

- (iii) HYPOTHETICAL: YES

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

GCACAGTTGA AGTGAACCTG C

21

102

## (2) INFORMATION FOR SEQ ID NO: 19:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "oligonucleotide"

## (iii) HYPOTHETICAL: YES

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

CGAGATCTGA ATTCATGGAT CAGTA

25

## (2) INFORMATION FOR SEQ ID NO: 20:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "oligonucleotide"

## (iii) HYPOTHETICAL: YES

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

CGAGATCTGA ATTCCTAAGG CATGCC

26

### Claims

1. A DNA sequence encoding a cell cycle interacting protein or encoding an immunologically active and/or functional fragment of such a protein, selected from the group consisting of:
  - (a) DNA sequences
    - (aa) comprising a nucleotide sequence encoding at least the mature form of a protein (LDV115) comprising the amino acid sequence as given in SEQ ID NO: 2;
    - (ab) comprising the nucleotide sequence as given in SEQ ID NO: 1;
    - (ac) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (aa) or (ab) under stringent hybridization conditions;
    - (ad) comprising an nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (aa) or (ab);
    - (ae) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (aa) to (ad);
  - (b) DNA sequences
    - (ba) comprising a nucleotide sequence encoding at least the mature form of a protein (LDV24) comprising the amino acid sequence as given in SEQ ID NO: 4;
    - (bb) comprising the nucleotide sequence as given in SEQ ID NO: 3;
    - (bc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (ba) or (bb) under stringent hybridization conditions;
    - (bd) comprising an nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (ba) or (bb);

- (be) comprising a nucleotide sequence encoding at least the cyclin-like interacting domain of the protein encoded by the nucleotide sequence of any one of (ba) to (bd);
- (c) DNA sequences
  - (ca) comprising a nucleotide sequence encoding at least the mature form of a protein (VB33) comprising the amino acid sequence as given in SEQ ID NO: 6;
  - (cb) comprising the nucleotide sequence as given in SEQ ID NO: 5;
  - (cc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (ca) or (cb) under stringent hybridization conditions;
  - (cd) comprising an nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (ca) or (cb);
  - (ce) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (ca) to (cd);
- (d) DNA sequences
  - (da) comprising a nucleotide sequence encoding at least the mature form of a protein (VB89) comprising the amino acid sequence as given in SEQ ID NO: 8;
  - (db) comprising the nucleotide sequence as given in SEQ ID NO: 7;
  - (dc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (da) or (db) under stringent hybridization conditions;
  - (dd) comprising an nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (da) or (db);
  - (de) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (da) to (dd);

(e) DNA sequences

- (ea) comprising a nucleotide sequence encoding at least the mature form of a protein (VBDAHP) comprising the amino acid sequence as given in SEQ ID NO: 10;
- (eb) comprising the nucleotide sequence as given in SEQ ID NO: 9;
- (ec) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (ea) or (eb) under stringent hybridization conditions;
- (ed) comprising a nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (ea) or (eb);
- (ee) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (ea) to (ed);

(f) DNA sequences

- (fa) comprising a nucleotide sequence encoding at least the mature form of a protein (VBDPBP) comprising the amino acid sequence as given in SEQ ID NO: 12;
- (fb) comprising the nucleotide sequence as given in SEQ ID NO: 11;
- (fc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (fa) or (fb) under stringent hybridization conditions;
- (fd) comprising a nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (fa) or (fb);
- (fe) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (fa) to (fd);

(g) DNA sequences

- (ga) comprising a nucleotide sequence encoding at least the mature form of a protein (VBHSF) comprising the amino acid sequence as given in SEQ ID NO: 14;
  - (gb) comprising the nucleotide sequence as given in SEQ ID NO: 13;
  - (gc) comprising a nucleotide sequence hybridizing with the complementary strand of a nucleotide sequence as defined in (ga) or (gb) under stringent hybridization conditions;
  - (gd) comprising an nucleotide sequence encoding a protein having an amino acid sequence at least 60 % identical to the amino acid sequence encoded by the nucleotide sequence of (ga) or (gb);
  - (ge) comprising a nucleotide sequence encoding at least the domain binding to CDKs of the protein encoded by the nucleotide sequence of any one of (ga) to (gd);
  - (h) DNA sequences, the nucleotide sequence of which is degenerate as a result of the genetic code to a nucleotide sequence of a DNA sequence as defined in any one of (a) to (g); and
  - (i) DNA sequences comprising a nucleotide sequence encoding a fragment of a protein encoded by a DNA sequence of any one of (a) to (h), wherein said fragment is capable of interacting with a cell cycle protein.
2. A method for identifying and obtaining cell cycle interacting proteins comprising a two-hybrid screening assay wherein CDC2a or CDC2b as a bait and a cDNA library of a plant cell suspension as prey are used.
  3. The method of claim 2, wherein said CDC2a is CDC2aAt and CDC2b is CDC2bAt.
  4. A DNA sequence encoding a cell cycle interacting protein obtainable by the method of claim 2 or 3.

5. A nucleic acid molecule of at least 15 nucleotides in length hybridizing specifically with a DNA sequence of claim 1 or 4 or with a complementary strand thereof.
6. A vector comprising a DNA sequence of claim 1 or 4.
7. The vector of claim 6 which is an expression vector wherein the DNA sequence is operatively linked to one or more control sequences allowing the expression in prokaryotic and/or eukaryotic host cells.
8. A host cell containing a vector of claim 6 or 7 or a DNA sequence of claim 1 or 4.
9. The host cell of claim 8 which is a bacterial, insect, fungal, plant or animal cell.
10. A method for the production of a cell cycle interacting protein or an immunologically active or functional fragment thereof comprising culturing a host cell of claim 8 or 9 under conditions allowing the expression of the protein and recovering the produced protein from the culture.
11. A cell cycle interacting protein or an immunologically active or functional fragment thereof encodable by a DNA sequence of claim 1 or 4 or obtainable by the method of claim 2; 3 or 10.
12. An antibody specifically recognizing the protein of claim 11 or a fragment or epitope thereof.
13. A method for the production of transgenic plants, plant cells or plant tissue comprising the introduction of a DNA sequence of claim 1, 4 or 5 or a vector of claim 6 or 7 into the genome of said plant, plant cell or plant tissue.
14. The method of claim 13 further comprising regenerating a plant from said plant tissue or plant cell.

15. A transgenic plant cell comprising a DNA sequence of claim 1 or 4 which is operably linked to regulatory elements allowing transcription and/or expression of the DNA sequence in plant cells or obtainable according to the method of claim 13 or 14.
16. The transgenic plant cell of claim 15 wherein said DNA sequence or said vector is stably integrated into the genome of the plant cell.
17. A transgenic plant or a plant tissue comprising plant cells of claim 15 or 16.
18. The transgenic plant of claim 17 in which plant cell division and/or growth is enhanced and/or wherein the plant is less sensitive to environmental stress compared to the corresponding wildtype plant.
19. A transgenic plant cell which contains stably integrated into the genome a DNA sequence of claim 1, 4 or 5 or part thereof or obtainable according to the method of claim 13 or 14, wherein the transcription and/or expression of the DNA sequence or part thereof leads to reduction of the synthesis of a cell cycle interacting protein in the cells.
20. The plant cell of claim 19, wherein the reduction is achieved by an antisense, sense, ribozyme, co-suppression, dominant mutant effect and/or a knock out mutatin in the gene.
21. A transgenic plant or plant tissue comprising plant cells of claim 19 or 20.
22. The transgenic plant of claim 21 which displays a deficiency in plant cell division and/or growth.



23. Harvestable parts or propagation material of plants of any one of claims 17, 18, 21 or 22 comprising plant cells of claim 15, 16, 19 or 20.
24. A regulatory sequence of a promoter regulating the expression of a nucleic acid molecule comprising the DNA sequence of any one of claim 1 or 4, said regulatory sequence being capable of conferring expression of a heterologous DNA sequence during various stages of the cell cycle.
25. A recombinant DNA molecule comprising the regulatory sequence of claim 24.
26. The recombinant DNA molecule of claim 25, wherein said regulatory sequence is operatively linked to a heterologous DNA sequence.
27. A host cell transformed with a regulatory sequence of claim 24 or a recombinant DNA molecule of claim 25 or 26.
28. A transgenic plant, plant tissue, or plant cell comprising the regulatory sequence of claim 24 or the recombinant DNA molecule of claim 25 or 26.
29. A method for the identification of an activator or inhibitor of cell cycle interacting proteins or their encoding genes comprising the steps of:
  - (a) culturing a plant cell or tissue or maintaining a plant comprising a recombinant DNA molecule comprising a readout system operatively linked to a regulatory sequence of claim 24 in the presence of a compound or a sample comprising a plurality of compounds under conditions which permit expression of said readout system;
  - (b) identifying or verifying a sample and compound, respectively, which leads to suppression or activation and/or enhancement of expression of said readout system in said plant, plant cell, or plant tissue.

30. A method for identifying and obtaining an activator or inhibitor of cell division comprising the steps of:
- (a) combining a compound to be screened with a reaction mixture containing the cell cycle interacting protein of claim 11 and a readout system capable of interacting with the protein under suitable conditions which permit interaction of the protein with said readout system;
  - (b) identifying or verifying a sample and compound, respectively, which leads to suppression or activation of the readout system.
31. A method of producing a therapeutic agent comprising the steps of the method of claim 30 and synthesizing the activator or inhibitor obtained or identified in step (b) or an analog or derivative thereof in an amount sufficient to provide said agent in a therapeutically effective amount to a patient.
32. A method of producing a plant effective agent comprising the steps of the method of claim 30 and synthesizing the activator or inhibitor obtained or identified in step (b) or an analog or derivative thereof in an effective amount sufficient to provide said agent in an effective amount suitable for the application in agriculture or plant cell and tissue culture.
33. A method of producing a therapeutic or plant effective composition comprising the steps of the method of claim 30 and combining the compound obtained or identified in step (b) or an analog or derivative thereof with a pharmaceutically acceptable carrier or with a plant cell and tissue culture acceptable carrier.
34. An activator or inhibitor of a cell division obtained by the method of any one of claims 30 to 32.

35. A composition comprising a DNA sequence of claim 1, 4 or 5, a vector of claim 6 or 7, a protein of claim 11, an antibody of claim 12, or the activator or inhibitor of claim 34.
36. The composition of claim 35 for use as a medicament, a diagnostic means, a kit or plant effective agent.
37. Use of a DNA sequence of claim 1, 4 or 5, the vector of claim 6 or 7, the protein of claim 11, the antibody of claim 12 or the activator or inhibitor of claim 34 for modulating the cell cycle in an animal or plant, plant cell division and/or growth, for influencing the activity of cell cycle proteins in a plant or animal cell, as positive or negative regulator of cell proliferation, for modifying the growth inhibition caused by environmental stress conditions, for use in a screening method for the identification of inhibitors or activators of cell cycle proteins, as growth regulator, herbicide or for inducing nematode resistance in plants.
38. Use of a DNA sequence of claim 1, 4 or 5 or the regulatory sequence of claim 24 as a marker gene in plant or animal cell and tissue culture or as a marker in marker-assisted plant breeding.
39. Use of the two-hybrid system as defined in claim 2 or 3 for the identification of cell cycle interacting proteins or activators or inhibitors of such proteins.
40. Use of a regulatory sequence of claim 24 or a recombinant DNA molecule of claim 25 or 26, for the expression of a heterologous DNA sequence during a stage of the cell cycle.

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### Abstract

Provided are DNA sequences encoding cell cycle interacting proteins as well as methods for obtaining the same. Furthermore, vectors comprising said DNA sequences are described, wherein the DNA sequences are operatively linked to regulatory elements allowing expression in prokaryotic and/or eukaryotic host cells. In addition, proteins encoded by said DNA sequences, antibodies to said proteins and methods for their production are provided. Also described is a method for controlling or altering growth characteristics of a plant and/or a plant cell comprising introduction and/or expression of one or more cell cycle regulatory proteins functional in a plant or parts thereof and/or one or more DNA sequences encoding such proteins. Also provided are regulatory sequences controlling the expression of the above described cell cycle interacting proteins. Methods for the identification of compounds being capable of activating or inhibiting the cell cycle are described as well. Further described are diagnostic compositions comprising the aforementioned DNA sequences, regulatory sequences, proteins, antibodies, inhibitors and activators. Furthermore, transgenic plant cells, plant tissue and plants containing the above-described DNA sequences and vectors are described as well as the use of the aforementioned DNA sequences, vectors, proteins, regulatory sequences, antibodies and/or compounds identified by the method of the invention in plant cell and tissue culture, plant breeding and/or agriculture.

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